

# Ultrasound elastography: a brief clinical history of an evolving technique



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## Keywords

ultrasound, elastography, history

received 19.03.2024

accepted 21.07.2024

published online 2024

## Bibliography

Ultrasound Int Open 2024; 10: a23786926

DOI 10.1055/a-2378-6926

ISSN 2509-596X

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## ABSTRACT

The history of the emerging elastographic technique is presented. Ultrasound imaging of elasticity and tissue strain has gained clinical acceptance as an established technique useful in routine daily clinical practice.

## Introduction

Where are we coming from and where are we going [1, 2]? Developments in ultrasound imaging as we know them today are not self-evident but have undergone a continued and sustained evolution. WFUMB has a website showing the history of ultrasound worldwide [<https://wfumb.info/pdfhistory-2/>]. In addition, EFSUMB has prepared an online e-book on the history of ultrasound, which displays the history of its members [<https://efsumb.org/history/>] and is a work in progress. The “History of ultrasound in medicine from its birth to date on the occasion of the 50-year anniversary of EFSUMB” [3] and the “History of ultrasound in obstetrics and gynecology from 1971 to 2021 on the occasion of the 50-year anniversary of EFSUMB” [4] have been published. An editorial and

booklet also summarize the birth and 50 years of EFSUMB [5, 6]. In addition, the history of contrast-enhanced ultrasound has been summarized [7]. Highlights of ultrasound over the past 70 years were summarized by Nielsen et al. [8]. The history of student ultrasound education (SUSE) is also being summarized in an ongoing study. The current article focuses on the history and development of elastography in ultrasound and how it became a major imaging modality in medical diagnosis. What followed the more than 2000 years of manual palpation?

Manual palpation during physical examination has been used in clinical practice for many centuries to distinguish harder areas from softer ones in the skin, organs, and/or tissues and to detect tumors. Palpation reflects various characteristics of a certain structure

including the position but also other features, including hardness, mobility, and pulsation. Palpation is subjective and mainly restricted to superficial anatomical structures or larger and deeper organs and neoplasia. In the Western World, the earliest description of this method was found in the Edwin Smith Papyrus scroll containing an ancient Egyptian medical text. It is believed that the author of this text was Imhotep, a priest, physician and presumed architect who lived between 3000 and 2500 B.C. A description of palpation techniques and results is also found in the *Corpus Hippocraticum* (Greece), a collection of ancient medical texts dating from the 5th to the 2nd century. Similar texts are found in the literature of the Far East but these have not been attributed to a single author.

Manual palpation still plays an important role in clinical practice, particularly in special situations, e. g., rectal digital examination of the prostate gland, intraoperative detection of certain structures, in self-examination of the female breast. However, it is a rather subjective method and is highly dependent on the experience of the person performing it. Elastographic methods are not a mere substitute for palpation but an additional technique that may allow further tissue characterization. The availability of semi-quantitative and quantitative methods to determine tissue stiffness allowed a real breakthrough in the diagnostic workup of patients including freehand, step and physiological displacement, low-frequency vibration, and radiation force techniques including acoustic emission shear wave and other impulse and displacement techniques.

## Early history of elastometry and elastography

Conventional ultrasound techniques are limited to image anatomy, the evaluation of tissue motion in real time, and the display of vessel anatomy blood flow mainly by Doppler techniques. In addition, emerging techniques exploit elastic properties of tissue including stiffness or hardness. The first elastometric and elastographic measurements using Young's or shear modulus were performed following classic mechanical models with the application of pressure and tension. From the physical point of view, elastography quantitatively maps Young's modulus, which corresponds to stiffness and shows the mechanical stress and strain of the material using a simplified approach via Hook's law [9]. However, as mechanical stress is not easily measurable *in vivo*, a strong assumption of homogeneous mechanical stress was initially assumed. Yet, considering different tissue structures within the human body, the application of pressure and tension as a homogeneous stress factor is not easily attainable [10–12]. In the beginning, simple (linear) properties of the underlying material were assumed with viscous and non-linear properties not taken into account.

Elasticity in general is described as the tendency of whatever material to resume its original size and shape after deformation by any kind of force or stress. Change in size or shape are defined as strain, which might be expressed as a ratio of a diameter (e. g., length) or as a histogram. Solid structures resist changes in shape and volume since they possess rigidity or shear elasticity, as well as volume elasticity. In contrast, fluids resist a change in volume, but not in shape as they possess only volume elasticity. The measurement of elastic properties (elastometry) of biological tissues was first described by King and Lawton. In their experiments, resonance was created with mechanical vibrations. They postulated that the

relative elasticity of the examined tissue could be obtained [13]. Using tension experiments, the tissue was described as a mechanically non-linear, anisotropic, and inhomogeneous viscoelastic medium [14]. Mechanical tissue properties were examined by determining the elasticity, particularly in muscle and heart samples [15–19]. In these experiments, a correlation between tissue elasticity and the acoustic reverberation coefficient was found [20]. The measurement and interpretation of superimposing waves have also been used for elastography of non-linear tissue properties [21], such as in femoral muscles using Doppler ultrasonographic techniques and in a model using a vibrator-based model with a mechanically moving piston [22]. The description of these techniques and their corresponding biological tissues was characterized by a lack of uniformly defined terminology.

Magnetic resonance elastography (MRE) was initially developed at the Mayo Clinic, Rochester, MN, in 1995 [23–27]. In this modality, a static shear modulus is calculated from the phase shift of the MR signal and the corresponding sites in three dimensions with high-speed volume acquisition. The extended scan times of MRE and the application of vibrators to generate appropriate shear waves were problematic due to the high demand regarding operation time [28–34]. The advantages include that MRE is not restricted by the presence of air, gas, or bone, whereas the advantages of USE include that elastometry of soft tissue strain and elasticity is faster, more convenient, more accurate, and more precise. For an extensive review of the history of ultrasonic elasticity imaging technology development, refer to the articles by Wells and Parker [9, 35].

## Definitions, Elastography, and Elastometry

Elastography is a qualitative method and describes the pictorial representation of tissue stiffness. Elastometry is a quantitative method with objective measurements. Currently, two main sonographic elastography techniques are used in medicine: compression elastography (also called strain or static elastography or simply elastography) and shear wave elastography (elastometry). The term “static elastography” was used in contrast to dynamic elastography (MRE, sonoelasticity) and transient elastography. Shear wave elastography is quantitative, and compression or static elastography is qualitative or comparative (harder or softer compared to the environment). Compression elastography is particularly useful for characterizing circumscribed changes, and shear wave elastography is useful for studying diffuse (infiltrative) changes of parenchymal organs.

Ophir and colleagues in Houston, Texas, USA first used the term “elastography” in 1991 [36]. This group was the first to present elastographic images comparing tissue stiffness in following publications [37–41]. External pressure was used to compress the tissue and the subsequent deformation was recorded using ultrasound with cross-correlation between the pre- and post-compression signals. Using a simple mechanical model, the modulus of elasticity can be derived from the measured strain and the applied stress (Hooke's law). The tissue strain was estimated from the deformation which was combined with assumptions about tissue geometry, homogeneous stress, and material models through image reconstruction algorithms that generated images of the underlying qualitative material elastic moduli, the so-called elasto-

gram [36]. As subsequent elasticity imaging methods were introduced, this method became known as “compression elastography”. Several years passed before compression elastography became commercially available. In fact, due to challenges with computation time and material model assumptions, commercial implementations do not generate “elastograms”, but rather bypass the reconstruction step and provide images of the tissue strain. As such, commercial implementation of compression elastography is broadly called “strain imaging”. Strain imaging became commercially available in the early 2000’s (i. e., “real-time elastography” by Hitachi, Tokyo, Japan was introduced in 2003). Siemens also introduced its “eSie Touch Elastography Imaging” with accepted clinical applications focusing on the differentiation of benign from malignant lesions in the breast and the prostate. The stiffness of the tissue is shown on a color image superimposed on the standard B-mode ultrasound image [10].

Since that time, all major ultrasound manufacturers have provided strain imaging features, and many other clinical applications have been explored and thousands of studies have been published.

Using a linear tissue model, stiffness parameters were postulated and derived from the observed tension in the target tissue. Cross-correlation was used to compare the measurements in the native and compressed states. A major disadvantage was the linear estimation function of the underlying tissue model, the assumption of spatially homogeneous stress, and the high number of measuring procedures. The first trials were performed in human muscles and breast [42].

### An anecdote about the color schema used in strain elastography

The attempt by scientific experts to call for a manufacturer-independent standardization of color coding at the EFSUMB guideline conference on elastography in Bologna in September 2012 was not able to prevail against the color associations of the company representatives, which had traditional meanings. For the Japanese manufacturing company Hitachi, “red” (the red circle in the center of the Japanese flag, the sun) symbolized vitality and strength, and thus has positive emotional connotations as the color of life. Thereby the color “red” stands for the sun and for the qualities clear, bright, brilliant, vital, benign and thus “soft(er)”. For manufacturers from Western cultures (for example, Supersonic Imagine), on the other hand, red is a color associated with aggressiveness and danger and is understood as a warning signal in a medical and technical context (fire). Therefore, ultrasound manufacturers from Europe and the USA code the tissue property “hard” with the color “red” in their elastography modules [43]. The manufacturer-dependent difference in the colors used to denote “stiffer” or “softer” must be taken into account during the examination.

### Ultrasound elastometry

This definition is focused on measurements. The term was used in the early years of development but has now been replaced by the term elastography.

