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# A *De novo* Mutation in the *COL1A1* Gene Leading to Severe Osteogenesis Imperfecta: Case Report and Review of the Literature

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# Abstract Introduction Oste ited skeletal dysplas

**Introduction** Osteogenesis imperfecta (OI) is the most common monogenic inherited skeletal dysplasia disorder. Mutations in the *COL1A1/COL1A2* gene cause ~85 to 90% of OI. Studies of cases have demonstrated that missense mutations are the primary cause of OI, with poor prognosis.

**Case Description** We report the case of a fetus with skeletal abnormalities and subcutaneous edema. Ultrasound imaging revealed suspected skeletal malformations, including hypoplastic long bones of all four limbs, poorly ossified calvarium, unrevealing nasal bones, and generalized subcutaneous edema. Whole-exome sequencing revealed a heterozygous mutation in *COL1A1* (c.2174G > T/p.(G725V), NM\_000088.3). According to the American College of Medical Genetics and Genomics guidelines, it was determined to be a pathogenic variant and identified as a de novo variant (PS2 + PP3\_strong + PM2\_supporting), which has not been reported in the HGMD, gnomAD, ClinVar, or other databases. This variation causes a glycine-to-valine substitution at position 725, located within the Gly-Xaa-Yaa repeat in the helical domain of the collagen molecule.

# Keywords

- osteogenesis imperfecta
- ► COL1A1 gene
- whole-exome sequencing
- prenatal diagnosis

the collagen molecule. **Conclusion** The *COL1A1* mutation (c.2174G > T/p.(G725V), NM\_000088.3) is a novel pathogenic variant of severe OI. Our study expanded the OI *COL1A1* gene variation profiles in the Chinese population and provided a theoretical foundation for prenatal diagnosis, genetic counseling, and obstetric management.

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As one of the most common congenital disorders, fetal skeletal dysplasia is characterized by genetic and phenotypic heterogeneity, making it difficult to differentiate between these diseases on ultrasound.<sup>1</sup> Osteogenesis imperfecta (OI) is the most prevalent monogenic inherited skeletal dysplasia disease. It is predominantly autosomal dominant, but a few autosomal recessive cases have been reported, and it is rarely attributed to X-linked inheritance.<sup>2</sup> The incidence of OI ranges from ~1:15,000 to 1:20,000 births.<sup>3</sup> The main clinical manifestations of OI include short stature, skeletal anomalies, increased skeletal fragility, multiple fractures, other skeletal system pathologies, and extraskeletal manifestations such as dentinogenesis imperfecta, malocclusion, pulmonary dysfunction, and hearing impairment.<sup>4</sup> In 1979, Sillence et al initially categorized OI into four types, I to IV, based on patients' phenotypic characteristics and severity. In 2019, the Chinese guidelines for rare disease diagnosis and treatment further classified it into five types, I to V. Among them, type II OI is fatal, and the phenotype is the most serious, with patients usually succumbing in the perinatal period, and there is no efficacious treatment for live births.<sup>5,6</sup> For fetuses with potential OI suggested by ultrasound, genetic testing to ascertain the etiology and prenatal intervention are optimal prevention and management strategies, respectively. In this study, we conducted whole-exome sequencing (WES) of the fetus, referring to prenatal ultrasound suggestive of OI, with normal copy number variation sequencing (CNV-seq) and classic karyotype. To validate the disease-associated candidate variant sites detected by WES, Sanger sequencing was used to elucidate the fetal genetic etiology and provide a basis for prenatal intervention, genetic counseling, and risk assessment of fertility in the family. Following the CARE reporting checklist, we present the following case.

#### **Case Presentation**

A 24-year-old Chinese Han primigravida was found to carry a fetus with suspected skeletal dysplasia, which prompted our attention. No abnormalities were observed in the first trimester, and the couple did not have a history of radiation substance exposure or medication use, as claimed by the couple of primigravida. Additionally, the couple was previously healthy and did not have a family history of skeletal dysplasia or consanguineous marriage. During the routine checkup in the second trimester, other prenatal examinations did not reveal any abnormalities. However, ultrasound imaging at 18 weeks of gestation revealed suspected skeletal malformations, including hypoplastic long bones of all four limbs, poorly ossified calvarium, unrevealing nasal bones, and generalized subcutaneous edema ( **Fig. 1**). Details of the suspected fetuses are shown in **-Table 1**. Subsequently, diagnostic screening was conducted by collecting amniotic fluid from fetal parents and peripheral blood samples.

