

Chemoenzymatic Total Synthesis of (+)-Oxycodone from Phenethyl Acetate

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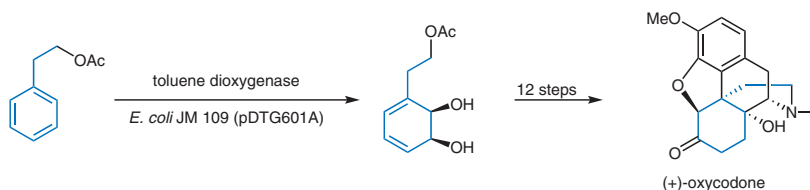
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Abstract The stereoselective total synthesis of unnatural (+)-oxycodone from phenethyl acetate is described. Absolute stereochemistry was established via microbial dihydroxylation of phenethyl acetate with the recombinant strain JM109 (pDTG601A) to the corresponding *cis*-cyclohexadienediol whose configuration provides for the absolute stereochemistry of the ring C of (+)-oxycodone. Intramolecular Heck cyclization was employed to establish the quaternary carbon at C-13, along with the dibenzodihydrofuran functionality. The C-14 hydroxyl was installed via SmI_2 -mediated radical cyclization. The synthesis of (+)-oxycodone was completed in a total of 13 steps and an overall yield of 1.5%. Experimental and spectral data are provided for all new compounds.

Key words enzymatic dihydroxylation, total synthesis, oxycodone, Parker's hydroamination, pinacol-type coupling

The semi-synthetic opioid (–)-oxycodone (**1**) (Figure 1), although found also in nature,¹ is a potent analgesic that is clinically prescribed for pain management.² It is taken by mouth and is available mixed with acetaminophen in immediate release tablet form, which contains oxycodone HCl (5 mg) and acetaminophen (325 mg) (Percocet®),³ as a single ingredient medication oxycodone HCl (60 mg and 80 mg) or as film coated, extended release tablet OxyContin®.⁴ The commercial route⁵ for the preparation of oxycodone is a two-step process from thebaine through the oxidation of the diene moiety with a peroxy acid to form an enone followed by hydrogenation.⁶ Thebaine is a minor constituent of opium and thus this fact limits the production of oxycodone. However, thebaine, as well as oripavine, are now also available from genetically modified poppies that produce much higher percentages of these alkaloids.⁷ These compounds are now supplied by Tasmanian Alkaloids, Inc.⁸

During the past 70 years, since the milestone synthesis of morphine by Gates,⁹ there have been more than 30 total syntheses of morphine and related alkaloids and the academic effort continues unabated.¹⁰ Even the most efficient synthesis reported by Rice¹¹ may not be suitable for scale-up in the industrial preparation of morphinans. Although the development of a truly practical total synthesis of any morphinan or an opiate-derived agent on a commercial scale seems like a distant dream we have attempted to design a method for the synthesis of oxycodone from readily available starting materials. A *de novo* preparation of oxycodone or any other medicinal opiate-derived agents for medicinal use may serve as an insurance against any future unforeseen events that may limit the supply of natural sources because of climate or political instabilities in the opium-producing regions. To date, there is only one published total synthesis of oxycodone.¹² Fukuyama and co-workers accomplished the total synthesis of (–)-oxycodone (**1**) from 2-bromoisovanillin in 24 steps in an overall yield of 0.016%. The key steps in their synthesis featured a direct intramolecular arylation of an aryl bromide, an oxidative dearomatization reaction, an intramolecular Michael addition, and a Hofmann rearrangement. Absolute stereochemistry was incorporated into the starting material by the use of Evans' oxazolidinone as a chiral auxiliary.

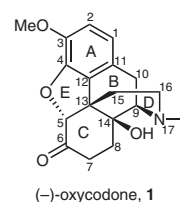


Figure 1 Structure of (–)-oxycodone (with numbering system shown)

Biographical sketches



Mary Ann A. Endoma-Arias was born in 1969 in Rizal, Philippines. After completing her BSc (1990), cum laude, at the University of the Philippines Diliman, in Quezon City, Philippines, she started her PhD studies at Virginia Tech, in Blacksburg, VA, under the supervision of Professor Tomas Hudlicky, in 1992. She moved with Professor Hudlicky in 1995 to the University of Florida, in Gainesville, FL, where she completed her

PhD in 1997. Upon completion of her PhD, she returned to the University of the Philippines where she rose to the rank of Associate Professor. Her main interest during her research career in the Philippines focused on the synthesis and conjugation of compounds with medicinal properties to liposomes and hydrophilic polymers for targeted delivery. During this time she continued to work closely with the

Hudlicky Research Group. Since 2010 she has been working with the Research group of Professor Hudlicky at Brock University in St. Catharines, ON, Canada as a Research Associate. In 2015 she emigrated to Canada with her husband and children. Her main research interest is the total synthesis of morphine and related alkaloids. To date, she has published 30 papers and 5 patents.



Mariia Makarova was born in 1990 in Samara, Russia. She completed her B.Sc. in 2013 at Samara State University, Samara, Russia. In January 2014 she started her M.Sc. stud-

ies at Brock University, St. Catharines, Canada under the supervision of Professor Tomas Hudlicky. During her second year of M.Sc. she transferred to the Ph.D. program.

Her Ph.D. research has been focused on the chemoenzymatic approach towards synthesis of morphine alkaloids. Her defense is scheduled on November 30, 2018.



Helen E. Dela Paz was born in 1975 in Rizal, Philippines. She obtained her B.Sc. in Food Technology at the University of the Philippines Diliman, in Quezon City, Philippines. She was given the opportunity to apply her knowledge of microbiology to fermentation technology. Thus, in 2001, she was offered a research position in the laboratories of Dr. Tomas Hudlicky at the University of

Florida, Gainesville, FL, USA. While at UF, she was introduced to the utility of a fermentor in the large scale microbial oxidation of aromatic compounds towards the production of value-added metabolites. After almost two years of work, she returned to the Philippines and took a job as a Science teacher. In 2014, she returned to the Hudlicky Research Group at Brock University to work as

a Researcher and join her sister who has been working there. Her research interest focuses on the production of known diene diols from aromatic compounds, and the identification of new ones. She has published a few papers related to biocatalysis and its application to organic synthesis.



Tomas Hudlicky was born in 1949 in Prague, Czechoslovakia, where he received his elementary and middle school education. After several years of working as a process chemist apprentice and in other odd jobs in pharmaceutical chemistry, it became apparent that higher education opportunities were closed to him. In 1968, he emigrated to the U.S. with his parents and sister. Hudlicky's educational experience continued at Blacksburg High School, from which he dropped out in the spring of 1969. Accepted as a probational student at Virginia Tech the following autumn, he received his B.S. in chemistry in 1973, and went on to pursue graduate studies at Rice University under the direction of Professor Ernest Wenkert in the field of indole alkaloid total syn-

thesis, earning his Ph.D. in 1977. He then spent a year at the University of Geneva working under the late Professor Wolfgang Oppolzer on the synthesis of isocomene. In 1978, he joined the faculty at the Illinois Institute of Technology as an Assistant Professor, and began the first phase of his research career in the field of general methods of synthesis for triquinane terpenes and other natural products containing five-membered rings by [4+1] cyclopentene, pyrroline, and dihydrofuran annulation methodologies. He returned to his alma mater, Virginia Tech, in 1982, and rose to the rank of Professor in 1988. One year later, at the 20-year class reunion of the Blacksburg High School class of 1969, he received his High School Diploma. The next phase of his research involved