### Freehand elastography

The development of freehand elastography was achieved by palpating the body using a hand-held transducer. The very first related ideas and reports were published by Christopher R. Hill [44], who inspired Jeff Bamber to explore this line of work in elastometry when he was a young postdoc [45–47]. The abstracts were presented at the World Federation for Ultrasound in Medicine and Biology conference Washington DC, United States (1988). This ended up being the strain imaging method implemented on most commercial systems, and differed from Ophir’s original concept of strain imaging, which used a motor-driven transducer. Eventually Ophir stopped developing that approach and switched to the freehand method introduced by Bamber et al. [48].

The work done by Bamber et al. then developed in two directions [49].

1. A combined autocorrelation algorithm for overcoming computer speed limitations, first published by Shiina et al. [50], was subsequently further developed and commercially released by Hitachi in about 2003.
2. A general strain imaging algorithm for freehand use mentioned in two prize-winning conference presentations [51] (winner of the BMUS oral presentation prize) and [52] (winner of the RSNA INFO-RAD presentation certificate of merit) eventually became the algorithm commercially released by Zonare Ltd., in about 2008, on their Z.one Ultra scanner [53].

### Sonoelastography

The terms sonoelastography and dynamic elastography broadly refer to dynamic elasticity imaging methods that use harmonic vibrations (with fixed frequencies, which differ from transient excitation) to mechanically excite tissue and reconstruction algorithms to estimate tissue material properties and ARFI-based techniques. The term sonoelastography was the result of a collaboration between the NIH (Rehabilitation Division) and the Tokyo Institute of Technology. Elasticity measurements were performed on patients and compared [54]. Krouskop et al. (1987) reported one of the first quantitative measurements of tissue elasticity using gated pulsed Doppler to track the velocity of internal tissue displacements generated by an externally applied vibration [22]. These first 1D non-invasive measurements of the elastic modulus of soft tissue in vivo were aimed at predicting the interaction of a prosthetic socket with an amputee’s residual limb.

Vibration amplitude sonoelastography was introduced by Lerner and Parker in 1987–88 [55–60]. Robert Lerner, a radiologist at the University of Rochester, had the experience of palpating prostate carcinomas that were not visible on ultrasound [35]. Subsequently, in 1987, Lerner and Parker conducted preliminary studies on vibration amplitude sonoelastography (sonoelasticity imaging) [57, 61]. The principle involved applying a low-frequency vibration (20–1000 Hz) externally to initiate internal vibrations in the tissue under investigation.

External vibrators generated shear waves in tissues, which were then monitored using Doppler sonography [57]. Transient elastography was introduced by Mathias Fink and collaborators from 1997 at the Laboratoire Ondes et Acoustique (now Institut Langevin) to

overcome the influence of boundary conditions observed in sonoelastography.

## Measurement of the stiffness of living tissue using ultrasonic radiation force

Sugimoto's group from the Tokyo Institute of Technology investigated the possibilities of tissue hardness measurements using the radiation force of focused ultrasound (ARFI) [62].

The authors used the force of a focused ultrasound beam to assess the longitudinal displacement of the tissue (like in strain elastography). A major possible disadvantage is that different elastic waves show significant interactions, such as conventional longitudinally orientated ultrasound waves (1500 m/s) modified by compression may be superimposed on the much weaker shear wave characteristics (1 m/s). However, in this study, the measurement is performed while avoiding the time zone affected by this shear wave after the radiation force is stopped. There is almost no effect on the displacement measurement in the focal region [63–66].

The deformation is measured by the pulsed Doppler method. As an advantage, this technique makes it possible to evaluate tissue hardness based on only the time dependence of relative displacement. By using the Voigt model, the viscoelastic value is derived from the change in deformation. Two-layer agar samples were employed, and a fundamental experiment was carried out to obtain the viscoelastic diagrams. These diagrams demonstrate the feasibility of the hardness measurement. The study was conducted between 1988 and 1993. For the first time, theoretical and experimental studies of *in vivo* measurement about tissue hardness were performed using focused ultrasound.

In 1988, in response to the request to display more qualitative information, such as hardness information, rather than just grayscale images, Professor Sadayuki Ueha of Tokyo Institute of Technology gave Tsuneyoshi Sugimoto, who was a master's student at the time, "tissue hardness measurement using radiation force" as a research topic. He continued this research as a doctoral research theme. The first problem was what was the hardness felt by the medical doctor. However, it was clear that, in general, biological tissue is a complex and viscoelastic body, and *in vivo* measurement is ultimately required. Because of that, no practical measurement method was established at that time. Therefore, with the cooperation of Professor Kouichi Itoh of Jichi Medical University, the relationship between the amount of pressurization and the amount of deformation was investigated for various human organs. As a result, it became clear that the hardness felt by human beings is related not only to elastic information but also to viscous information. After that, T. Sugimoto measured the deformation of tissue in the living body and its temporal decay due to the radiation pressure of focused ultrasound using the pulse echo method or the pulse Doppler method. A method to extract the feature quantity related to hardness from the deformation measurement was proposed. Moreover, in order to extend the proposed method to *in vivo* hardness evaluation, relative value measurement is indispensable. Then, a viscoelastic diagram was devised with elastic hardness on the vertical axis and viscous hardness on the horizontal axis. Furthermore, in order to evaluate the viscoelastic value of the bio-

logical tissue on the basis of the time change of displacement, a theoretical study was performed using the Voigt viscoelastic model, after the biological tissue was deformed by the radiation pressure of focused ultrasonic waves. As a result of the examination, it was clarified that the index value corresponding to each viscoelastic value of the model can be derived from the time change of displacement and the viscoelastic diagram can be created.

## Strain imaging, principles, and basic mechanisms

During conventional B-mode sonography with grayscale imaging, organs, including circumscribed lesions within the organs, can be palpated with the transducer by applying compression. Using this technique, the relative strain (or compression) of the target lesion can be evaluated in comparison to the surrounding tissues [67]. This can be semi-quantitated using a defined scale. The aim of this procedure is to gain information regarding tissue properties such as consistency, rigidity, size, and deformability. Ultrasound frequencies from 1 to 20 MHz with pulse repetitive frequencies of 1–10 kHz are used. In strain images, softer tissues will show a higher degree of deformation than stiffer tissues for a given compression (or applied stress). This deformation is visualized by ultrasonographic image processing with the examination being performed using standard ultrasound probes without any additional devices (pressure measurements, vibrators, etc.) similar to Doppler examination. The calculation of the strain is performed in real time with the results superimposed onto the conventional B-mode image [68].

## Probe systems, standardization of manual compression

### Clinical strain measurement techniques

Relative strain is obtained by measurements and transformed into images with the extended autocorrelation method by the ultrasound systems. Color coding with a blending of transparency up to 100% color saturation represents data translation of the obtained strain values. Using a measuring menu, a region of interest (ROI) within the elastographic window of the target lesion can be positioned and relative strain data obtained as the mean (%) of the areas within the ROI. These values can numerically supplement the color data, as is the case with thyroid examinations [69–73].

### Strain Ratio

#### Fat-to-lesion relationship

Linear correlations between different strain parameters were first investigated using appropriately sophisticated phantoms with embedded target structures. These results permit the definition of these properties more quantitatively in terms of strain ratios. If ROI "A" is positioned over the lesion to be examined (e. g., a tumor appearing in blue) and a second ROI "B" is placed over a reference tissue, e. g., fat deposits, then the lesion A (ROI A) is expected to appear with very low values (low elasticity) in relation to the reference structure (fat ROI B) which will show much higher values (high elasticity). In the clinical environment, these observations correspond to characteristics found in the human breast. The fatty tissue of this

organ can be regarded as the reference structure from which strain parameters can be determined and expressed as the ratio B/A (fat-to-lesion), which is a dimensionless number. If this ratio is around 1.2, a lesion is highly likely to be benign, whereas for a ratio of greater than 8.5 malignancy is suspected. This ratio comparing the tissue of interest to fat benefits from the fact that fat tissue has very similar stiffness values from patient to patient. The fat-to-lesion ratio reduces other variabilities due to the experiment and technology. Consequently, the number of false-positive test results was reduced, particularly for benign lesions. A cut-off of 3.2 was defined [74–79].

### Strain Histogram

More recent developments regarding elastography data processing include signal averaging of the last recorded strain sequence with subsequent quantitative color coding. With the strain-histology mode, an ROI can be placed within the elastobox with discrimination of the elastography data. The difference with respect to the above-described method is that discrimination of elastographically generated data is possible using this technique. This ROI is placed over a suspected area (with blue color elements) and a histogram of the relative stretching in a blue area is generated within the range 0 (blue) to 255. This ROI is of statistical importance with regard to size and circumference. Quantitative values (from 0 to 255) are then available based on the data derived from appropriate numerical tables. In addition, a statistical distribution of the non-stretchable areas (blue) can be given. Within the ROIs blue areas will be recorded in clusters and calculated with regard to area and circumference, thereby enabling a statistical description (complexity, standard deviation). This technology is thought to noninvasively evaluate parenchymatous organs such the liver for staging fibrosis in addition to the classic gold standard of biopsy. However, the results that have been obtained in European cohorts do not allow the technique to be recommended in this setting [80, 81].