After obtaining consent from the couple, we received a 20mL sample of amniotic fluid via amniocentesis at 18 weeks of gestation. The amniotic fluid sample obtained was aliquoted into two tubes. One tube was utilized for chromosome karyotype analysis, and the other tube (QIAamp DNA Blood Mini Kit produced by German company Qiagen) was used to extract genomic DNA for CNV-seq and Trio-WES; 2-mL samples of each spouse's peripheral blood were collected to extract genomic DNA (Magnetic Blood Genomic DNA Kit from Tiangen Biochemical Technology Company) for Trio-WES.

Karyotype analysis revealed no abnormalities in the chromosome number or structure (**-Fig. 2A**). CNV-seq analysis did not reveal any chromosome aneuploidy or CNVs in the genome, with definite pathogenicity of more than 100 kb (**-Fig. 2B**). We identified a heterozygous mutation of *COL1A1* (c.2174G > T/p.(G725V), NM\_000088.3), which were not present in either parent. Sanger sequencing corroborated the sequencing results for the whole exon group (**-Fig. 2**), and this variation was confirmed to be a de novo variation through sequencing of the parental samples.

The mutation was deemed pathogenic based on multiple in silico predictions of variant pathogenicity (**-Table 2**), with a Rare Exome Variant Ensemble Learner (REVEL) software prediction score of 0.997. This variation was absent from the ESP6500, ExAC, and 1000 Genome databases. Following the American College of Medical Genetics and Genomics (ACMG) guidelines, the variation was pathogenic (PS2 + PP3\_strong + PM2\_supporting). This mutation was not present in HGMD, gnomAD, ClinVar, or other databases, indicating that it was a novel mutation site.

Collagen type 1 (Col1) is the principal connective tissue that forms the body's skeleton, sclere, skin, and teeth. Collagen type 1 is a protein composed of  $\alpha$  1 and  $\alpha$  2 chains. We used Uniport (https://www.uniprot.org/) to process protein conservation analysis at the mutation site 725, and the results showed that it is a conservation site for various species (**> Fig. 2D**).

#### Discussion

Ol is a congenital skeletal dysplasia with a genetic classification of 22 types (OI types I–XXII) (https://omim.org). However, there is considerable overlap in the clinical phenotypes of these different types. As of November 11, 2022, the OI and Ehlers–Danlos' syndrome variant databases have included 20 genes associated with OI. Studies have demonstrated that pathogenic mutations cause ~85 to 90% of OI in the type I collagen ligand  $\alpha$ 1 subunit/ $\alpha$ 2 subunit (*COL1A1/COL1A2*). Despite the identification of thousands of pathogenic mutation types, a precise genotype–phenotype correlation has yet to be established.<sup>7</sup>

In clinical practice, OI is classified into four types (I–IV) based on the clinical phenotype and severity. Type I OI is characterized by the mildest clinical phenotype, typically without skeletal malformations. The clinical phenotype of type II OI is the most grievous, with the most affected individuals dying in the perinatal period due to severe bone deformities. Some severe cases also present with fetal edema.<sup>8</sup> Type III OI is severe, presenting with progressive skeletal malformation and high-frequency fractures as well as abnormal teeth and hearing loss, and has a poor prognosis. Type IV OI is of moderate severity, between types I and III.<sup>5,6</sup>