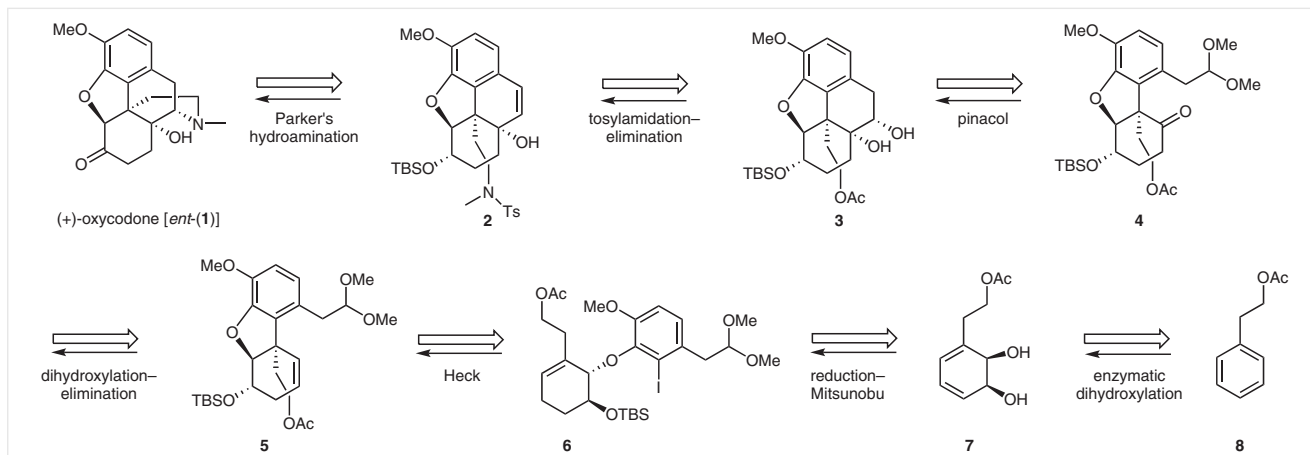
the investigation of *cis*-cyclohexadienediols in enantioselective synthesis. In 1995, he moved to University of Florida in Gainesville. In 2003, Dr. Hudlicky accepted an offer from Brock University where he currently holds a position as Canada Research Chair. His current research interests include the development of enantioselective synthetic methods, bacterial dioxygenase-mediated degradation of aromatics, design and synthesis of fluorinated inhalation anesthetic agents, synthesis of morphine and Amaryllidaceae alkaloids, and design of unnatural oligosaccharide conjugates with new molecular properties. His hobbies are skiing, hockey, martial arts, and music.

Scheme 1 outlines our retrosynthesis of (+)-oxycodone. Disconnection of ring D leads to styrene **2**. In the forward sense, Parker's hydroamination can be utilized to construct the C-9 stereogenic center. The tosylamide group necessary for the cyclization reaction is derived from acetate **3**, which is envisioned to be prepared from the keto acetal **4** following a deprotection of acetal and a SmI_2 -mediated pinacol-type coupling reaction. The key intermediate **4** can be obtained from alkene **5** via dihydroxylation followed by selective mesylation of the less hindered hydroxyl group and the elimination of the mesylate to reveal the ketone functionality in **4**. Alkene **5** can be obtained in two steps from alcohol **7** via a sequence of steps that involves a Mitsunobu coupling with an iodophenol acetal to furnish aryl ether **6** followed by an intramolecular Heck reaction. The absolute stereochemistry in **7** is incorporated via microbial dihydroxylation with toluene dioxygenase, overexpressed in *E. coli* JM109 (pDTG601A), in the whole-cell fermentation of phenethyl acetate (**8**).¹³ The enzymatically derived arene *cis*-dihydrodiols such as **7** have found widespread use in enantioselective synthesis of natural products.¹⁴

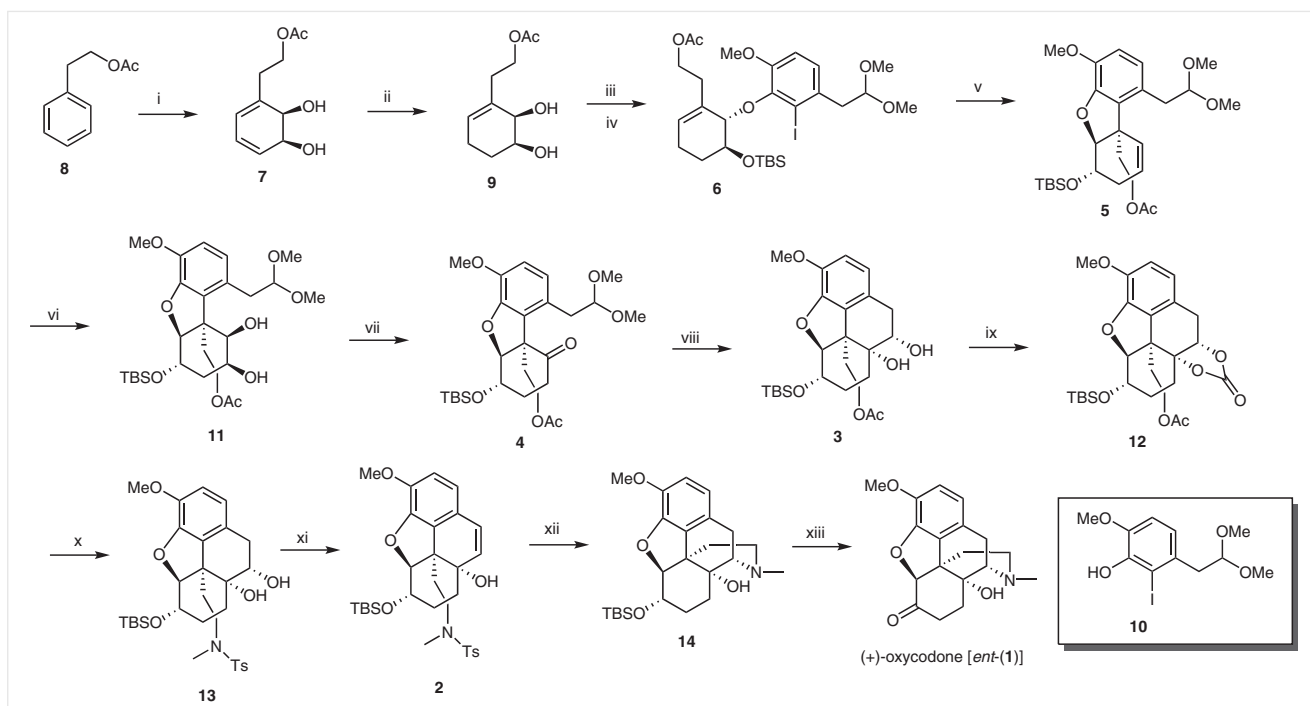
The synthesis began with the microbial dihydroxylation of phenethyl acetate (**8**) (Scheme 2) in a whole cell fermentation with *E. coli* JM109 (pDTG601A) to afford the intermediate cyclohexadiene diol **7** (obtained in 5 gL^{-1} yield),¹⁵ which was subjected to a selective reduction of the less hindered alkene to afford the known diol **9** (85% yield). The distal, less hindered, hydroxyl in diol **9** was protected with *tert*-butyldimethylsilyl chloride and the proximal allylic alcohol was then coupled with iodophenol **10**,¹⁷ derived from isovanillin, via a Mitsunobu reaction to furnish ether **6** (45% yield over two steps). A subsequent intramolecular Heck reaction of **6** produced olefin **5** (87% yield) whose dihydroxylation led to diol **11** (81% yield). This compound possesses the features of the ACE rings of oxycodone. The diol functionality was converted to ketone **4** via mesylation of the less hindered hydroxyl group followed by DBU-catalyzed

elimination of the resulting mesylate (63% yield over two steps). With the attainment of **4**, deprotection of the acetal followed by a pinacol-type coupling of the intermediate keto aldehyde using SmI_2 was conducted to afford diol **3**, tentatively assigned as the *cis*-isomer (65% yield over two steps).¹⁸ Protection of diol **3** with a carbonate group to afford **12** (80% yield) allowed for the introduction of the tosylamide functionality via methanolysis of the acetate followed by Mitsunobu coupling of the resulting alcohol with *N*-methyl *p*-toluenesulfonyl amide. The carbonate moiety in the crude tosylamide was hydrolyzed to afford diol **13** (67% yield over three steps). It would have been desirable to isolate the intermediate tosylamide carbonate but the difficulty in the separation of residual *N*-methyl *p*-toluenesulfonyl amide from the desired product rendered this process impractical. The less hindered hydroxyl of **13** was converted to the corresponding mesylate and subjected to DBU-catalyzed elimination affording alkene **2** (70% yield over two steps), which was the precursor for the key Parker's hydroamination step to complete the ring system of oxycodone. Thus, treatment of **2** with Li in liquid ammonia as reported by Parker¹⁹ in the total synthesis of hydrocodone afforded the oxycodol ether **14** (76% yield).²⁰ Deprotection of the TBS group in **14** followed by oxidation of the alcohol to ketone afforded *ent*-oxycodone [*ent*-(**1**)] (59% yield over two steps).