### Transient Elastography

Transient elastography began at the laboratory of Mathias Fink, in Paris, France in the late 1990s [82–85]. The principle of transient elastography is to measure the speed of shear waves generated by a low-frequency transient vibration generated at the surface of a medium. The displacements induced in the medium by the propagation of the shear waves are tracked using ultrasound with a high frame rate (typically higher than 1000 frames per second). Initial developments by Catheline during his PhD under the supervision of Fink consisted of a transmission setup in which a mini-shaker and a single element ultrasound transducer were located on opposite sides of the to-be-measured medium [85, 86]. It was during the Sandrin PhD that the technique was extended to work in reflection mode, with the ultrasound probe directly located on the mini-shaker, thus resulting in a more compact and clinically applicable device. This research was the origin of the shear elasticity probe [83], while the diffraction effects associated with shear wave generation were better understood [87]. The transient elastography technique was initially a 1D technique. To extend transient elastography to 2D imaging, Fink had the idea of modifying the electronics of a time-reversal mirror made of an ultrasonic array (that he developed

in the early 90s) to make an ultrafast ultrasound scanner able to acquire several thousands of images per second to track the propagation of shear waves in 2D [88].

Fink and Sandrin developed this ultrafast imaging ultrasound scanner [89] to obtain elasticity maps of biological tissues [67, 69]. The initial idea of Fink's group was to put the ultrasonic array on a low-frequency shaker to both generate the shear wave in the body and to observe it with the ultrasound array [88]. However, the probe was heavy and not easy to manipulate.

They later decided to use ultrasonic radiation force to generate the shear waves using the supersonic mode. Interestingly, Fink, Sandrin, and Manneville also applied the ultrafast imaging ultrasound scanner to flow imaging including vortex [90, 91]. The Princeps patent of transient elastography [Princeps Transient Elastography patent US 6,770,033] which was filed in 1999 covers different embodiments of the technology. Initially proposed for the food industry (yogurt and camembert) applications, transient elastography ultimately became established in the medical field ▶ **Table 1**.

### Transient elastography in the food industry

The results from applications in the food industry were less successful than initial expectations [92]. Real-time assessment of the viscoelastic properties of yogurt during manufacturing was performed in 1998. Although encouraging results were obtained based on yogurt purchased at the grocery store, about 15 days after manufacturing, tests at the manufacturing site were very disappointing. As a matter of fact, fresh yogurt was ultrasound transparent in the first couple of days, thus preventing any attempt to perform ultrasound elastography. In the case of meat or muscle, tissue anisotropy leads to major difficulties triggered by the complexity of the shear wave propagation. Another problem is the attenuation of ultrasound waves by fat which was a major obstacle to the non-invasive determination of the stage of maturation of cheese such as camembert (unpublished data).

### Transient elastography in the medical field

In June 2001, Laurent Sandrin and colleagues founded the firm Echosens ([www.echosens.com](http://www.echosens.com)) to further develop the transient elastography technique which was still immature. They created algorithms to reliably measure shear wave speed in vivo and designed an elastography device able to control and synchronize the low-frequency vibration while performing pulse-echo ultrasound acquisitions. The improved technique named Vibration-Controlled Transient Elastography (VCTE) was patented in 2002 [Vibration-Controlled Transient Elastography patent US 7,578,789] ▶ **Fig. 1**.

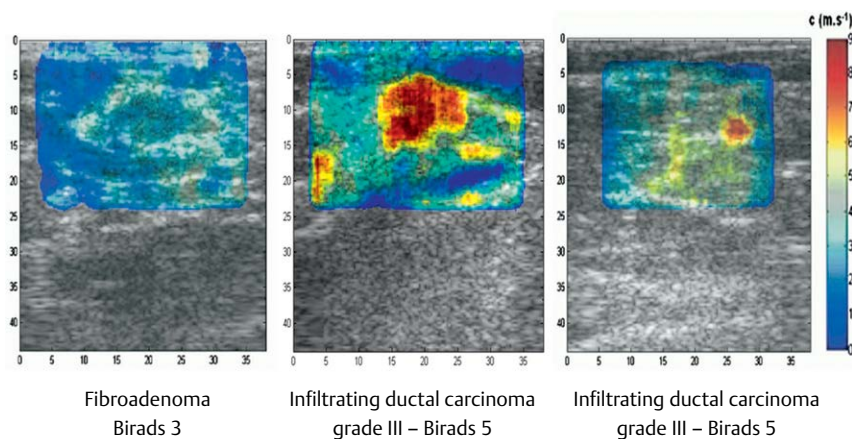
In parallel, they initiated a marketing survey to identify the potential market applications. During a meeting together with clinicians, the disadvantages regarding focal lesions such as breast and prostate cancers were discussed. The argument was that the differentiation of benign from malignant tissue by elastography might not be entirely reliable since even benign tissue can show different degrees of stiffness. A new important criterion regarding the evaluation of liver stiffness for fibrosis assessment was proposed. As stated in Hippocrates' (c. 460 – c. 370 BC) aphorisms, liver fibrosis is a well-known continuous process clearly associated with increased stiffness. Therefore, the Echosens team decided to focus



► **Table 1** Modern ultrasonographic elastography according to Cui et al. 2022 [126].

Technique	Measurement	Stimulation	Method	Indicator	Company	System
Strain imaging	Strain or displacement	Manual compression	Strain elastography	Elasticity score Strain ratio E/B size ratio	Esaote Hitachi Aloka GE Philips Toshiba Ultronix Mindray Samsung Siemens	ElaXto Real-time tissue elastography Elastography ElastoScan eSieTouch elasticity imaging
Shear wave imaging	Shear wave speed	Mechanical vibration	Transient elastography	Young's modulus (kPa)	Echosens	FibroScan
		ARFI	p-SWE	Shear wave speed (m/s) Young's modulus (kPa)	Siemens Philips	VTQ ElastPQ
			2D SWE	Shear wave speed (m/s) Young's modulus (kPa)	Siemens SuperSonic Imagine	VTIQ SWE
			3D SWE	Shear wave speed (m/s) Young's modulus (kPa)	SuperSonic Imagine	SWE

VTQ: Virtual Touch quantification; VTIQ: Virtual Touch image quantification; SWE: ShearWave elastography; ARFI: Acoustic radiation force impulse; p-SWE: Point-SWE.



► **Fig. 1** First clinical images of shear wave elastography (SWE) using ultrafast ultrasound imaging and acoustic radiation force generation for breast cancer diagnosis.

their efforts on in-vivo liver parenchyma stiffness evaluation. The clinical usefulness of transient elastography to evaluate quite homogeneous and continuously increasing fibrosis accumulation appeared promising.

The technique was initially tested during a pilot study at Institut Mutualiste Montsouris (Paris, France) in 2001–2002. Stiffness results obtained with VCTE were compared with the fibrosis stages determined by pathologists from liver biopsies [93]. In a following

multicentric study, the first clinically relevant results with evaluation of patients with chronic hepatitis C were published [94]. In December 2003, FibroScan obtained CE marking and was introduced on the market as the first commercially available noninvasive quantitative elastography device.

With FibroScan, shear waves are not only measured but can also be visually demonstrated. The tip of the probe is made of an ultrasound transducer mounted on the shaft of an electrodynamic ac-

tuator. The tip is actuated by transient excitation generated by the actuator, which induces shear waves in the tissues. Interestingly, only the longitudinal component of the shear waves is registered [87] since the displacements are measured on the ultrasound axis which is also the propagation axis of the shear waves. The controlled attenuation parameter (CAP) to detect and grade liver steatosis was introduced in 2010 [95]. Echosens was the first manufacturer to commercially offer stiffness measurement and ultrasound attenuation measurements in vivo. Since 2001, the company continues to develop the VCTE technique which is used in all devices named FibroScan [10].

## Acoustic Radiation Force Impulse (ARFI) based shear wave elastography

ARFI was developed by different researchers and commercially introduced by the Siemens company for the noninvasive measurement of liver stiffness. Sarvazyan et al. first proposed radiation force to make modulus maps [96]. Nightingale et al. first demonstrated in vivo use of radiation force of focused ultrasound to generate images [97] by measuring, point by point, the local displacement induced in tissue by the radiation force of a focused ultrasonic beam. Fink's group then decided to extend the radiation force concept to directly create a transient shear wave of high-amplitude to replace the one created by the mechanical shaker in 2D transient elastography. The two fundamental ideas behind the SSI technique are as follows: first, they proposed interleaved focused beam transmissions for shear wave generation and ultrafast plane wave transmissions for simultaneous ultrafast imaging. Second, they proposed to electronically control the radiation force pattern to create a fast moving "seismic" source of shear waves. If the source moves at a supersonic speed, it generates a high amplitude plane shear wave like in the "sonic boom" created by a supersonic plane. It was a very efficient way to produce shear waves in tissue. Tanter and Fink patented this SSI technique in September 2002 that led to the creation of Supersonic Imagine company in 2005. Bercoff, Tanter, and Fink used ultrafast imaging to detect the shear wave propagation produced by this supersonic seismic source [98]. This was the origin of the so-called supersonic shear wave scanner [98]. The supersonic shear imaging technique utilizes the underlying ultrafast ultrasound imaging technique developed for transient 2D elastography, but it replaces the vibrator with acoustic radiation pressure. First clinical validation of shear wave elastography using the SSI technique was published in 2008 [95] for breast cancer diagnosis. For a detailed review of the concept of ultrafast ultrasound imaging and applications in elastography and beyond, refer to [99].

Siemens introduced the first commercial SWE system in 2008. The Aixplorer (Supersonic Imagine, Aix-en-Provence, France) was then introduced in 2009 by Supersonic Imagine first for breast applications [98, 100–102] and other manufacturers followed [103].