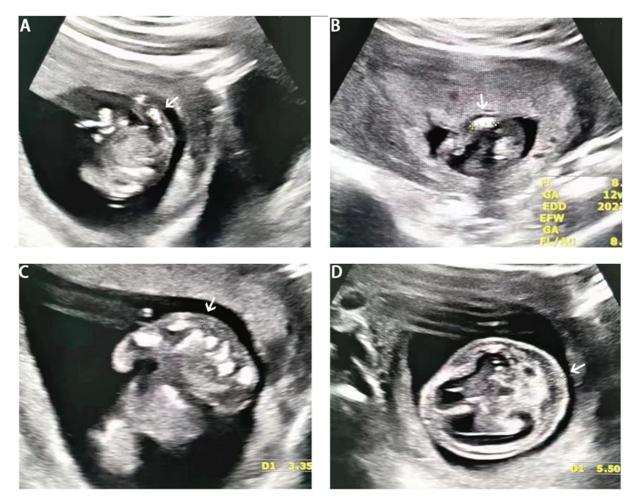


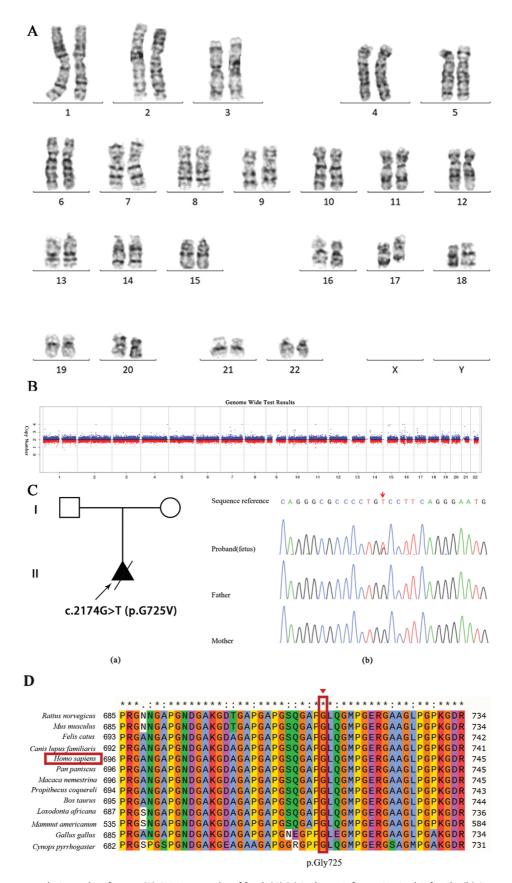
Fig. 1 Ultrasound images of cases. (A) Humeral length: 7 mm. (B) Femur length: 8 mm. (C) The abdomen was ~3.5 mm thick. (D) The scalp was ~5.5 mm thick.

Ultrasonographic measurements	Measured value (mm)	Normal range in 18 weeks' gestation (mm)
Biparietal diameter	42	34–45
Head circumference	59	128–161
Transverse diameter of the cerebellum	16	15.9–20.5
Abdominal circumference	108	110–141
Femoral length	8	21–29
Humeral length	7	22.3-30.1
Fetal weight	118 (g)	184–269 (g)
Fetal skin thickness	Scalp thickness: 5.5 Abdominal skin thickness: 3.5	-
Other	The nasal bone of the fetus was not shown, the lip structure was not clearly displayed, the ossification of the skull ring was not obvious, the cerebellum was visible, the midline of the brain was in the middle, and the width of bilateral ventricles was $\sim$ 4 mm	-

Table 1 Ultrasonographic measurements of suspected fetal skeletal dysplasia

The *COL1A1* is located at 17q21.33, with a total length of 18 kb and 51 exons. Code-shifting, nonsense, and splicing mutations of *COL1A1* primarily lead to premature termina-

tion codons, producing hazardous and degradable truncated gene expression products. OI caused by such mutations typically presents mild clinical manifestations. Missense



**Fig. 2** (A) Karyotype analysis results of cases. (B) CNV-seq results of fetal. (C) (a) Pedigree information in this family. (b) Sanger validation peak plots for the proband (fetus) and family members. The fetal *COL1A1* gene has c.2174G > T / p.(G725V) heterozygous variation, which parents do not carry. (D) Protein conservation analysis in the mutation site 725 showed that it is a conservation site for various species (https://www.uniprot.org/). CNV-seq, copy number variation sequencing.

Protein prediction algorithm	Score	Interpretation
Scale-Invariant Feature Transform (SIFT)	0	Damaging
Polymorphism Phenotyping Version 2_HDIV (Polyphen2 HDIV)	1	Probably damaging
Polymorphism Phenotyping Version 2_HVAR (Polyphen2 HVAR)	0.999	Probably damaging
Mutation Taster	1	Disease-causing
Protein Variation Effect Analyzer (PROVEAN)	-7.53	Damaging
Rare Exome Variant Ensemble Learner (REVEL)	0.997	Damaging
Combined Annotation-Dependent Deletion (CADD)	34	Damaging
phyloP	7.867	Conserved
phastCons	1	Conserved
SiPhy	18.429	Conserved