In conclusion, a short chemoenzymatic synthesis of *ent*-oxycodone has been accomplished in 13 steps from phenethyl acetate. Further improvements in this short synthesis will address installation of the *N*-methyltosylamide side chain earlier in the synthesis thus eliminating the need to use a carbonate protecting group during the synthetic sequence. In particular, the acetate functionality in **5** will be converted to the tosylamide. Furthermore, an SmI_2 -mediated nitron-keto coupling is being investigated to afford an amino alcohol instead of diol intermediate for subsequent cyclization to the pendant acetate side chain. These



Scheme 1 Retrosynthetic analysis of the route to *ent*-oxycodone



Scheme 2 Synthesis of *ent*-oxycodone. *Reagents and conditions:* i) *E. coli* JM 109 (pDTG601A), 5 gL⁻¹; ii) potassium azodicarboxylate, MeOH, AcOH, 85%; iii) TBSCl, imidazole, CH₂Cl₂, 54%; iv) **10**, TMAD, *n*-Bu₃P, THF, 0 °C to r.t., 83%; v) Ag₂CO₃, dppp, Pd(OAc)₂, DMF, reflux, 87%; vi) cat. OsO₄, NMO, acetone/H₂O, 81%; vii) a. MsCl, NEt₃, CH₂Cl₂, b. DBU, toluene, reflux, 63% over 2 steps; viii) a. aq TFA, toluene, 50 °C, b. Sml₂, THF, -78 °C, 65% over 2 steps; ix) carbonyldiimidazole, toluene, 80 °C, 80%; x) a. K₂CO₃, MeOH, b. TsNHMe, TMAD, *n*-Bu₃P, THF, c. aq NaOH, MeOH, 67% over 3 steps; xi) a. MsCl, NEt₃, CH₂Cl₂, b. DBU, toluene, reflux, 70% over 2 steps; xii) Li, *t*-BuOH, THF, liq NH₃, -78 °C, 76%; xiii) a. TBAF, THF, b. Dess–Martin periodinane, CH₂Cl₂, 59% yield over 2 steps.

improvements, as well as the 2nd generation synthesis of the natural enantiomer, are currently being investigated and will be reported in due course.

Inoculum was obtained from viable cells stored at -78 °C in cryovials. They were grown in suitable media as previously described.^{15b} Substrate was fed in 5 mL increments over the course of 3 h with metabolites being harvested in the usual manner. All non-aqueous reactions were conducted in an argon atmosphere using standard Schlenk techniques for the exclusion of moisture and air. CH₂Cl₂ was distilled from CaH₂; THF and toluene were dried over Na/benzophenone. Analytical TLC was performed on Silicycle 60 Å 250 mm TLC plates with F-254 indicator. Flash column chromatography was performed using silica gel 60 (230–400 mesh). Melting points were recorded on a Hoover Unimelt apparatus and are uncorrected. IR spectra were obtained on a PerkinElmer One FT-IR spectrophotometer. Optical rotation was measured on a PerkinElmer 341 polarimeter at a wavelength of 589 nm. ¹H and ¹³C spectra were recorded on a 300 MHz and 400 MHz Bruker spectrometer. All chemical shifts are referenced to TMS or residual nondeuterated solvent. Data of proton spectra are reported as follows: chemical shift in ppm [multiplicity (standard abbreviations), coupling constants (Hz), integration]. ¹³C NMR spectra were recorded with complete proton decoupling and the chemical shifts are reported in ppm (δ) relative to solvent resonance as internal standard. Mass spectra and high-resolution mass spectra were performed by the Analytical Division at Brock University.

2-[(5*S*,6*R*)-5,6-Dihydroxycyclohexa-1,3-dien-1-yl]ethyl Acetate (**7**)¹⁶

This compound was prepared according to a literature procedure.¹⁶ [α]_D²⁰ +32.07 (*c* = 0.4, CHCl₃) {Lit.¹⁶ [α]_D²⁸ +40.7 (*c* = 2.0, CHCl₃)}; *R*_f = 0.18 (hexanes/EtOAc 1:2).

IR (CHCl₃): 3392, 2955, 2925, 1735, 1383, 1366, 1238, 1037, 803 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 5.90 (ddd, *J* = 9.5, 5.2, 0.9 Hz, 1 H), 5.80 (dd, *J* = 9.3, 3.4 Hz, 1 H), 5.75–5.70 (m, 1 H), 4.30–4.14 (m, 3 H), 4.09 (d, *J* = 6.0 Hz, 1 H), 2.97 (s, 2 H), 2.57–2.48 (m, 2 H), 2.01 (s, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 171.5, 137.7, 128.2, 124.7, 121.6, 70.1, 68.5, 63.1, 33.3, 21.1.

MS (EI): *m/z* = 198, 120, 107, 91, 75.

HRMS (EI): *m/z* calcd for C₁₀H₁₄O₄: 198.0892; found: 198.0889.

2-[(5*S*,6*R*)-5,6-Dihydroxycyclohex-1-enyl]ethyl Acetate (**9**)¹⁷

To a stirred mixture of diol **7** (150 mg, 0.76 mmol) and potassium azodicarboxylate (PAD) (200 mg, 1.03 mmol) in MeOH (4 mL) at 0 °C was added dropwise (over 15 min) AcOH (0.5 mL, 8.75 mmol) in MeOH (1 mL). The reaction was complete after 30 min, as monitored by TLC (2:1 EtOAc/hexanes). The mixture was concentrated via rotary evaporation to afford a crude product that was resuspended in CH₂Cl₂ (10 mL), and the CH₂Cl₂ layer was washed with sat. aq NaHCO₃ (5 mL). The organic extract was dried (MgSO₄), filtered, and evaporated to remove solvent to afford a crude oily residue that was chromatographed

on silica gel using 2:1 EtOAc/hexanes as eluent to afford **9** as an oil; yield: 128 mg (85%); $[\alpha]_D^{20}$ -65.9 ($c = 1.0$, CHCl_3) {Lit.¹⁷ $[\alpha]_D^{20}$ -53.0 ($c = 0.2$, CHCl_3)}; $R_f = 0.25$ (hexanes/EtOAc 1:2).

¹H NMR (300 MHz, CDCl_3): $\delta = 5.63$ (t, $J = 3.7$ Hz, 1 H), 4.37–4.26 (m, 1 H), 4.22–4.12 (m, 1 H), 4.03 (d, $J = 3.8$ Hz, 1 H), 3.79–3.71 (m, 1 H), 2.57–2.33 (m, 3 H), 2.23–2.05 (m, 3 H), 2.04 (s, 3 H), 1.75–1.62 (m, 2 H).

¹³C NMR (75 MHz, CDCl_3): $\delta = 171.4$, 133.6, 127.6, 69.5, 68.6, 63.3, 33.9, 24.9, 24.1, 20.9.

2-((5S,6S)-5-[(*tert*-Butyldimethylsilyloxy)-6-[3-(2,2-dimethoxyethyl)-2-iodo-6-methoxyphenoxy]cyclohex-1-en-1-yl]ethyl Acetate (**6**)

To a stirred solution of alcohol **9** (3.1 g, 10.0 mmol) and 3-(2,2-dimethoxyethyl)-2-iodo-6-methoxyphenol¹⁵ (**10**; 3.7 g, 10.9 mmol) at -10 °C in THF (40 mL) was added $n\text{Bu}_3\text{P}$ (3.5 mL, 14.0 mmol), followed by TMAD (2.3 g, 13.0 mmol). The reaction mixture was allowed to warm up to r.t. and stirred for 16 h at r.t. The solvent was removed by rotary evaporation and the residue was chromatographed on silica gel using hexanes/EtOAc as eluent (4:1 to 2:1) to afford the product **6** as an oil; yield: 5.2 g (83%); $[\alpha]_D^{20}$ $+48.4$ ($c = 1.2$, CH_2Cl_2); $R_f = 0.58$ (2:1 hexanes/EtOAc).