ARFI-based shear wave elastography is an ultrasound technique, which is capable of measuring tissue elasticity quantitatively and also qualitatively (color-coded) when using 2D-SWE technology. Frequency-defined push pulses, with higher intensity and longer duration compared to that used for conventional B-mode images, lead to tissue vibrations at a region of interest (ROI) contributing

to the generation of shear waves (the higher the frequency, the higher the registered velocity). The velocity of these shear waves can be determined using the same ultrasound transducer. Based on certain assumptions, this velocity can be converted to tissue stiffness (kPa) using the Young's modulus  $E = 3\rho v_s^2$  [9]. As ARFI-based shear wave elastography did not rely on ultrafast frame rates, the shear wave propagation was not tracked in real time but rather reconstructed in a stroboscopic manner using several successive radiation force pushes and different imaging lines. This repetition of radiation force transmissions resulted in a higher acoustic energy deposit and thus limited the elastography frame rate and capabilities of ARFI-based shear wave elastography for real-time mapping of elasticity. McAleavey et al. developed a method called spatially modulated ultrasound radiation force (SMURF) [104]. In this method, instead of a sharp focal area, a spatially modulated pattern is used to create a unique spatial frequency within a tissue region.

## Fibrosis staging

It can be summarized that although VCTE has by far the highest level of evidence (more than 4,000 peer-reviewed publications in 2023), the number of publications on SWE has increased significantly in recent years due to the rapid evolution and spread of radiation force-based technologies. Several studies have demonstrated that both pSWE and 2D SWE techniques have similar accuracy to VCTE for the evaluation of liver fibrosis [5, 6, 103, 105]. However, VCTE and FibroScan are still considered to be the reference technique and the reference device, respectively, given the simplicity of use and high level of standardization of the technology.

## 2D Shear Wave Elastography

Two-dimensional shear wave elastography (2D-SWE) began in the laboratory of Mathias Fink, in Paris, France in the late 1990s [82–85, 89]. Standard ultrasound imaging was not able to efficiently track shear wave propagation in 2D due to its limited maximum frame rate of several hundred images per second. Fink and Sandrin [89] developed ultrafast ultrasound imaging which was initially obtained by only focusing in receive mode. Frame rates of several thousands of images per second were obtained and used to track shear wave propagation in 2D (both longitudinal and transverse tissue displacements) [82]. The absence of transmission focus yields a reduced penetration depth and lower contrast on B-mode ultrafast images. To solve this problem, Fink's group later introduced (in 2009) a multiple plane waves approach (today called coherent compound plane wave imaging) to synthesize virtual transmission focusing [106]. Bercoff, Tanter, and Fink further developed the technology in combination with radiation force for the generation of shear waves [107]. They first focused on studies with human breast, liver, and prostate, aiming at differentiating benign from malignant lesions and later on musculoskeletal applications [108, 109] and cardiovascular pathologies [110, 111]. With Jacques Souquet and Claude Cohen-Bacrie, they founded Supersonic Imagine with other colleagues in 2005. Both Echosens and Supersonic Imagine originated from Fink's laboratory.

## Further Developments

Echosens pioneered the field of liver stiffness and ultrasound attenuation assessments. Other manufacturers are developing or have already developed software for quantifying US beam attenuation. Preliminary results show that proprietary technologies implemented in ultrasound imaging systems seem more accurate than CAP for grading liver steatosis [112, 113]. Another available method for quantifying liver steatosis is based on the computation of the speed of sound. Initial results appear promising [114–116]. The backscatter coefficient also seems accurate for quantifying liver fat [117, 118].

## Modern elastography in clinical practice

Currently, the terms “elastography” and “elasticity imaging” are used broadly to describe imaging methods that provide information about tissue stiffness, including strain imaging, compression elastography, sonoelastography, acoustic radiation force impulse imaging, ultrasound shear wave elastography, magnetic resonance elastography, optical coherence elastography, et cetera.

Presently the following ultrasonic elastographic methods are widely being used in clinical medicine: strain imaging, ARFI imaging (both provide relative images of tissue stiffness), shear wave elastography (including vibration-controlled transient elastography), and cardiac strain imaging [119]. Shear wave elastography is quantitative, whereas strain and ARFI imaging yield qualitative images (harder vs. softer compared to the surrounding structures). The qualitative methods have mainly been used to characterize circumscribed lesions and heterogeneous tissues, whereas shear wave elastography allows both the evaluation of diffuse infiltrations of organs and circumscribed lesions [68, 120, 121].

Elastography has now found wide application including in pediatric patients with a large number of publications [122]. In addition to the assessment of liver fibrosis, the status assessment of breast lesions and thyroid nodules, multiparametric imaging of the prostate in the search for prostate carcinoma, there is a variety of other applications including the status assessment of lymph nodes, the assessment of the strength of any focal lesions compared to the surrounding parenchyma, the assessment of the age of vascular thrombi, assessment of the vascular wall, intestinal wall assessment in Crohn's disease, and last but not least for neurosurgery of brain tumors [35, 70]. Shear wave elastography was also recently identified as “the new wave of cardiovascular biomechanics” [123] since myocardial stiffness is assumed to be a very relevant biomarker of cardiac pathologies [124]. Recent studies suggest that diastolic myocardial elasticity is higher in heart failure with preserved ejection fraction (HFpEF) patients. Systolic myocardial elasticity was also shown to noninvasively measure cardiac myocardial contractility. Another challenging application is the combination of strain elastography and semiquantitative methods with endoscopic ultrasound.

By quantifying the elasticity of muscles, joints, ligaments, nerves, and soft-tissue musculoskeletal masses, shear wave elastography is also very promising for the follow-up of various traumatic and pathologic conditions of the musculoskeletal system [125].

## Elastography in endoscopic ultrasound (EUS)

Endoscopic ultrasound strain elastography is an important investigative component in the evaluation of focal lesions in the pancreas, gastrointestinal tract, and all structures in the sonographic window of EUS and lymph nodes [127–129]. Elastography in EUS evolved with the use of electronic EUS probes. Strain elastography in EUS is not without controversy. In order to be able to evaluate the findings, investigators must familiarize themselves with the method and study data. Profound background knowledge is required. The findings must be viewed in the overall context of B-mode, color Doppler imaging, and contrast-enhanced harmonic EUS imaging data. In EUS, strain elastography is used to evaluate the hardness of lymph nodes [130–133], pancreatic lesions [130, 134–142], and intramural wall structures of the gastrointestinal tract. The result of strain elastography in EUS is only one piece of the overall examination and is subject to some influencing factors. Elastography is possible with both longitudinal and radial probes. The transducer of the EUS probe is coupled to the wall with slight pressure. The interesting lesion should be centered. The elastography window, the region of interest, must be sufficiently large to detect the lesion of interest in comparison to surrounding tissues. By comparing the elasticity of the lesion with the surrounding tissue, the strain ratio can be calculated [131, 141, 143, 144]. Additional software can be used to calculate the mean histogram value from multiple images [132, 137]. Difficulties in deriving a strain elastography image in EUS result from very small lesions, deeply located lesions, and pulsations from the heart or large vessels. Limitations regarding the differentiation between benign and malignant lesions are caused by fibrosis (lymph nodes and pancreas) [136–139, 145], tumor necrosis, and liquid and semiliquid components. A lymph node or pancreatic lesion must be assessed in the overall evaluation of sonomorphology, color duplex imaging, and contrast-enhanced harmonic EUS together with strain elastography. Elastography in EUS is quite helpful for making clinical decisions. Lymph nodes can be selected for EUS-FNA using strain elastography. A pancreatic lesion that is hypervascularized and endosonographically soft is highly unlikely to correspond to ductal adenocarcinoma [146].

## Current terminology and guidelines on the use of ultrasound elastography

The first European Federation of Societies for Ultrasound in Medicine and Biology (EFSUMB) guidelines and recommendations on the clinical use of elastography described the basic principles and technology and clinical applications in detail [147–149]. This was followed by the publication of the World Federation for Ultrasound in Medicine and Biology (WFUMB) 2015 and the Society of Radiologists in Ultrasound (SRU) 2015 [150]. All of these guidelines list the relative advantages of the different elastographic methods. The 2013 EFSUMB guidelines and recommendations on the clinical use of elastography were updated in 2017, with a focus on the assessment of diffuse liver disease [103, 105]. An update was also provided by WFUMB for liver elastography [80] and by the SRU [151]. In



addition to the 2013 EFSUMB guidelines, updates for non-hepatic applications were published in 2019 [70].

The basic principles of elastography remain unchanged since they were outlined in the first part of the original EFSUMB guidelines and WFUMB guidelines on this subject [147, 148]. The term *elastography* is considered a type of remote palpation that allows measurement and display of biomechanical properties associated with the elastic restoring forces in the tissue that act against shear deformation. This view unifies the different types of elastography, namely strain imaging, acoustic radiation force impulse imaging, and shear wave elastography (SWE) and explains why they all display images with contrast for the same underlying information, associated with the shear elastic modulus. The type of elastography most suited for the assessment of diffuse liver disease is SWE, which measures the speed of a shear wave in the liver. In the 2017 update [103, 105], VCTE is regarded as a type of SWE, although it differs from other SWE methods because it uses a body-surface vibration to create a shear wave, which then travels to the liver, whereas the other methods use acoustic radiation force to remotely create the shear wave within the liver. VCTE also generates shear waves at lower frequencies (around 50 Hz) compared to radiation force-based methods (around 200 Hz) leading to different shear wave speed values due to shear wave dispersion.