Table 2 Effects of COL1A1 mutations predicted with in silico tools

mutations in COL1A1 are associated with the production of proteins with abnormal structures. The mutant subunits participate in and disrupt the triple helix assembly of type I collagen, exacerbating the dominant-negative effect. These mutations typically result in severe phenotypes.<sup>9</sup> Studies in China, Italy, Poland, and other countries have demonstrated that missense mutations are the primary cause of OI with poor prognosis, the majority of which involve the substitution of glycine in the Gly-X-Y (X and Y are random amino acids) repeat sequences of the collagen molecular helix domain with other amino acids.<sup>10–13</sup> The substitution of glycine with branched nonpolar or charged amino acids likely results in a more severe form of OI.<sup>14</sup> Studies conducted in other contexts have corroborated that de novo mutations, which have not been subjected to purifying selection, are associated with a more severe clinical presentation of OI than inherited COL1A1 mutations.15,16

This was the first pregnancy for couples who were not blood-related. No significant skeletal dysplasia or similar clinical manifestations were observed in the family history. Obstetric ultrasonography revealed severe limb shortening, poor skull ossification, absence of nasal bones, and generalized subcutaneous edema. According to research findings, fetuses afflicted with lethal skeletal dysplasia display considerable and premature reduction in the length of their long bones as well as disproportionate growth of the fetal abdomen and cranium. Notably, the ratio of femoral length to abdominal circumference serves as a reliable indicator of skeletal dysplasia lethality. Specifically, 92 to 96% of cases showed a ratio of femoral length to abdominal circumference below 0.16, which is a sign of fatal skeletal dysplasia.<sup>17</sup> The available sonographic parameters indicate the presence of a hypoplastic fetal skeletal system, which is indicative of severe OI. Studies have demonstrated that when a pregnant woman and her spouse have no apparent skeletal system malformations, prenatal ultrasound imaging reveals abnormalities of the fetal skeletal system in conjunction with other system malformations, which are likely caused by chromosomal abnormalities or CNVs. When fetal malformations are mainly observed in the skeletal system, they are usually

monogenic in origin.<sup>18</sup> In this study, karyotyping and CNVseq did not reveal any abnormalities, suggesting that nonchromosomal numerical, structural, and CNVs were associated with skeletal malformations and other ultrasonic manifestations. Trio-WES revealed that the fetus in this case carried the heterozygous c.2174G > T/p.(G725V) variant, which is categorized as a pathogenic variant based on ACMG guidelines. Neither of the parents had this mutation site, which was not present in the database or reported in the literature. Among the candidate genes, the missense variant in COL1A1 appeared to be the most frequently observed pathogenic variant associated with OI. Therefore, the identified mutation appears to have arisen spontaneously during the early stages of human embryonic development. Based on this and the ultrasound-specific clinical presentation, the fetus was initially diagnosed with perinatal lethal OI. The fetal COL1A1 gene had a c.2174G > T/p.(G725V) variation, causing a glycine-to-valine substitution at position 725, which is located within the Gly-Xaa-Yaa repeat in the helical domain of the collagen molecule and is a branched nonpolar amino acid, which might be the most responsible for the more severe fetal clinical phenotype. We have summarized the mutated genes of severe fetal OI caused by missense mutations in COL1A1 and their associated clinical ultrasonic manifestations reported in the literature (**-Table 3**). Ultrasound images exhibit high similarity, primarily displaying a shortened humerus, shortened femur, thoracic stenosis, and a poorly ossified calvarium. This finding was consistent with the ultrasound findings reported in the present study. Consequently, in the absence of a prenatal genetic diagnosis, ultrasound during pregnancy suggests that the aforementioned fetal manifestations could be utilized as the initial diagnostic basis. The patient chose to terminate their pregnancy after genetic confirmation. Regrettably, owing to the lack of aborted tissue, we were unable to definitively ascertain whether it was a perinatal lethal OI.