IR (film): 2930, 2855, 1738, 1642, 1472, 1249, 1078, 776 cm^{-1} .

¹H NMR (300 MHz, CDCl_3): $\delta = 7.01$ (d, $J = 8.4$ Hz, 1 H), 6.86 (d, $J = 8.4$ Hz, 1 H), 5.87 (d, $J = 4.5$ Hz, 1 H), 4.58 (s, 1 H), 4.56 (t, $J = 5.7$ Hz, 1 H), 4.21 (m, 2 H), 4.05 (m, 1 H), 3.87 (s, 3 H), 3.36 (s, 6 H), 3.09 (d, $J = 5.7$ Hz, 2 H), 2.47–2.61 (m, 2 H), 2.28–2.37 (m, 2 H), 2.04 (s, 3 H), 2.00–2.06 (m, 1 H), 1.61–1.70 (m, 1 H), 0.77 (s, 9 H), -0.13 (s, 3 H), -0.18 (s, 3 H).

¹³C NMR (75 MHz, CDCl_3): $\delta = 171.1$, 150.2, 146.8, 132.9, 130.4, 130.0, 125.8, 112.1, 104.5, 79.3, 67.5, 63.7, 55.5, 54.2, 54.1, 44.4, 33.7, 25.6, 21.0, 20.7, 17.9, -5.1 , -5.2 .

MS (EI+): m/z (%) = 657 (100), 565 (10), 297 (10), 237 (25).

HRMS (EI+): m/z calcd for $\text{C}_{27}\text{H}_{41}\text{IO}_7\text{Si}$ [$M - 2\text{H}$]: 632.1666; found: 632.1657.

2-((5aS,6S,9aR)-6-[(*tert*-Butyldimethylsilyloxy)-1-(2,2-dimethoxyethyl)-4-methoxy-5a,6,7,9a-tetrahydrodibenzo[*b,d*]furan-9a-yl]ethyl Acetate (**5**)^{16a}

To a stirred solution of ether **6** (3.02 g, 4.76 mmol) in DMF (55 mL) was added $\text{Pd}(\text{OAc})_2$ (168 mg, 0.71 mmol), Ag_2CO_3 (3.9 g, 14.3 mmol), and dppp (590 mg, 1.43 mmol). The resulting mixture was heated to reflux for 3 h. The cooled reaction mixture was diluted with $\text{Et}_2\text{O}/\text{H}_2\text{O}$ (100 mL/50 mL). The layers were separated and the aqueous phase was further extracted with Et_2O (2 \times 50 mL). The combined organic extracts were washed with brine, dried (MgSO_4), filtered, and concentrated to afford a residue that was chromatographed on silica gel using hexanes/EtOAc (8:1 to 4:1) as eluent to afford the product **5** as an oil; yield: 2.1 g (87%).

2-((5aS,6S,8S,9R,9aS)-6-[(*tert*-Butyldimethylsilyloxy)-1-(2,2-dimethoxyethyl)-8,9-dihydroxy-4-methoxy-5a,6,7,8,9,9a-hexahydrodibenzo[*b,d*]furan-9a-yl]ethyl Acetate (**11**)

To a stirred solution of alkene **5** (1.5 g, 2.95 mmol) in acetone/ H_2O (30 mL/9 mL) was added *N*-methylmorpholine oxide (NMO; 345 mg, 2.95 mmol) followed by a catalytic amount of $\text{K}_2\text{OsO}_4 \cdot 2\text{H}_2\text{O}$. The resulting solution was stirred at r.t. for 2 d whereupon the mixture was diluted with EtOAc/ H_2O (30 mL/15 mL). The layers were separated and the aqueous phase was further extracted with EtOAc (2 \times 20 mL). The

combined organic extracts were dried (MgSO_4), filtered, and concentrated to afford a residue that was chromatographed on silica gel using hexanes/EtOAc (1:1 to 1:2) as eluent to afford the product **11** as an oil; yield: 1.3 g (81%); $[\alpha]_D^{20}$ $+16.6$ ($c = 16.7$, CH_2Cl_2); $R_f = 0.36$ (1:1 hexanes/EtOAc).

IR (film): 3429, 2951, 2856, 1738, 1629, 1433, 1042 cm^{-1} .

¹H NMR (300 MHz, CDCl_3): $\delta = 6.72$ (d, $J = 8.4$ Hz, 1 H), 6.64 (d, $J = 8.4$ Hz, 1 H), 4.57 (dd, $J = 3.3$, 7.8 Hz, 1 H), 4.50 (dd, $J = 3.6$, 3.6 Hz, 1 H), 3.90–3.98 (m, 2 H), 3.80 (s, 3 H), 3.37 (s, 3 H), 3.33 (d, $J = 3.3$ Hz, 1 H), 3.25 (s, 3 H), 2.84–2.90 (m, 2 H), 2.22–2.30 (m, 2 H), 2.03 (td, $J = 3.3$, 13.2 Hz, 1 H), 1.78 (dt, $J = 4.2$, 13.2 Hz, 1 H), 0.85 (s, 9 H), 0.08 (s, 3 H), 0.06 (s, 3 H).

¹³C NMR (75 MHz, CDCl_3): $\delta = 170.7$, 148.3, 143.3, 128.8, 125.1, 121.8, 112.0, 106.2, 86.2, 76.7, 75.1, 68.8, 64.0, 61.4, 55.8, 55.7, 53.4, 52.6, 34.5, 33.0, 32.1, 25.7, 20.8, 17.9, -5.09 , -5.09 .

MS (EI+): m/z (%) = 483 (30), 451 (25), 359 (50), 289 (100), 259 (30), 167 (50), 149 (60), 121 (40).

HRMS (EI+): m/z calcd for $\text{C}_{27}\text{H}_{44}\text{O}_4\text{Si}$: 540.2755; found: 540.2734.

2-((5aS,6S,9aR)-6-[(*tert*-Butyldimethylsilyloxy)-1-(2,2-dimethoxyethyl)-4-methoxy-9-oxo-5a,6,7,8,9,9a-hexahydrodibenzo[*b,d*]furan-9a-yl]ethyl Acetate (**4**)

To a stirred solution of diol **11** (1.41 g, 2.61 mmol) in CH_2Cl_2 (37 mL) at 0 °C was added NEt_3 (724 μL , 5.22 mmol) followed by MsCl (303 μL , 3.91 mmol). The resulting solution was stirred at 0 °C for 30 min, and then diluted with $\text{CH}_2\text{Cl}_2/\text{aq NaHCO}_3$ (20 mL/30 mL). The layers were separated and the aqueous layer was further extracted with CH_2Cl_2 (2 \times 15 mL). The organic layers were combined, dried (MgSO_4), filtered, and concentrated to afford a residue that was used without purification in the next step. To the crude mesylate (obtained from the previous step) dissolved in toluene (20 mL) was added DBU (5 mL). The mixture was heated to reflux for 5 h. The reaction mixture was allowed to cool to r.t. after which it was diluted with EtOAc/sat. aq NH_4Cl (20 mL/20 mL). The layers were separated and the aqueous layer was further extracted with EtOAc (2 \times 20 mL). The organic layers were combined, dried (MgSO_4), filtered, and concentrated to afford a residue that was chromatographed on silica gel using hexanes/EtOAc as eluent (2:1) to afford the product **4** as an oil; yield: 850 mg (63% over 2 steps); $[\alpha]_D^{20}$ -60.2 ($c = 17.9$, CH_2Cl_2); $R_f = 0.44$ (2:1 hexanes/EtOAc).

IR (film): 2952, 2857, 1739, 1713, 1625, 1506, 1233, 1075, 836 cm^{-1} .