A variety of methods that measure shear wave speed should be grouped under the term SWE [81, 147, 148]. Ultrasound elastography uses ultrasonic echoes to observe tissue displacement as a function of time and space after applying a force that is either dynamic (e. g., thumping or vibrating) or varying so slowly that it is considered “quasi-static” (e. g., probe palpation). The displacement measurements may be represented in an elasticity image (elastogram), or as a local measurement, in one of three ways:

- Tissue displacement may be displayed directly, as in the method known as acoustic radiation force impulse (ARFI) imaging,
- Tissue strain may be calculated from the spatial gradient of displacement and displayed, producing what is termed strain elastography (SE), or
- When the force is dynamic only, the time-varying displacement data may be used to record at various positions the arrival times of propagating shear waves. These are used to calculate either regional values of shear wave speed (without making images) using methods referred to as transient elastography (TE, including VCTE) and point shear wave elastography (pSWE), or images of shear wave speed using methods referred to as 2D (or 3D) shear wave elastography (SWE). All of the methods in this third group come under the heading of SWE [80, 103, 105].

In order to properly introduce the method of elastography to the general public, the corresponding “How to perform” recommendations are available in addition to the guidelines [152–154].

## Conclusions and future perspectives

Compared to more than 2000 years of palpation, it is possible to look back on the successful and effective development of elastography thanks to the effective work of a large number of teams in

the last 35 years. Elastography is an important additional imaging modality in many specialties. It should be actively used by investigators. In the evaluation of liver fibrosis, histology can be omitted in a large number of patients. In the overall context with B-mode sonography, color Doppler imaging, and contrast-enhanced US or EUS, lesions and their status can be more reliably delineated. Developments that compensate for the limitations of elastography, e. g., increased fat content, would be desirable to more accurately predict fibrosis even in hepatic steatosis. More accurate tissue analysis would be helpful to further delineate focal lesions. In contrast to the modern technique, there is still a widespread perception that elastography is mainly nice and colorful but not very helpful. Through continuing education and communication of information, this misconception should be eliminated so that examiners are even better able to make accurate diagnoses.

## Acknowledgement

The authors thank Hubert Allgayer, Jeff Bamber, Giovanna Ferraioli and Kathy Nightingale for advice.

## Conflict of Interest

The authors declare that they have no conflict of interest.

## References

- [1] Dietrich CF, Cantisani V. Current status and perspectives of elastography. *Eur J Radiol* 2014; 83: 403–404
- [2] Bamber JC. Comment on new technology--ultrasound elastography. *Ultraschall Med* 2008; 29: 319–320
- [3] Dietrich CF, Bolondi L, Duck F, Evans DH, Ewertsen C, Fraser AG et al. History of Ultrasound in Medicine from its birth to date (2022), on occasion of the 50 Years Anniversary of EFSUMB. A publication of the European Federation of Societies for Ultrasound In Medicine and Biology (EFSUMB), designed to record the historical development of medical ultrasound. *Med Ultrason* 2022; 24: 434–450
- [4] Merz E, Evans DH, Dong Y, Jenssen C, Dietrich CF. History of ultrasound in obstetrics and gynaecology from 1971 to 2021 on occasion of the 50 years anniversary of EFSUMB. *Med Ultrason* 2023; 25: 175–188
- [5] Jenssen C, Ewertsen C, Piscaglia F, Dietrich CF, Gilja OH, Sidhu PS et al. 50th years anniversary of EFSUMB: Initial roots, maturation, and new shoots. *Ultraschall Med* 2022; 43: 227–231
- [6] Christian Jenssen CE, Christoph F. Dietrich, Alina Popescu, Lynne Rudd European Federation of Societies for Ultrasound in Medicine and Biology (EFSUMB), London, United Kingdom. 50 Years of EFSUMB. Past, Present and Future. 2022.
- [7] Dietrich CF, Greis C. History of contrast enhanced ultrasound. In: Lyshchik A, Dietrich CF, Sidhu P, Wilson S, eds. *Fundamentals in contrast enhanced ultrasound (CEUS)*. Philadelphia: Elsevier; 2019: 4–8
- [8] Nielsen MB, Sogaard SB, Bech Andersen S, Skjoldbye B, Hansen KL, Rafaelsen S et al. Highlights of the development in ultrasound during the last 70 years: A historical review. *Acta Radiol* 2021; 62: 1499–1514

- [9] Wells PN, Liang HD. Medical ultrasound: imaging of soft tissue strain and elasticity. *J R Soc Interface* 2011; 8: 1521–1549
- [10] Gennisson JL, Deffieux T, Fink M, Tanter M. Ultrasound elastography: principles and techniques. *Diagn Interv Imaging* 2013; 94: 487–495
- [11] Dewart RJ. Ultrasound elastography: principles, techniques, and clinical applications. *Crit Rev Biomed Eng* 2013; 41: 1–19
- [12] Tang A, Cloutier G, Szeverenyi NM, Sirlin CB. Ultrasound Elastography and MR Elastography for Assessing Liver Fibrosis: Part 1, Principles and Techniques. *AJR American journal of roentgenology* 2015; 205: 22–32
- [13] King AL, Plawton RW. Elasticity of Body Tissues: Chicago Year Book Publ. 1950 u. 1960, Medical Physics, Vol II, Vol.III; 1960.
- [14] Hartung C. Zur Biomechanik weicher Gewebe. Düsseldorf: VDI-Verlag; 1975
- [15] Mol CR, Breddels PA. Ultrasound velocity in muscle. *J Acoust Soc Am* 1982; 71: 455–461
- [16] Levinson SF. Ultrasound propagation in anisotropic soft tissues: the application of linear elastic theory. *J Biomech* 1987; 20: 251–260
- [17] O'Donnell M, Mimbs JW, Miller JG. Relationship between collagen and ultrasonic backscatter in myocardial tissue. *J Acoust Soc Am* 1981; 69: 580–588
- [18] Madaras EI, Perez J, Sobel BE, Mottley JG, Miller JG. Anisotropy of the ultrasonic backscatter of myocardial tissue: II. Measurements in vivo. *J Acoust Soc Am* 1988; 83: 762–769
- [19] Wear KA, Milunski MR, Wickline SA, Perez JE, Sobel BE, Miller JG. Contraction-related variation in frequency dependence of acoustic properties of canine myocardium. *J Acoust Soc Am* 1989; 86: 2067–2072
- [20] Hete B, Shung KK. A study of the relationship between mechanical and ultrasonic properties of dystrophic and normal skeletal muscle. *Ultrasound Med Biol* 1995; 21: 343–352
- [21] Zhang D, Gong X, Ye S. Acoustic nonlinearity parameter tomography for biological specimens via measurements of the second harmonics. *J Acoust Soc Am* 1996; 99: 2397–2402
- [22] Krouskop TA, Dougherty DR, Vinson FS. A pulsed Doppler ultrasonic system for making noninvasive measurements of the mechanical properties of soft tissue. *Journal of rehabilitation research and development* 1987; 24: 1–8
- [23] Muthupillai R, Lomas DJ, Rossman PJ, Greenleaf JF, Manduca A, Ehman RL. Magnetic resonance elastography by direct visualization of propagating acoustic strain waves. *Science* 1995; 269: 1854–1857
- [24] Muthupillai R, Ehman RL. Magnetic resonance elastography. *Nat Med* 1996; 2: 601–603
- [25] Muthupillai R, Rossman PJ, Lomas DJ, Greenleaf JF, Riederer SJ, Ehman RL. Magnetic resonance imaging of transverse acoustic strain waves. *Magn Reson Med* 1996; 36: 266–274
- [26] Manduca A, Muthupillai R. Visualization of Tissue Elasticity by Magnetic Resonance Elastography. *Lecture Notes in Computer Science Bd 1131* 1996; 63–68
- [27] Manduca A. Multispectral image visualization with nonlinear projections. *IEEE transactions on image processing: a publication of the IEEE Signal Processing Society* 1996; 5: 1486–1490
- [28] Catheline S, Gennisson JL, Delon G, Fink M, Sinkus R, Abouelkaram S et al. Measuring of viscoelastic properties of homogeneous soft solid using transient elastography: an inverse problem approach. *J Acoust Soc Am* 2004; 116: 3734–3741
- [29] Sinkus R, Tanter M, Catheline S, Lorenzen J, Kuhl C, Sondermann E et al. Imaging anisotropic and viscous properties of breast tissue by magnetic resonance-elastography. *Magn Reson Med* 2005; 53: 372–387
- [30] Sinkus R, Tanter M, Xydeas T, Catheline S, Bercoff J, Fink M. Viscoelastic shear properties of in vivo breast lesions measured by MR elastography. *Magn Reson Imaging* 2005; 23: 159–165
- [31] Sinkus R, Siegmann K, Xydeas T, Tanter M, Clausen C, Fink M. MR elastography of breast lesions: understanding the solid/liquid duality can improve the specificity of contrast-enhanced MR mammography. *Magn Reson Med* 2007; 58: 1135–1144
- [32] Robert B, Sinkus R, Gennisson JL, Fink M. Application of DENSE-MR-elastography to the human heart. *Magn Reson Med* 2009; 62: 1155–1163
- [33] Salameh N, Larrat B, Abarca-Quinones J, Pallu S, Dorvillius M, Leclercq I et al. Early detection of steatohepatitis in fatty rat liver by using MR elastography. *Radiology*. 2009; 253: 90–97
- [34] Larrat B, Pernot M, Aubry JF, Dervishi E, Sinkus R, Seilhean D et al. MR-guided transcranial brain HIFU in small animal models. *Phys Med Biol* 2010; 55: 365–388
- [35] Parker KJ, Doyle MM, Rubens DJ. Imaging the elastic properties of tissue: the 20 year perspective. *Phys Med Biol* 2011; 56: R1–R29
- [36] Ophir J, Cespedes I, Ponnekanti H, Yazdi Y, Li X. Elastography: a quantitative method for imaging the elasticity of biological tissues. *Ultrason Imaging* 1991; 13: 111–134
- [37] Cespedes I, Ophir J, Ponnekanti H, Maklad N. Elastography: elasticity imaging using ultrasound with application to muscle and breast in vivo. *Ultrason Imaging* 1993; 15: 73–88
- [38] Ponnekanti H, Ophir J, Cespedes I. Ultrasonic imaging of the stress distribution in elastic media due to an external compressor. *Ultrasound Med Biol* 1994; 20: 27–33
- [39] Ophir J, Miller RK, Ponnekanti H, Cespedes I, Whittaker AD. Elastography of beef muscle. *Meat science* 1994; 36: 239–250
- [40] Ponnekanti H, Ophir J, Huang Y, Cespedes I. Fundamental mechanical limitations on the visualization of elasticity contrast in elastography. *Ultrasound Med Biol* 1995; 21: 533–543
- [41] Ophir J, Alam SK, Garra BS, Kallel F, Konofagou EE, Krouskop T et al. Elastography: Imaging the elastic properties of soft tissues with ultrasound. *J Med Ultrason* (2001) 2002; 29: 155
- [42] Chen EJ, Novakofski J, Jenkins WK, Brien WDO. Young's modulus measurements of soft tissues with application to elasticity imaging. *IEEE Transactions on Ultrasonics, Ferroelectrics, and Frequency Control* 1996; 43: 191–194
- [43] Jansen C, Lauer N, Burmester E, Will U, Hocke M, Dietrich C. „Art meets Science“ – Gedanken zu Schnittstellen zwischen Kunst und Wissenschaft. *Das Spannungsfeld von Information und ästhetischer Faszination in Endoskopie und Bildgebung. Endoskopie heute* 2013; 26: 2–12
- [44] Dickinson RJ, Hill CR. Measurement of soft tissue motion using correlation between A-scans. *Ultrasound Med Biol* 1982; 8: 263–271
- [45] Tristram M, Barbosa DC, Cosgrove DO, Nassiri DK, Bamber JC, Hill CR. Ultrasonic study of in vivo kinetic characteristics of human tissues. *Ultrasound Med Biol* 1986; 12: 927–937
- [46] Hill CR, Tristram M, Bamber JC, Blaszczyk M, Barbosa DC, Cosgrove D et al. Ultrasonic remote palpation (URP): Use of shear elastic modulus to differentiate pathology. *J Ultrasound Med*. 1988; 7: S129
- [47] Bamber JC, Hasan P, Cook-Martin G, Bush NL. Parametric imaging of tissue shear and flow imaging using B-scan decorrelation rate. *J Ultrasound Med* 1988; 7: S135–S136
- [48] Bamber JC, Bush NL. Freehand Elasticity Imaging Using Speckle Decorrelation Rate. In: Tortoli P, Masotti L, editors. *Acoustical Imaging*. Boston, MA: Springer US; 1996: 285–292
- [49] Bamber JC, Barbone PE, Bush NL, Cosgrove D, Doyely MM, Fuechsel FG et al. Progress in Freehand Elastography of the Breast. *IEICE Transactions on Information and Systems* 2002; 85: 5–14