OI is one of the primary causes of skeletal dysplasia in fetuses for which no adequate therapy exists. Currently, nonlethal patients are managed symptomatically, primarily through lifestyle modification, medications, surgery, and

References	Gene nucleotide change	Pathogenicity level (evidence)	Fetal gestational age	Radiographic finding	Extraskeletal manifestations in ultrasound examination	Fetal sample for WES
ji et al <sup>21</sup>	Case 1: c.1634G > A (p.Gly545Asp)	1	Case 1: 17 wk	Case 1: The fetus is characterized by short humeri (10 mm) and short femora (10 mm). Skull ossification is poor with borderline personality disorder (41 mm) and circumference (150 mm)	Case 1: A narrow chest and notch can be seen in the junction of the chest and abdomen, and the maxi- mal pocket of amniotic fluid was 36 mm	Case 1: Amniotic fluid
Tanner et al <sup>22</sup>	Case 1: c.3290G > T (p.Gly1097Val)	1	Case 1: 21 <sup>4/7</sup> wk	Case 1: Extremely short limbs corresponding roughly to the gestational age of 12 wk. The thorax was so small; computed tomography showed the fetus was abnormally short and poorly ossified long bones and absent ossification of the skull were evident. The fetus was aborted at 22 wk	1	Case 1: Tissue after fetal abortion
Huang et al <sup>23</sup>	Case 1: c.1822G > A (p.Gly608Ser)	1	Case 1: 20 wk	Case 1: Short and incurved limbs, a small thorax, bowing long bones, and a reduced echogenic ring around the intracranial structures: Fetus was aborted at 17 wk	1	Case 1: Tissue after fetal abortion
Zhang et al <sup>24</sup>	Case 1: c.994G > A (p.Gly332Arg) Case 2: c.2362C > A (p.Gly788Ser) Case 3: c.2444G > C (p.Gly815Ala) Case 4: c.3505G > A (p.Gly1169Ser) Case 5: c.3541G > A (p.Gly1181Ser)	Case 1: P (PS2 + PM1 + PM2 + PP3 + PP4) Case 2: LP (PM1 + PM2 + PP3 + PP4) Case 3: LP (PM1 + PM2 + PP3 + PP4) Case 4: LP (PS2 + PM1 + PM2 + PP3 + PP4) Case 5: LP (PS2 + PM1 + PM2 + PP3 + PP4)	Case 1: 24 <sup>6/7</sup> wk Case 2: 31 <sup>3/7</sup> wk Case 3: 30 <sup>1/7</sup> wk Case 4: 26+ wk Case 5: 21+ wk	Case 1: Fetal whole body bone sono- graphic changes, clinical suspicion of OI Case 2: The fetal femur and humerus were $\sim 4$ wk+ less than the corrected gestational age, and no obvious abnormality was found before 31 wk Case 3: The long bones of limbs are less than gestational age, and bilat- eral femurs are slightly curved Case 4: Fetal femurs are short and slightly curved on both sides Case 5: The long bones of the fetus limbs are less than 4 standard deviations, and the thoracic cavity is narrow; the nasal bone is absent	Case 3: The diameter of the oval is larger, and the left ventricle has strong light spots Case 5: The eye spacing is widened; the scalp edema, the brain parenchyma is poor, the subdural effusion, and the abdominal fluid is effusion	Unknown
Yang et al <sup>25</sup>	Case 1: c.1678G > A (p.Gly560Ser) Case 2: c.2101G > A	Case 1: P (PP2 + PM2 + PM5 strong + PS4 sup- porting + PS2 + PP3)	Case 1: 23 <sup>4/7</sup> wk Case 2: 22 <sup>6/7</sup> wk Case 3: 22 wk; 20 <sup>6/7</sup>	Case 1: The long bones of the fetus' limbs were short and curved. The fetus was aborted at $24^{1/7}$ wk	Case 4: nuchal translucen- cy thickening (6.0 mm), anasarca	Case 1: Umbili- cal cord Case 2:

Table 3 Gene and clinical manifestation of fetal severe OI caused by multiple missense mutations COL 1A1

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+ + <sup>2</sup> N + <sup>4</sup>
Case 1: PS2_very strong + PM5 + PM2 supporting + PP3
Case 1: PVS1 + PM2 + Case 1: 22 wk PM5 + PP2 + PP3 Case 2: PS2 + PM2 + Case 2: 24 <sup>5/7</sup> wk PP3 Case 3: PS2 + PM1 + PM2 + PP3 + PP5
Case 1: PVS1 + PM2 + Case 1: 22 wk PM5 + PP2 + PP3 Case 2: 24 <sup>517</sup> wk Case 2: PS2 + PM2 + Case 3: 20 <sup>617</sup> wk PP3 Case 3: PS2 + PM1 + PM2 + PP3 + PP5