¹H NMR (300 MHz, CDCl_3): $\delta = 6.78$ (d, $J = 8.4$ Hz, 1 H), 6.74 (d, $J = 8.4$ Hz, 1 H), 4.81 (dd, $J = 2.4$, 3.0 Hz, 1 H), 4.43 (t, $J = 5.4$ Hz, 1 H), 4.18 (t, $J = 3.3$ Hz, 1 H), 4.01–4.05 (m, 1 H), 3.82–3.89 (m, 1 H), 3.85 (s, 3 H), 3.27 (s, 6 H), 2.65 (d, $J = 5.4$ Hz, 2 H), 2.39–2.58 (m, 2 H), 2.24–2.29 (m, 1 H), 2.03–2.15 (m, 2 H), 1.93 (s, 3 H), 1.77–1.86 (m, 1 H), 0.85 (s, 9 H), 0.08 (s, 3 H), 0.06 (s, 3 H).

¹³C NMR (75 MHz, CDCl_3): $\delta = 207.8$, 170.7, 148.9, 142.9, 127.1, 126.0, 123.0, 112.8, 104.7, 91.7, 67.1, 61.0, 59.2, 55.9, 53.4, 53.1, 34.3, 33.2, 32.7, 25.5, 24.4, 20.8, 17.9, -4.9 , -5.1 .

MS (EI+): m/z (%) = 522 (20), 490 (100), 430 (40), 373 (50), 341 (95), 313 (50), 199 (75), 111 (38).

HRMS (EI+): m/z calcd for $\text{C}_{27}\text{H}_{42}\text{O}_8\text{Si}$: 522.2649; found: 522.2622.

2-((3S,3aS,3a1R,9S,9aR)-3-[(*tert*-Butyldimethylsilyloxy)-9,9a-dihydroxy-5-methoxy-1,2,3,3a,3a1,8,9,9a-octahydrophenanthro[4,5-*bcd*]furan-3a1-yl]ethyl Acetate (**3**)

To a stirred solution of keto acetal **4** (440 mg, 0.84 mmol) in toluene (10 mL) was added 50% aq TFA (1.0 mL). The biphasic mixture was heated to 50 °C for 30 min, then it was allowed to cool down to r.t. The

reaction mixture was diluted with EtOAc/sat. aq NaHCO₃ (10 mL/5 mL). The layers were separated and the aqueous phase was further extracted with EtOAc (2 × 10 mL). The organic layers were combined, dried (MgSO₄), filtered, and concentrated to afford a residue that was used as crude in the next step. To the crude keto acetal (obtained from the previous step) dissolved in THF (3 mL) at -78 °C was added SmI₂ (14 mL, 0.1 M in THF, 1.40 mmol). The resulting deep blue reaction mixture was stirred at -78 °C for 30 min. The cooling bath was removed and the mixture was diluted with EtOAc/sat. aq NaHCO₃ (20 mL/15 mL). The biphasic mixture was allowed to warm up to r.t., then the layers were separated. The aqueous layer was further extracted with EtOAc (3 × 20 mL). The organic layers were combined, dried (MgSO₄), filtered, and concentrated to afford a residue that was chromatographed on silica gel using hexanes/EtOAc as eluent (2:1) to afford the product **3** as an oil; yield: 262 mg (65% yield over 2 steps); [α]_D²⁰ +41.5 (c = 0.9, CH₂Cl₂); R_f = 0.33 (1:1 hexanes/EtOAc).

IR (film): 3435, 2952, 2930, 2855, 1736, 1713, 1632, 1504, 1257, 1030 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 6.72 (d, J = 8.1 Hz, 1 H), 6.61 (d, J = 8.1 Hz, 1 H), 4.80 (dd, J = 3.3, 1.2 Hz, 1 H), 4.18–4.27 (m, 1 H), 3.95–4.05 (m, 1 H), 3.97–4.03 (m, 1 H), 3.88–3.91 (m, 1 H), 3.86 (s, 3 H), 3.28 (dd, J = 8.1, 17.0 Hz, 1 H), 3.02 (dd, J = 3.0, 17.0 Hz, 1 H), 2.47–2.55 (m, 1 H), 2.21–2.31 (m, 1 H), 1.96 (s, 3 H), 1.65–1.71 (m, 1 H), 1.56–1.60 (m, 1 H), 1.30–1.36 (m, 1 H), 1.05–1.13 (m, 1 H), 0.96 (s, 9 H), 0.21 (s, 3 H), 0.16 (s, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 170.0, 145.8, 142.4, 130.2, 124.8, 119.8, 113.7, 91.6, 74.5, 73.6, 69.4, 61.8, 56.4, 49.8, 33.9, 33.8, 31.7, 25.7, 22.7, 20.9, 18.0, -5.0, -5.2.

MS (EI+): m/z (%) = 361 (90), 343 (100), 315 (60), 305 (95), 287 (40), 249 (80).

HRMS (EI+): m/z calcd for C₂₅H₃₈O₇Si: 478.2387; found: 478.2391.

2-((5a*S*,8a*R*,8a1*R*,11*S*,11a*S*)-11-[(*tert*-Butyldimethylsilyl)oxy]-2-methoxy-7-oxo-5a,8a1,9,10,11,11a-hexahydro-5*H*-furo[2',3',4',5':4,5]-phenanthro[8a,9-d][1,3]dioxol-8a-1-yl)ethyl Acetate (**12**)

To a stirred solution of diol **3** (100 mg, 0.21 mmol) in toluene (3 mL) was added carbonyldiimidazole (102 mg, 0.63 mmol). The resulting mixture was heated to 80 °C for 3 h, then allowed to cool down to r.t. The solvent was removed by rotary evaporation and the residue was chromatographed on silica gel using hexanes/EtOAc (2:1) as eluent to afford the carbonate **12** as an oil; yield: 75 mg (71%); [α]_D²⁰ +51.6 (c = 1.0, CH₂Cl₂); R_f = 0.65 (1:1 hexanes/EtOAc).

IR (film): 3436, 2953, 2931, 2856, 1806, 1738, 1637, 1235, 1067 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 6.79 (d, J = 8.1 Hz, 1 H), 6.69 (d, J = 8.1 Hz, 1 H), 4.67 (d, J = 6.3 Hz, 1 H), 4.06–4.12 (m, 1 H), 3.86 (s, 3 H), 3.69–3.73 (m, 1 H), 3.39–3.43 (m, 1 H), 3.42 (dd, J = 8.1, 15.3 Hz, 1 H), 2.95 (dd, J = 8.1, 15.3 Hz, 1 H), 2.14–2.19 (m, 1 H), 2.00–2.11 (m, 1 H), 1.88 (s, 3 H), 1.69–1.84 (m, 2 H), 1.33–1.43 (m, 1 H), 0.90 (s, 9 H), 0.14 (s, 3 H), 0.05 (s, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 170.6, 145.5, 144.5, 127.4, 120.4, 120.3, 115.8, 96.7, 86.2, 82.4, 74.1, 60.3, 56.3, 50.0, 36.0, 33.0, 30.9, 26.4, 25.8, 25.6, 20.7, 18.0, -4.6, -5.0.

MS (EI+): m/z (%) = 447 (70), 419 (35), 387 (40), 343 (100), 313 (40), 269 (35), 117 (25).

HRMS (EI+): m/z calcd for C₂₆H₃₆O₈Si: 504.2179; found: 504.2172.