- [50] Shiina T, Doyley MM, Bamber JC. editors. Strain imaging using combined RF and envelope autocorrelation processing. Proceedings IEEE Ultrasonics Symposium; 1996.
- [51] Bamber JC, Fuechsel FG, Bush NL, Tranquart F, Cosgrove DO, Miller NM et al. Freehand elasticity imaging of breast masses: a preliminary clinical study. *European J Ultrasound* 2001; 13: S13
- [52] Fuechsel FG, Bush NL, Tranquart F, Bamber JC, Cosgrove DO, Miller NR. Ultrasound freehand elastography: Evaluation of diagnostic potential in clinical breast imaging. *Radiology* 2001; 221: 188
- [53] Doyley MM, Bamber JC, Fuechsel F, Bush NL. A freehand elastographic imaging approach for clinical breast imaging: system development and performance evaluation. *Ultrasound Med Biol* 2001; 27: 1347–1357
- [54] Levinson SF, Shinagawa M, Sato T. Sonoelastic determination of human skeletal muscle elasticity. *J Biomech* 1995; 28: 1145–1154
- [55] Lerner RM. Sono-elasticity'-Images derived from Ultrasound Signals in Mechanically Vibrated Targets: Report of the Commission of the European Communities Paper-Nr.: EUR11816EN; 1988.
- [56] Parker KJ, Huang SR, Musulin RA, Lerner RM. Tissue response to mechanical vibrations for "sonoelasticity imaging". *Ultrasound Med Biol* 1990; 16: 241–246
- [57] Lerner RM, Huang SR, Parker KJ. "Sonoelasticity" images derived from ultrasound signals in mechanically vibrated tissues. *Ultrasound Med Biol* 1990; 16: 231–239
- [58] Lee F Jr., Bronson JP, Lerner RM, Parker KJ, Huang SR, Roach DJ. Sonoelasticity imaging: results in in vitro tissue specimens. *Radiology* 1991; 181: 237–239
- [59] Rubens DJ, Hadley MA, Alam SK, Gao L, Mayer RD, Parker KJ. Sonoelasticity imaging of prostate cancer: in vitro results. *Radiology* 1995; 195: 379–383
- [60] Gao L, Parker KJ, Alam SK, Lerner RM. Sonoelasticity imaging: theory and experimental verification. *J Acoust Soc Am* 1995; 97: 3875–3886
- [61] Lerner RM, Parker KJ. Sonoelasticity images derived from ultrasound signals in mechanically vibrated targets Proc. 7th European Communities Workshop (Oct 1987, Nijmegen, The Netherlands). 1987.
- [62] Sugimoto T, Ueha S, Itoh K. editors. Tissue hardness measurement using the radiation force of focused ultrasound. Proc IEEE Ultrasonics Symposium; 1990.
- [63] Sugimoto T, Ueha S, Itoh K. Tissue Hardness Measurement using ultrasonic radiation force. *J J Appl Phys* 1992; Suppl 31: 166–168
- [64] Sugimoto T, Ueha S, Itoh K. Tissue hardness measurement using ultrasonic radiation force, theory and experiment using Voigt model. *J Med Ultrasonics* 1993; 20–5: 277–283
- [65] Sugimoto T, Ueha S, Itoh K. A Evaluation Method of Tissue Hardness Using Relaxation Elastic Moduli Study of Measurement Theory and in vitro Experiment. *Jpn J Medical Electronics and Biological Engineering* 1991; 29: 635–641
- [66] Sugimoto T, Ueha S, Itoh K. Tissue Basic research on hardness measurement of biological tissues: -In vivo measurement theory and experiments using Voigt model. Proc of the spring meeting the Acoustical Society of Japan 1993; II: 753–754
- [67] Dietrich CF, Barr RG, Farrokh A, Dighe M, Hocke M, Jenssen C et al. Strain Elastography - How To Do It? *Ultrasound Int Open* 2017; 3: E137–E149
- [68] Dietrich CF. Echtzeit-Gewebeelastographie. Anwendungsmöglichkeiten nicht nur im Gastrointestinaltrakt. *Endoskopie Heute* 2010; 23: 177–212
- [69] Dietrich CF, Muller T, Bojunga J, Dong Y, Mauri G, Radzina M et al. Statement and Recommendations on Interventional Ultrasound as a Thyroid Diagnostic and Treatment Procedure. *Ultrasound Med Biol* 2018; 44: 14–36
- [70] Saftoiu A, Gilja OH, Sidhu PS, Dietrich CF, Cantisani V, Amy D et al. The EFSUMB Guidelines and Recommendations for the Clinical Practice of Elastography in Non-Hepatic Applications: Update 2018. *Ultraschall Med* 2019; 40: 425–453
- [71] Friedrich-Rust M, Vorlaender C, Dietrich CF, Kratzer W, Blank W, Schuler A et al. Evaluation of Strain Elastography for Differentiation of Thyroid Nodules: Results of a Prospective DEGUM Multicenter Study. *Ultraschall Med* 2016; 37: 262–270
- [72] Dighe M, Barr R, Bojunga J, Cantisani V, Chammas MC, Cosgrove D et al. Thyroid Ultrasound: State of the Art. Part 2 – Focal Thyroid Lesions. *Med Ultrason* 2017; 19: 195–210
- [73] Cosgrove D, Barr R, Bojunga J, Cantisani V, Chammas MC, Dighe M et al. WFUMB Guidelines and Recommendations on the Clinical Use of Ultrasound Elastography: Part 4. Thyroid. *Ultrasound Med Biol* 2017; 43: 4–26
- [74] Farrokh A, Treu L, Ohlinger R, Fliieger C, Maass N, Schafer FK. A Prospective Two Center Study Comparing Breast Cancer Lesion Size Defined by 2D Shear Wave Elastography, B-Mode Ultrasound, and Mammography with the Histopathological Size. *Ultraschall Med* 2019; 40: 212–220
- [75] Farrokh A, Maass N, Treu L, Heilmann T, Schafer FK. Accuracy of tumor size measurement: comparison of B-mode ultrasound, strain elastography, and 2D and 3D shear wave elastography with histopathological lesion size. *Acta Radiol* 2019; 60: 451–458
- [76] Farrokh A, Schaefer F, Degenhardt F, Maass N. Comparison of Two Different Ultrasound Devices Using Strain Elastography Technology in the Diagnosis of Breast Lesions Related to the Histologic Results. *Ultrasound Med Biol* 2018; 44: 978–985
- [77] Barr RG, Nakashima K, Amy D, Cosgrove D, Farrokh A, Schafer F et al. WFUMB guidelines and recommendations for clinical use of ultrasound elastography: Part 2: breast. *Ultrasound Med Biol* 2015; 41: 1148–1160
- [78] Farrokh A, Wojcinski S, Degenhardt F. Evaluation of real-time tissue sono-elastography in the assessment of 214 breast lesions: limitations of this method resulting from different histologic subtypes, tumor size and tumor localization. *Ultrasound Med Biol* 2013; 39: 2264–2271
- [79] Jiang M, Li CL, Luo XM, Chuan ZR, Lv WZ, Li X et al. Ultrasound-based deep learning radiomics in the assessment of pathological complete response to neoadjuvant chemotherapy in locally advanced breast cancer. *Eur J Cancer* 2021; 147: 95–105
- [80] Ferraioli G, Wong VW, Castera L, Berzigotti A, Sporea I, Dietrich CF et al. Liver Ultrasound Elastography: An Update to the World Federation for Ultrasound in Medicine and Biology Guidelines and Recommendations. *Ultrasound Med Biol* 2018; 44: 2419–2440
- [81] Ferraioli G, Filice C, Castera L, Choi BI, Sporea I, Wilson SR et al. WFUMB guidelines and recommendations for clinical use of ultrasound elastography: Part 3: liver. *Ultrasound Med Biol* 2015; 41: 1161–1179
- [82] Tanter M, Bercoff J, Sandrin L, Fink M. Ultrafast compound imaging for 2-D motion vector estimation: application to transient elastography. *IEEE Trans Ultrason Ferroelectr Freq Control* 2002; 49: 1363–1374
- [83] Sandrin L, Tanter M, Gennisson JL, Catheline S, Fink M. Shear elasticity probe for soft tissues with 1-D transient elastography. *IEEE Trans Ultrason Ferroelectr Freq Control* 2002; 49: 436–446
- [84] Sandrin L, Tanter M, Catheline S, Fink M. Shear modulus imaging with 2-D transient elastography. *IEEE Trans Ultrason Ferroelectr Freq Control* 2002; 49: 426–435
- [85] Catheline S, Thomas JL, Wu F, Fink MA. Diffraction field of a low frequency vibrator in soft tissues using transient elastography. *IEEE Trans Ultrason Ferroelectr Freq Control* 1999; 46: 1013–1019