Table 3 (Continued)

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References	Gene nucleotide change	Pathogenicity level (evidence)	Fetal gestational age	Radiographic finding	Extraskeletal manifestations in ultrasound examination	Fetal sample for WES
Yang et al <sup>29</sup>	Case 1: c.2605G > T (p.Gly869Cvs Case 2: c.1804G > A (p.Gly602Arg)	1	Case 1: 24 wk Case 2: 25 wk	Case 1: Fetal extremities were short for gestational age. The femurs, humeri, and fibulas were bent, and some of which were telephone receiver-shaped Case 2: Fetal nasal bones were not shown. The thorax was narrow and bell-shaped. Fetal limb bones were short for gestational age, and some were abnormally bending. The fetus was aborted at 25 wk	1	Case 1: Umbilical cord blood Case 2: Aborted fetuses and amniotic fluid samples
Abbreviation: Ol, Os	Abbreviation: Ol, Osteogenesis imperfecta.					

rehabilitative training.<sup>19,20</sup> Therefore, regular prenatal checkups, especially prenatal ultrasound examinations, are paramount for such diseases. Prompt prenatal diagnosis is necessary for fetuses with a family history or ultrasound findings suggestive of skeletal dysplasia, for early prenatal intervention and prevention of the birth of affected infants.

# Conclusion

In conclusion, we present a case of OI in a fetus with skeletal dysplasia and generalized subcutaneous edema. The c.2174G > T/p.(G725V) de novo mutation of *COL1A1* in our patient was a novel pathogenic mutation of OI.

## **Ethics Approval**

The samples used in this study were collected with appropriate informed consent and approval from the Ethics Committee of Shandong Provincial Hospital, affiliated with Shandong First Medical University (LCYJ: NO. 2018-003).

## Authors' Contribution

Y.L. and Y.T. wrote the original draft of this manuscript. J.L. and Y.W. visualized the study. X.W. contributed to the conceptualization; methodology; project administration, writing, review, and editing.

## Patient Consent

Images and information from individual participants were obtained from the individual's free prior informed consent.

#### Data Availability

All the data involved in this study are available in the manuscript, tables, and figures.

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# **Conflict of Interest**

None declared.

#### Acknowledgment

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#### References

- <sup>1</sup> Savarirayan R, Rossiter JP, Hoover-Fong JE, et al; Skeletal Dysplasia Management Consortium. Best practice guidelines regarding prenatal evaluation and delivery of patients with skeletal dysplasia. Am J Obstet Gynecol 2018;219(06):545–562
- 2 Glorieux FH, Moffatt P. Osteogenesis imperfecta, an ever-expanding conundrum. J Bone Miner Res 2013;28(07):1519–1522
- 3 Forlino A, Marini JC. Osteogenesis imperfecta. Lancet 2016;387 (10028):1657–1671
- 4 Marini JC, Forlino A, Bächinger HP, et al. Osteogenesis imperfecta. Nat Rev Dis Primers 2017;3:17052
- 5 Sillence DO, Senn A, Danks DM. Genetic heterogeneity in osteogenesis imperfecta. J Med Genet 1979;16(02):101–116

Table 3 (Continued)