N-(2-((5a*S*,8a*R*,8a1*R*,11*S*,11a*S*)-11-[(*tert*-Butyldimethylsilyl)oxy]-2-methoxy-7-oxo-5a,8a1,9,10,11,11a-hexahydro-5*H*-furo[2',3',4',5':4,5]-phenanthro[8a,9-d][1,3]dioxol-8a-1-yl)ethyl)-*N*,4-dimethylbenzenesulfonamide (**13**)

To a stirred solution of carbonate **12** (50 mg, 0.10 mmol) in MeOH (2 mL) was added K₂CO₃ (50 mg, 0.36 mmol). The resulting suspension was stirred at r.t. for 2 h. The solvent was removed by rotary evaporation and the residue was diluted with EtOAc/sat. aq NaHCO₃ (10 mL/5 mL). The layers were separated and the aqueous phase was further extracted with EtOAc (2 × 10 mL). The organic layers were combined, dried (MgSO₄), filtered, and concentrated to afford a residue that was used as crude in the next step. To the crude alcohol (obtained from the previous step) and *N*-methyl *p*-toluenesulfonyl amide (20 mg, 0.11 mmol) in THF (1.5 mL) at -10 °C was added *n*Bu₃P (35 μL, 0.14 mmol), followed by TMAD (23 mg, 0.13 mmol). The reaction mixture was allowed to warm up to r.t. and stirred for 16 h at r.t. The solvent was removed by rotary evaporation to afford a residue that was used without purification in the next step. To the crude tosylamide carbonate (obtained from the previous step) dissolved in MeOH (2 mL) was added aq 3 N NaOH (0.5 mL). The resulting cloudy solution was stirred at r.t. for 30 min. The solvent was removed by rotary evaporation and the residue was diluted with EtOAc/H₂O (10 mL/5 mL). The aqueous layer was further extracted with EtOAc (2 × 10 mL). The organic layers were combined, dried (MgSO₄), filtered, and concentrated to afford a residue that was chromatographed on silica gel using hexanes/EtOAc (2:1) as eluent to afford the product **13** as an oil; yield: 40 mg (67% yield over 3 steps); [α]_D²⁰ +18.4 (c = 1.1, CH₂Cl₂); R_f = 0.50 (1:1 hexanes/EtOAc).

IR (film): 3466, 2928, 2855, 1633, 1505, 1159 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.61 (d, J = 8.4 Hz, 1 H), 7.27 (d, J = 8.4 Hz, 1 H), 6.73 (d, J = 8.1 Hz, 1 H), 6.61 (d, J = 8.1 Hz, 1 H), 4.75 (s, 1 H), 4.71 (d, J = 3.0 Hz, 1 H), 3.89–3.93 (m, 2 H), 3.87 (s, 3 H), 3.56 (d, J = 4.8 Hz, 1 H), 3.50 (t, J = 6.6 Hz, 1 H), 3.38–3.40 (m, 1 H), 3.26 (dd, J = 8.1, 17.4 Hz, 1 H), 2.99 (dd, J = 3.0, 17.4 Hz, 1 H), 2.78 (td, J = 4.8, 12.0 Hz, 1 H), 2.69 (s, 3 H), 2.42 (s, 3 H), 2.23–2.40 (m, 2 H), 1.46–1.61 (m, 2 H), 1.39–1.46 (m, 2 H), 0.96 (s, 9 H), 0.22 (s, 3 H), 0.15 (s, 3 H).

¹³C NMR (75 MHz, CDCl₃): δ = 145.5, 143.1, 142.5, 135.2, 129.6, 127.4, 124.6, 119.9, 113.8, 91.7, 74.4, 73.7, 69.6, 56.5, 49.7, 46.9, 34.9, 33.9, 33.8, 31.8, 25.7, 22.9, 21.5, 18.0, -5.0, -5.2.

MS (EI+): m/z (%) = 603 (20), 546 (30), 528 (10), 343 (12), 313 (12), 198 (100).

HRMS (EI+): m/z calcd for C₃₁H₄₅NO₇SSi: 603.2686; found: 603.2679.

N-(2-((3*S*,3a*S*,3a1*R*,9a*S*)-3-[(*tert*-Butyldimethylsilyl)oxy]-9a-hydroxy-5-methoxy-1,2,3,3a,3a1,9a-hexahydrophenanthro[4,5-*bcd*]furan-3a1-yl)ethyl)-*N*,4-dimethylbenzenesulfonamide (**2**)

To a stirred solution of diol **13** (150 mg, 0.25 mmol) in CH₂Cl₂ (5 mL) at 0 °C was added NEt₃ (63 μL, 0.50 mmol) followed by MsCl (29 μL, 0.37 mmol). The resulting solution was stirred at 0 °C for 30 min, and then it was diluted with CH₂Cl₂/aq NaHCO₃ (10 mL/10 mL). The layers were separated and the aqueous layer was further extracted with CH₂Cl₂ (2 × 10 mL). The organic layers were combined, dried (MgSO₄), filtered, and concentrated to afford a residue that was used without purification in the next step. To the crude mesylate (obtained from the previous step) dissolved in toluene (3 mL) was added DBU (1 mL). The mixture was heated to reflux for 1 h. The mixture was allowed to cool to r.t. after which it was diluted with EtOAc/sat. aq NH₄Cl (10 mL/10 mL). The layers were separated and the aqueous phase was further extracted with EtOAc (2 × 10 mL). The organic layers were combined, dried (MgSO₄), filtered, and concentrated to afford a residue that was

chromatographed on silica gel using hexanes/EtOAc (2:1) as eluent to afford the product **2** as a viscous oil; yield: 101 mg (70% over 2 steps); $[\alpha]_D^{20} +59.98$ ($c = 0.3$, CHCl_3); $R_f = 0.6$ (2:1, hexanes/EtOAc).

IR (neat): 3511, 2952, 2927, 2854, 1742, 1631, 1598, 1505, 1333, 1272, 1157, 1115 cm^{-1} .

^1H NMR (300 MHz, CDCl_3): $\delta = 7.53$ (d, $J = 8.2$ Hz, 2 H), 7.25 (d, $J = 8.2$ Hz, 2 H), 6.69 (d, $J = 8.0$ Hz, 1 H), 6.61 (d, $J = 8.0$ Hz, 1 H), 6.25 (d, $J = 9.6$ Hz, 1 H), 5.62 (d, $J = 9.6$ Hz, 1 H), 4.47 (d, $J = 6.3$ Hz, 1 H), 3.87 (s, 3 H), 3.53 (ddd, $J = 10.9, 6.1, 4.5$ Hz, 1 H), 3.04–2.96 (m, 2 H), 2.62 (s, 3 H), 2.40 (s, 3 H), 2.18–2.03 (m, 1 H), 2.02–1.89 (m, 1 H), 1.80–1.59 (m, 2 H), 1.52–1.40 (m, 2 H), 0.90 (s, 9 H), 0.13 (s, 3 H), 0.04 (s, 3 H).

^{13}C NMR (75 MHz, CDCl_3): $\delta = 145.4, 144.7, 143.2, 137.2, 135.1, 129.7, 129.6, 127.5, 123.6, 123.2, 118.2, 113.8, 97.3, 75.8, 73.3, 56.7, 50.3, 46.9, 35.2, 34.4, 33.0, 25.9, 25.2, 21.6, 18.2, -4.5, -4.9$.

MS (EI+): m/z (%) = 436 (62), 432 (59), 416 (43), 374 (39), 225 (26), 198 (23), 157 (24), 125 (40), 93 (28), 71 (26), 57 (100).

HRMS (EI+): m/z calcd for $\text{C}_{31}\text{H}_{44}\text{NO}_6\text{Si}$: 585.2580; found: 586.2647.

(4S,4aR,7S,7aS,12bR)-7-[(tert-Butyldimethylsilyloxy]-9-methoxy-3-methyl-2,3,4,4a,5,6,7,7a-octahydro-1H-4,12-methanobenzofuro[3,2-e]isoquinolin-4a-ol (**14**)

To a stirred solution of tosylamide **2** (50 mg, 0.085 mmol) in THF (8 mL) was added *t*-BuOH (100 μL , 1.05 mmol). The solution was cooled to -78°C and ammonia was condensed (30 mL) to the reaction mixture. Li (60 mg, 8.65 mmol) was added in three portions over 10 min. The resulting deep blue solution was stirred at -78° for 10 min. It was quenched by the sequential addition of solid NH_4Cl (4 g), MeOH (20 mL), and sat. aq NH_4Cl (20 mL). The reaction mixture was allowed to warm up to r.t. after which it was diluted with CH_2Cl_2 /sat. aq NH_4Cl (30 mL/30 mL). The layers were separated and the aqueous phase was further extracted with CH_2Cl_2 (2×30 mL). The organic layers were combined, dried (MgSO_4), filtered, and concentrated to afford a residue that was chromatographed on silica gel using CH_2Cl_2 /MeOH (9:1) as eluent to afford the product **14** as a solid; yield: 28 mg (76%); mp 120 – 122°C (MeOH); $[\alpha]_D^{20} +66.4$ ($c = 0.3$, CH_2Cl_2); $R_f = 0.43$ (9:1 CH_2Cl_2 /MeOH).