- [86] Catheline S. Interférométrie-speckle ultrasonore: application à la mesure d'élasticité. Thèse de doctorat. Université Paris VII, 1998.
- [87] Sandrin L, Cassereau D, Fink M. The role of the coupling term in transient elastography. *J Acoust Soc Am* 2004; 115: 73–83
- [88] Bercoff J, Chaffai S, Tanter M, Sandrin L, Catheline S, Fink M et al. In vivo breast tumor detection using transient elastography. *Ultrasound Med Biol* 2003; 29: 1387–1396
- [89] Sandrin L, Catheline S, Tanter M, Hennequin X, Fink M. Time-resolved pulsed elastography with ultrafast ultrasonic imaging. *Ultrasound Imaging* 1999; 21: 259–272
- [90] Sandrin L, Manneville S, Fink M. Ultrafast two-dimensional ultrasonic speckle velocimetry: A tool in flow imaging. *Applied Physics Letters* 2001; 78: 1155–1157
- [91] Manneville S, Sandrin L, Fink M. Investigating a stretched vortex with ultrafast two-dimensional ultrasonic speckle velocimetry. *Physics of Fluids* 2001; 13: 1683–1690
- [92] Povey MJW. Ultrasonics in food engineering Part II: Applications. *Journal of Food Engineering* 1989; 9: 1–20
- [93] Sandrin L, Fourquet B, Hasquenoph JM, Yon S, Fournier C, Mal F et al. Transient elastography: a new noninvasive method for assessment of hepatic fibrosis. *Ultrasound Med Biol* 2003; 29: 1705–1713
- [94] Ziolo M, Handra-Luca A, Kettaneh A, Christidis C, Mal F, Kazemi F et al. Noninvasive assessment of liver fibrosis by measurement of stiffness in patients with chronic hepatitis C. *Hepatology* 2005; 41: 48–54
- [95] Sasso M, Beaugrand M, de Ledinghen V, Douvin C, Marcellin P, Poupon R et al. Controlled attenuation parameter (CAP): a novel VCTE guided ultrasonic attenuation measurement for the evaluation of hepatic steatosis: preliminary study and validation in a cohort of patients with chronic liver disease from various causes. *Ultrasound Med Biol* 2010; 36: 1825–1835
- [96] Sarvazyan AP, Rudenko OV, Swanson SD, Fowlkes JB, Emelianov SY. Shear wave elasticity imaging: a new ultrasonic technology of medical diagnostics. *Ultrasound Med Biol* 1998; 24: 1419–1435
- [97] Nightingale KR, Palmeri ML, Nightingale RW, Trahey GE. On the feasibility of remote palpation using acoustic radiation force. *J Acoust Soc Am* 2001; 110: 625–634
- [98] Bercoff J, Tanter M, Fink M. Supersonic shear imaging: a new technique for soft tissue elasticity mapping. *IEEE Trans Ultrason Ferroelectr Freq Control* 2004; 51: 396–409
- [99] Tanter M, Fink M. Ultrafast imaging in biomedical ultrasound. *IEEE Trans Ultrason Ferroelectr Freq Control* 2014; 61: 102–119
- [100] Tanter M, Bercoff J, Athanasiou A, Deffieux T, Gennisson JL, Montaldo G et al. Quantitative assessment of breast lesion viscoelasticity: initial clinical results using supersonic shear imaging. *Ultrasound Med Biol* 2008; 34: 1373–1386
- [101] Athanasiou A, Tardivon A, Tanter M, Sigal-Zafrani B, Bercoff J, Deffieux T et al. Breast lesions: quantitative elastography with supersonic shear imaging--preliminary results. *Radiology*. 2010; 256: 297–303
- [102] Cosgrove DO, Berg WA, Dore CJ, Skyba DM, Henry JP, Gay J et al. Shear wave elastography for breast masses is highly reproducible. *Eur Radiol* 2012; 22: 1023–1032
- [103] Dietrich CF, Bamber J, Berzigotti A, Bota S, Cantisani V, Castera L et al. EFSUMB Guidelines and Recommendations on the Clinical Use of Liver Ultrasound Elastography, Update 2017 (Long Version). *Ultraschall Med* 2017; 38: e16–e47
- [104] McAleavey S, Collins E, Kelly J, Elegebe E, Menon M. Validation of SMURF estimation of shear modulus in hydrogels. *Ultrasound Imaging* 2009; 31: 131–150
- [105] Dietrich CF, Bamber J, Berzigotti A, Bota S, Cantisani V, Castera L et al. EFSUMB Guidelines and Recommendations on the Clinical Use of Liver Ultrasound Elastography, Update 2017 (Short Version). *Ultraschall Med* 2017; 38: 377–394
- [106] Montaldo G, Tanter M, Bercoff J, Benec N, Fink M. Coherent plane-wave compounding for very high frame rate ultrasonography and transient elastography. *IEEE Trans Ultrason Ferroelectr Freq Control* 2009; 56: 489–506
- [107] Fink M, Tanter M. Multiwave imaging and super resolution. *Physics Today* 2010; 63: 28–33
- [108] Gennisson JL, Deffieux T, Mace E, Montaldo G, Fink M, Tanter M. Viscoelastic and anisotropic mechanical properties of in vivo muscle tissue assessed by supersonic shear imaging. *Ultrasound Med Biol* 2010; 36: 789–801
- [109] Hug F, Tucker K, Gennisson JL, Tanter M, Nordez A. Elastography for Muscle Biomechanics: Toward the Estimation of Individual Muscle Force. *Exerc Sport Sci Rev* 2015; 43: 125–133
- [110] Pernot M, Lee WN, Bel A, Mateo P, Couade M, Tanter M et al. Shear Wave Imaging of Passive Diastolic Myocardial Stiffness: Stunned Versus Infarcted Myocardium. *JACC Cardiovasc Imaging* 2016; 9: 1023–1030
- [111] Couade M, Pernot M, Prada C, Messas E, Emmerich J, Bruneval P et al. Quantitative assessment of arterial wall biomechanical properties using shear wave imaging. *Ultrasound Med Biol* 2010; 36: 1662–1676
- [112] Ferraioli G, Maiocchi L, Raciti MV, Tinelli C, De Silvestri A, Nichetti M et al. Detection of Liver Steatosis With a Novel Ultrasound-Based Technique: A Pilot Study Using MRI-Derived Proton Density Fat Fraction as the Gold Standard. *Clin Transl Gastroenterol* 2019; 10: e00081
- [113] Fujiwara Y, Kuroda H, Abe T, Ishida K, Oguri T, Noguchi S et al. The B-Mode Image-Guided Ultrasound Attenuation Parameter Accurately Detects Hepatic Steatosis in Chronic Liver Disease. *Ultrasound Med Biol* 2018; 44: 2223–2232
- [114] Imbault M, Faccinnetto A, Osmanski BF, Tissier A, Deffieux T, Gennisson JL et al. Robust sound speed estimation for ultrasound-based hepatic steatosis assessment. *Phys Med Biol* 2017; 62: 3582–3598
- [115] Imbault M, Dioguardi Burgio M, Faccinnetto A, Ronot M, Bendjador H, Deffieux T et al. Ultrasonic fat fraction quantification using in vivo adaptive sound speed estimation. *Phys Med Biol* 2018; 63: 215013
- [116] Stahl P, Becchetti C, Korta Martiartu N, Berzigotti A, Frenz M, Jaeger M. First-in-human diagnostic study of hepatic steatosis with computed ultrasound tomography in echo mode. *Commun Med (Lond)* 2023; 3: 176
- [117] Zhu Y, Yin H, Zhou D, Zhao Q, Wang K, Fan Y et al. A prospective comparison of three ultrasound-based techniques in quantitative diagnosis of hepatic steatosis in NAFLD. *Abdom Radiol (NY)*. 2024; 49: 81–92. DOI: 10.1007/s00261-023-04078-7
- [118] Ferraioli G, Berzigotti A, Barr RG, Choi BI, Cui XW, Dong Y et al. Quantification of Liver Fat Content with Ultrasound: A WFUMB Position Paper. *Ultrasound Med Biol* 2021; 47: 2803–2820
- [119] Amzulescu MS, De Craene M, Langet H, Pasquet A, Vancraeynest D, Pouleur AC et al. Myocardial strain imaging: review of general principles, validation, and sources of discrepancies. *Eur Heart J Cardiovasc Imaging* 2019; 20: 605–619
- [120] Frey H. [Realtime elastography. A new ultrasound procedure for the reconstruction of tissue elasticity]. *Der Radiologe* 2003; 43: 850–855
- [121] Ignee A, Jenssen C, Hocke M, Dong Y, Wang WP, Cui XW et al. Contrast-enhanced (endoscopic) ultrasound and endoscopic ultrasound elastography in gastrointestinal stromal tumors. *Endosc Ultrasound* 2017; 6: 55–60
- [122] Dietrich CF, Ferraioli G, Sirli R, Popescu A, Sporea I, Pienar C et al. General advice in ultrasound based elastography of pediatric patients. *Med Ultrason* 2019; 21: 315–326
- [123] Sengupta PP, Chandrashekar Y. The New Wave of Cardiovascular Biomechanics. *JACC Cardiovasc Imaging* 2019; 12: 1297–1299