- 6 Byers PH, Krakow D, Nunes ME, Pepin MAmerican college of medical genetics. Genetic evaluation of suspected osteogenesis imperfecta (OI). Genet Med 2006;8(06):383–388
- 7 Sykes B, Ogilvie D, Wordsworth P, Anderson, Jones N. Osteogenesis imperfecta is linked to both type I collagen structural genes. Lancet 1986;2(8498):69–72
- 8 Steiner RD, Basel D. COL1A1/2 Osteogenesis imperfecta. In: Adam MP, Everman DB, Mirzaa GM, et al., eds. GeneReviews®. Seattle (WA): University of Washington, Seattle; 2005
- 9 Pollitt R, McMahon R, Nunn J, et al. Mutation analysis of COL1A1 and COL1A2 in patients diagnosed with osteogenesis imperfecta type I-IV. Hum Mutat 2006;27(07):716
- 10 Gauba V, Hartgerink JD. Synthetic collagen heterotrimers: structural mimics of wild-type and mutant collagen type I. J Am Chem Soc 2008;130(23):7509–7515
- 11 Xi L, Zhang H, Zhang ZL. Clinical and genetic analysis in 185 Chinese probands of osteogenesis imperfecta. J Bone Miner Metab 2021;39(03):416–422
- 12 Maioli M, Gnoli M, Boarini M, et al. Genotype-phenotype correlation study in 364 osteogenesis imperfecta Italian patients. Eur J Hum Genet 2019;27(07):1090–1100
- 13 Augusciak-Duma A, Witecka J, Sieron AL, et al. Mutations in the COL1A1 and COL1A2 genes associated with osteogenesis imperfecta (OI) types I or III. Acta Biochim Pol 2018;65(01):79–86
- 14 Marini JC, Forlino A, Cabral WA, et al. Consortium for osteogenesis imperfecta mutations in the helical domain of type I collagen: regions rich in lethal mutations align with collagen binding sites for integrins and proteoglycans. Hum Mutat 2007;28(03):209–221
- 15 Mei Y, Zhang H, Zhang Z. Comparing clinical and genetic characteristics of *de novo* and inherited *COL1A1/COL1A2* variants in a large Chinese cohort of osteogenesis imperfecta. Front Endocrinol (Lausanne) 2022;13:935905
- 16 Zhytnik L, Maasalu K, Duy BH, et al. De novo and inherited pathogenic variants in collagen-related osteogenesis imperfecta. Mol Genet Genomic Med 2019;7(03):e559
- 17 Milks KS, Hill LM, Hosseinzadeh K. Evaluating skeletal dysplasias on prenatal ultrasound: an emphasis on predicting lethality. Pediatr Radiol 2017;47(02):134–145

- 18 Kucińska-Chahwan A, Roszkowski T, Nowakowska B, et al. Extended genetic testing in fetuses with sonographic skeletal system abnormalities. Ultrasound Obstet Gynecol 2022;59(05): 660–667
- 19 Morello R. Osteogenesis imperfecta and therapeutics. Matrix Biol 2018;71-72:294–312
- 20 Tournis S, Dede AD. Osteogenesis imperfecta a clinical update. Metabolism 2018;80:27-37
- 21 Ji Y, Ikram A, Ma Z, Peppelenbosch MP, Pan Q. Co-occurrence of heterozygous mutations in COL1A1 and SERPINF1 in a high-risk pregnancy complicated by osteogenesis imperfecta. J Genet 2019; 98(02):51
- 22 Tanner L, Vainio P, Sandell M, Laine J. Novel COL1A1 mutation c.3290G>T associated with severe form of osteogenesis imperfecta in a fetus. Pediatr Dev Pathol 2017;20(05):455–459
- 23 Huang H, Liu J, Zhang G. A novel de novo mutation in COL1A1 leading to osteogenesis imperfecta confirmed by zebrafish model. Clin Chim Acta 2021;517:133–138
- 24 Zhang L, Pan L, Teng Y, Liang D, Li Z, Wu L. Molecular diagnosis for 55 fetuses with skeletal dysplasias by whole-exome sequencing: a retrospective cohort study. Clin Genet 2021;100(02): 219–226
- 25 Yang K, Liu Y, Wu J, et al. Prenatal cases reflect the complexity of the *COL1A1/2* associated osteogenesis imperfecta. Genes (Basel) 2022;13(09):2
- 26 Zhuang J, Chen C, Chen Y, et al. Identification of a rare variant of c. 1777G>A (p.G593S) in the COL1A1 gene as the etiology of recurrent osteogenesis imperfecta by whole-exome sequencing. Front Pediatr 2022;10:816090
- 27 Cao J, Chen A, Tian L, Yan L, Li H, Zhou B. Application of whole exome sequencing in fetal cases with skeletal abnormalities. Heliyon 2022;8(07):e09819
- 28 Li R, Wang J, Wang L, Lu Y, Wang C. Two novel mutations of COL1A1 in fetal genetic skeletal dysplasia of Chinese. Mol Genet Genomic Med 2020;8(03):e1105
- 29 Yang Y, Wang M, Wang H. Prenatal trio-based whole exome sequencing in fetuses with abnormalities of the skeletal system. Mol Genet Genomics 2022;297(04):1017–1026