IR (film): 3411, 2927, 2853, 1634, 1500, 1447, 1257, 1108 cm^{-1} .

^1H NMR (300 MHz, CDCl_3): $\delta = 6.75$ (d, $J = 8.1$ Hz, 1 H), 6.62 (d, $J = 8.1$ Hz, 1 H), 4.42 (d, $J = 6.3$ Hz, 1 H), 3.89 (s, 3 H), 3.42–3.48 (m, 1 H), 3.13 (d, $J = 18.3$ Hz, 1 H), 2.81 (d, $J = 5.4$ Hz, 1 H), 2.59 (dd, $J = 5.4, 18.3$ Hz, 1 H), 2.38 (s, 3 H), 2.17–2.22 (m, 1 H), 1.92–2.01 (m, 1 H), 1.55–1.58 (m, 2 H), 1.42–1.46 (m, 1 H), 0.91 (s, 9 H), 0.13 (s, 3 H), 0.03 (s, 3 H).

^{13}C NMR (75 MHz, CDCl_3): $\delta = 144.3, 143.9, 132.6, 125.3, 118.3, 114.8, 96.5, 73.9, 70.3, 64.8, 57.0, 46.7, 45.6, 42.7, 30.4, 29.7, 27.4, 25.8, 22.0, -4.6, -5.0$.

MS (EI+): m/z (%) = 431 (20), 374 (100), 313 (15), 157 (15), 75 (20), 69 (35), 57 (20).

HRMS (EI+): m/z calcd for $\text{C}_{24}\text{H}_{37}\text{NO}_4\text{Si}$: 431.2492; found: 431.2482.

(+)-Oxycodone [*ent*-(**1**)]

To a stirred solution of ether **14** (15 mg, 0.035 mmol) in THF (1 mL) was added TBAF (174 μL , 0.174 mmol). The reaction mixture was stirred at r.t. for 3 h after which it was diluted with EtOAc/ H_2O (10 mL/3 mL). The layers were separated and the aqueous layer was further extracted with CH_2Cl_2 (2×10 mL). The organic layers were combined, dried (MgSO_4), filtered, and concentrated to afford a residue that was used as crude in the next step. To the crude alcohol (obtained from the previous step) dissolved in CH_2Cl_2 (3 mL) was added Dess–Martin periodinane (60.6 mg, 0.143 mmol). The reaction mixture

was stirred at r.t. for 2 h. It was diluted with sat. aq $\text{Na}_2\text{S}_2\text{O}_3$ (1 mL), followed by sat. aq NaHCO_3 (1 mL). The layers were separated and the aqueous phase was further extracted with CH_2Cl_2 (2×10 mL). The organic layers were combined, dried (MgSO_4), filtered, and concentrated to afford a residue that was chromatographed on silica gel using CH_2Cl_2 /MeOH (9:1) as eluent to afford the product *ent*-(**1**) as a solid; yield: 6.5 mg (59% over 2 steps); mp 206 – 208°C (Lit.¹² mp 207.4 – 209.5°C); $[\alpha]_D^{20} +205$ ($c = 0.3$, CHCl_3) [Lit.¹² $[\alpha]_D^{20} -207$ ($c = 0.09$, CHCl_3)].

^1H NMR (300 MHz, CDCl_3): $\delta = 6.69$ (d, $J = 8.1$ Hz, 1 H), 6.62 (d, $J = 8.1$ Hz, 1 H), 4.66 (s, 1 H), 3.89 (s, 3 H), 3.15 (d, $J = 18.6$ Hz, 1 H), 3.01 (ddd, $J = 5.1, 14.4$ Hz, 1 H), 2.87 (d, $J = 5.8$ Hz, 1 H), 2.55 (dd, $J = 5.8, 18.6$ Hz, 1 H), 2.40 (s, 3 H), 2.36–2.51 (m, 2 H), 2.28 (dt, $J = 3.3, 14.4$ Hz, 1 H), 2.12–2.18 (m, 1 H), 1.83–1.90 (m, 1 H), 1.64 (dd, $J = 3.3, 14.4$ Hz, 1 H), 1.55–1.60 (m, 1 H).

^{13}C NMR (75 MHz, CDCl_3): $\delta = 208.7, 145.1, 143.0, 129.5, 125.1, 119.6, 115.0, 90.5, 70.5, 64.7, 56.9, 50.3, 45.4, 42.8, 36.2, 31.5, 30.6, 22.1$.

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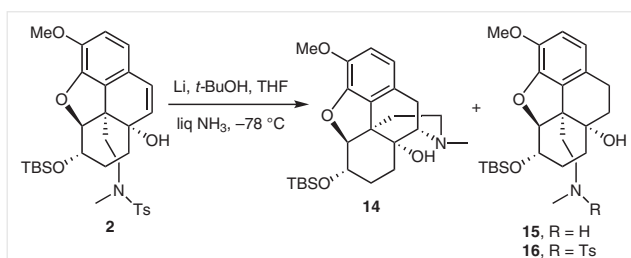
Supporting Information

Supporting information for this article is available online at <https://doi.org/10.1055/s-0037-1611335>. Included are spectral data for compounds **2**, **3**, **4**, **6**, **11**, **12**, **13**, **14**, *ent*-(**1**), **16**.

References

- (1) Jakubska, A.; Przado, D.; Steininger, M.; Aniol-Kwiatkowska, A.; Kadej, M. *Appl. Ecol. Env. Res.* **2005**, *3*, 29.
- (2) (a) Lin, H.-C.; Wang, Z.; Boyd, C.; Simoni-Wastila, L.; Buu, A. *Addict. Behav.* **2018**, *76*, 348. (b) Li, Y.; Hong, R. A.; Robbins, B.; Gibbons, K. M.; Holman, A. E.; Caird, M. S.; Farley, F. A.; Abbott, M. D.; Burke, M. C. *Spine* **2018**, *43*, E98.
- (3) <https://www.rexall.ca/articles/view/894/Percocet>.
- (4) <https://www.drugs.com/dosage/oxycotin.html>.
- (5) Chapman, R.; Rider, L. S.; Hong, Q.; Kyle, D.; Kupper, R. US Patent 7674800B2, **2004**.
- (6) (a) Freund, M.; Speyer, E. DE Patent 286431, **1914**. (b) Freund, M.; Speyer, E. DE Patent 296916, **1916**. (c) Freund, M.; Speyer, E. *Angew. Chem.* **1914**, *27*, 380. (d) Lutz, R. E.; Small, L. J. *Org. Chem.* **1939**, *4*, 220. (e) Kok, G. B.; Scammells, P. J. *RSC Adv.* **2012**, *2*, 11318.
- (7) (a) Millgate, A. G.; Pogson, B. J.; Wilson, I. W.; Kutchan, T. M.; Zenk, M. H.; Gerlach, W. L.; Fist, A. J.; Larkin, P. J. *Nature* **2004**, *431*, 413. (b) Fist, A. J.; Byrne, C. J.; Gerlach, W. L. US Patent 606749, **2000**. (c) Fist, A. J. US Patent Application 20090227796 A1, **2009**. (d) Thodey, K.; Galanie, S.; Smolke, C. D. *Nat. Chem. Biol.* **2014**, *10*, 837.
- (8) <https://tasalk.com.au/>.
- (9) Gates, M.; Tschudi, G. J. *Am. Chem. Soc.* **1952**, *74*, 1109.