- [124] Villemain O, Correia M, Mousseaux E, Baranger J, Zarka S, Podetti I et al. Myocardial Stiffness Evaluation Using Noninvasive Shear Wave Imaging in Healthy and Hypertrophic Cardiomyopathic Adults. *JACC Cardiovasc Imaging* 2019; 12: 1135–1145
- [125] Taljanovic MS, Gimber LH, Becker GW, Latt LD, Klauser AS, Melville DM et al. Shear-Wave Elastography: Basic Physics and Musculoskeletal Applications. *Radiographics: a review publication of the Radiological Society of North America, Inc* 2017; 37: 855–870
- [126] Cui XW, Li KN, Yi AJ, Wang B, Wei Q, Wu CG et al. Ultrasound elastography. *Endosc Ultrasound* 2022; 11: 252–274
- [127] Dietrich CF, Saftoiu A, Janssen C. Real time elastography endoscopic ultrasound (RTE-EUS), a comprehensive review. *Eur J Radiol* 2014; 83: 405–414
- [128] Dietrich CF, Bibby E, Janssen C, Saftoiu A, Iglesias-Garcia J, Havre RF. EUS elastography: How to do it? *Endosc Ultrasound* 2018; 7: 20–28
- [129] Dietrich CF, Burmeister S, Hollerbach S, Arcidiacono PG, Braden B, Fusaroli P et al. Do we need elastography for EUS? *Endosc Ultrasound* 2020; 9: 284–290
- [130] Giovannini M, Thomas B, Erwan B, Christian P, Fabrice C, Benjamin E et al. Endoscopic ultrasound elastography for evaluation of lymph nodes and pancreatic masses: a multicenter study. *World J Gastroenterol* 2009; 15: 1587–1593
- [131] Larsen MH, Fristrup C, Hansen TP, Hovendal CP, Mortensen MB. Endoscopic ultrasound, endoscopic sonoelastography, and strain ratio evaluation of lymph nodes with histology as gold standard. *Endoscopy* 2012; 44: 759–766
- [132] Saftoiu A, Vilmann P, Hassan H, Gorunescu F. Analysis of endoscopic ultrasound elastography used for characterisation and differentiation of benign and malignant lymph nodes. *Ultraschall Med* 2006; 27: 535–542
- [133] Dietrich CF, Janssen C, Arcidiacono PG, Cui XW, Giovannini M, Hocke M et al. Endoscopic ultrasound: Elastographic lymph node evaluation. *Endosc Ultrasound* 2015; 4: 176–190
- [134] Hirche TO, Ignee A, Barreiros AP, Schreiber-Dietrich D, Jungblut S, Ott M et al. Indications and limitations of endoscopic ultrasound elastography for evaluation of focal pancreatic lesions. *Endoscopy* 2008; 40: 910–917
- [135] Saftoiu A, Vilmann P, Gorunescu F, Janssen J, Hocke M, Larsen M et al. Efficacy of an artificial neural network-based approach to endoscopic ultrasound elastography in diagnosis of focal pancreatic masses. *Clin Gastroenterol Hepatol* 2012; 10: 84–90 e1
- [136] Janssen J, Schlorer E, Greiner L. EUS elastography of the pancreas: feasibility and pattern description of the normal pancreas, chronic pancreatitis, and focal pancreatic lesions. *Gastrointestinal endoscopy* 2007; 65: 971–978
- [137] Saftoiu A, Vilmann P, Gorunescu F, Gheonea DI, Gorunescu M, Ciurea T et al. Neural network analysis of dynamic sequences of EUS elastography used for the differential diagnosis of chronic pancreatitis and pancreatic cancer. *Gastrointest Endosc* 2008; 68: 1086–1094
- [138] Saftoiu A, Vilmann P, Gorunescu F, Janssen J, Hocke M, Larsen M et al. Accuracy of endoscopic ultrasound elastography used for differential diagnosis of focal pancreatic masses: a multicenter study. *Endoscopy* 2011; 43: 596–603
- [139] Dietrich CF, Hirche TO, Ott M, Ignee A. Real-time tissue elastography in the diagnosis of autoimmune pancreatitis. *Endoscopy* 2009; 41: 718–720
- [140] Iglesias-Garcia J, Larino-Noia J, Abdulkader I, Forteza J, Dominguez-Munoz JE. Quantitative endoscopic ultrasound elastography: an accurate method for the differentiation of solid pancreatic masses. *Gastroenterology*. 2010; 139: 1172–1180
- [141] Itokawa F, Itoi T, Sofuni A, Kurihara T, Tsuchiya T, Ishii K et al. EUS elastography combined with the strain ratio of tissue elasticity for diagnosis of solid pancreatic masses. *Journal of gastroenterology* 2011; 46: 843–853
- [142] Ignee A, Janssen C, Arcidiacono PG, Hocke M, Moller K, Saftoiu A et al. Endoscopic ultrasound elastography of small solid pancreatic lesions: a multicenter study. *Endoscopy* 2018; 50: 1071–1079
- [143] Fusaroli P, Saftoiu A, Mancino MG, Caletti G, Eloubeidi MA. Techniques of image enhancement in EUS (with videos). *Gastrointestinal endoscopy* 2011; 74: 645–655
- [144] Puga-Tejada M, Del Valle R, Oleas R, Egas-Izquierdo M, Arevalo-Mora M, Baquerizo-Burgos J et al. Endoscopic ultrasound elastography for malignant pancreatic masses and associated lymph nodes: Critical evaluation of strain ratio cutoff value. *World journal of gastrointestinal endoscopy* 2022; 14: 524–535
- [145] Dietrich CF, Hocke M. Elastography of the Pancreas, *Current View. Clin Endosc* 2019; 52: 533–540
- [146] Dietrich CF, Sahai AV, D'Onofrio M, Will U, Arcidiacono PG, Petrone MC et al. Differential diagnosis of small solid pancreatic lesions. *Gastrointest Endosc* 2016; 84: 933–940
- [147] Bamber J, Cosgrove D, Dietrich CF, Fromageau J, Bojunga J, Calliada F et al. EFSUMB guidelines and recommendations on the clinical use of ultrasound elastography. Part 1: Basic principles and technology. *Ultraschall Med* 2013; 34: 169–184
- [148] Cosgrove D, Piscaglia F, Bamber J, Bojunga J, Correias JM, Gilja OH et al. EFSUMB guidelines and recommendations on the clinical use of ultrasound elastography. Part 2: Clinical applications. *Ultraschall Med* 2013; 34: 238–253
- [149] Cui XW, Friedrich-Rust M, De Molo C, Ignee A, Schreiber-Dietrich D, Dietrich CF. Liver elastography, comments on EFSUMB elastography guidelines 2013. *World J Gastroenterol* 2013; 19: 6329–6347
- [150] Barr RG, Ferraioli G, Palmeri ML, Goodman ZD, Garcia-Tsao G, Rubin J et al. Elastography Assessment of Liver Fibrosis: Society of Radiologists in Ultrasound Consensus Conference Statement. *Radiology* 2015; 276: 845–861
- [151] Barr RG, Wilson SR, Rubens D, Garcia-Tsao G, Ferraioli G. Update to the Society of Radiologists in Ultrasound Liver Elastography Consensus Statement. *Radiology* 2020; 296: 263–274
- [152] Dietrich CF, Shi L, Wei Q, Dong Y, Cui XW, Lowe A et al. What does liver elastography measure? Technical aspects and methodology. *Minerva Gastroenterol (Torino)* 2021; 67: 129–140
- [153] Ferraioli G, Barr RG, Farrokh A, Radzina M, Cui XW, Dong Y et al. How to perform shear wave elastography. Part I. *Med Ultrason* 2022; 24: 95–106
- [154] Ferraioli G, Barr RG, Farrokh A, Radzina M, Cui XW, Dong Y et al. How to perform shear wave elastography. Part II. *Med Ultrason* 2022; 24: 196–210