- (10) (a) Rinner, U.; Hudlicky, T. *Top. Curr. Chem.* **2012**, *309*, 33. (b) Zezula, J.; Hudlicky, T. *Synlett* **2005**, 388. (c) Taber, D. F.; Neubert, T. D.; Schlecht, M. F. In *Strategies and Tactics in Organic Synthesis*; Harmata, M., Ed.; Elsevier: London, **2004**, 353. (d) Novak, B. H.; Hudlicky, T.; Reed, J. W.; Mulzer, J.; Trauner, D. *Curr. Org. Chem.* **2000**, *4*, 343. (e) Hudlicky, T.; Butora, G.; Fearnley, S.; Gum, A.; Stabile, M. In *Studies in Natural Products Chemistry*; Atta-ur-Rahman, Ed.; Elsevier: Amsterdam, **1996**, 43.
- (11) Rice, K. J. *Org. Chem.* **1980**, *45*, 3135.
- (12) Kimishima, A.; Umihara, J.; Mizoguchi, A.; Yokoshima, S.; Fukuyama, T. *Org. Lett.* **2014**, *16*, 6244.
- (13) Zylstra, G. J.; Gibson, D. T. *J. Biol. Chem.* **1989**, *264*, 14940.
- (14) For use of these metabolites in synthesis, see: (a) Taher, E. S.; Banwell, M. G.; Buckler, J. N.; Yan, Q.; Lan, P. *Chem. Rec.* **2018**, *18*, 239. (b) Banwell, M. G.; Bolte, B.; Buckler, J. N.; Chang, E. L.; Lan, P.; Taher, E. S.; White, L. V.; Willis, A. C. *J. Proc. Royal Soc. New South Wales* **2016**, *Parts 1 & 2*, 34. (c) Lewis, S. E. In *Arene Chemistry: Reaction Mechanisms and Methods for Aromatic Compounds*; Mortier, J., Ed.; Wiley-VCH: Weinheim, **2015**, 915. (d) Lewis, S. E. *Chem. Commun.* **2014**, *50*, 2821. (e) Griffen, J. A.; Kenwright, S. J.; Abou-Shehata, S.; Wharry, S.; Moody, T. S.; Lewis, S. E. *Org. Chem. Front.* **2014**, *1*, 79. (f) Rinner, U. In *Comprehensive Chirality*; Carreira, E. M.; Yamamoto, H., Ed.; Elsevier: Amsterdam, **2012**, 240. (g) Bon, J.-Y.; Lee, B.; Banwell, M. G.; Cade, I. A. *Chim. Oggi Chem. Today* **2012**, *30*, 22. (h) Hudlicky, T.; Reed, J. W. *Synlett* **2009**, 685. (i) Hudlicky, T.; Reed, J. W. *Chem. Soc. Rev.* **2009**, *38*, 3117. (j) Boyd, D. R.; Bugg, T. D. H. *Org. Biomol. Chem.* **2006**, *4*, 181. (k) Johnson, R. A. *Org. React.* **2004**, *63*, 117. (l) Banwell, M. G.; Edwards, A. J.; Harfoot, K. A.; Jolliffe, K. A.; McLeod, M. D.; McRae, K. J.; Stewart, S. G.; Vogtle, M. *Pure Appl. Chem.* **2003**, *75*, 223. (m) Boyd, D. R.; Sharma, N. D. *J. Mol. Catal. B: Enzym.* **2002**, *19–20*, 31. (n) Boyd, D. R.; Sharma, N. D.; Allen, C. C. R. *Curr. Opin. Biotechnol.* **2001**, *12*, 564. (o) Hudlicky, T.; Gonzalez, D.; Gibson, D. T. *Aldrichimica Acta* **1999**, *32*, 35. (p) Boyd, D. R.; Sheldrake, G. N. *Nat. Prod. Rep.* **1998**, *15*, 309. (q) Reed, J. W.; Hudlicky, T. In *Advances in Asymmetric Synthesis*; Hassner, A., Ed.; JAI Press: Greenwich, **1995**, 271. (r) Brow, S. M.; Hudlicky, T. In *Organic Synthesis: Theory and Applications*; Hudlicky, T., Ed.; JAI Press: Greenwich, **1993**, 113. (s) Carless, H. A. J. *Tetrahedron: Asymmetry* **1992**, *3*, 795. (t) Widdowson, D. A.; Ribbons, D. W.; Thomas, D. D. *Janssen Chim. Acta* **1990**, *8*, 3.
- (15) (a) Hudlicky, T.; Endoma, M. A. A.; Butora, G. *J. Chem. Soc., Perkin Trans. 1* **1996**, 2187. (b) Endoma, M. A.; Bui, V. P.; Hansen, J.; Hudlicky, T. *Org. Process Res. Dev.* **2002**, *6*, 525.
- (16) (a) Endoma-Arias, M. A. A.; Hudlicky, J. R.; Simionescu, R.; Hudlicky, T. *Adv. Synth. Cat.* **2014**, *356*, 333. (b) Hudlicky, T.; Endoma-Arias, M. A. A. *Chem. Eur. J.* **2016**, *22*, 14540.
- (17) Koizumi, H.; Yokoshima, S.; Fukuyama, T. *Chem. Asian J.* **2010**, *5*, 2191.
- (18) Oxidation of the diol **3** to afford a keto alcohol followed by NaBH₄ reduction afforded an epimeric diol. Attempts to form the acetonide of this supposed *trans*-isomer failed. The diol **3** was smoothly converted to the acetonide. Unpublished observations.
- (19) (a) Parker, K. A.; Fokas, D. *J. Am. Chem. Soc.* **1992**, *114*, 9688. (b) Parker, K. A.; Fokas, D. *J. Org. Chem.* **2006**, *71*, 449.
- (20) The outcome of the hydroamination was dependent on the condition of the reaction. Initial experiments resulted in the formation of an undesirable side product **15** (isolated as **16**) from the reduction of the cinnamyl alcohol moiety (Scheme 3). The formation of this product was eliminated by reversing the order of addition of reagents. Thus, Li was added last in the reaction and in several portions (see experimental section). The same phenomenon has been observed originally by Birch²¹ and later by Hall²² in their studies with dissolving metal reduction of cinnamyl alcohols.
- N-(2-((3*S*,3*aS*,3*a1R*,9*aR*)-3-[(*tert*-Butyldimethylsilyloxy]-9*a*-hydroxy-5-methoxy-1,3,3*a*,8,9,9*a*-hexahydrophenanthro[4,5-*bcd*]furan-3*a1*(2*H*)-yl)ethyl)-*N*,4-dimethylbenzenesulfonamide (**16**)**
Mp 67–68 °C (MeOH); [α]_D²⁰ +2.0 (c = 0.2, CH₂Cl₂); R_f = 0.5 (2:1 hexanes/EtOAc).
IR (film): 3500, 2925, 2854, 1735, 1600, 1461, 1338, 1257, 1160, 830 cm⁻¹.
¹H NMR (300 MHz, CDCl₃): δ = 7.57 (d, *J* = 8.4 Hz, 2 H), 7.27 (d, *J* = 8.4 Hz, 2 H), 6.76 (d, *J* = 8.1 Hz, 1H), 6.63 (d, *J* = 8.1 Hz, 1 H), 4.55 (d, *J* = 4.8 Hz, 1 H), 3.87 (s, 3H), 3.60 (m, 1 H), 3.34–3.38 (m, 1 H), 2.84–2.97 (m, 2 H), 2.71 (s, 3 H), 2.65–2.72 (m, 1 H), 2.43 (s, 3 H), 2.12–2.21 (m, 2 H), 1.86–1.96 (m, 1 H), 1.40–1.81 (m, 4 H), 0.91 (s, 9 H), 0.14 (s, 3 H), 0.07 (s, 3 H).
¹³C NMR (75 MHz, CDCl₃): δ = 143.1, 142.7, 135.0, 131.4, 129.6, 127.3, 125.4, 120.3, 114.3, 93.9, 77.2, 72.8, 71.5, 56.7, 49.7, 47.2, 35.2, 33.0, 31.8, 25.8, 25.0, 24.3, 21.5, 18.0, -4.8, -5.2.
MS (EI+): *m/z* (%) = 587 (10), 530 (20), 512 (25), 439 (25), 403 (20), 345 (40), 343 (70), 327 (50), 315 (45), 198 (100), 183 (30), 97 (25).
HRMS (EI+): *m/z* calcd for C₃₁H₄₅NO₆SSi: 587.2737; found: 587.2726.



Scheme 3

- (21) Birch, A. J. *J. Chem. Soc.* **1945**, 809.
(22) Hall, S. S. *J. Org. Chem.* **1973**, *38*, 1738.