

# From Anti-EBV Immune Responses to the EBV Diseasome via Cross-reactivity

Darja Kanduc<sup>1</sup> Yehuda Shoenfeld<sup>2,3</sup>

Global Med Genet 2020;7:51-63.

Address for correspondence Darja Kanduc, PhD, Department of Biosciences, Biotechnologies and Biopharmaceutics, University of Bari, Via Orabona 4, Bari 70125, Italy (e-mail: dkanduc@gmail.com).

### **Abstract**

### **Keywords**

- ► EBV epitopes
- systemic lupus erythematosus
- ► cross-reactivity
- autoimmunity
- negative selection
- self-reactive lymphocytes
- pathogenic autoantibodies

Sequence analyses highlight a massive peptide sharing between immunoreactive Epstein-Barr virus (EBV) epitopes and human proteins that—when mutated, deficient or improperly functioning—associate with tumorigenesis, diabetes, lupus, multiple sclerosis, rheumatoid arthritis, and immunodeficiencies, among others. Peptide commonality appears to be the molecular platform capable of linking EBV infection to the vast EBV-associated diseasome via cross-reactivity and questions the hypothesis of the "negative selection" of self-reactive lymphocytes. Of utmost importance, this study warns that using entire antigens in anti-EBV immunotherapies can associate with autoimmune manifestations and further supports the concept of peptide uniqueness for designing safe and effective anti-EBV immunotherapies.

### Introduction

The connection between Epstein-Barr virus (EBV) and Burkitt's lymphoma (BL) was discovered in 1964. Almost contemporaneously, high anti-EBV antibody levels were found in BL. Since then, EBV infection has been associated with a wide spectrum of malignancies that, besides BL, comprehends different types of lymphomas, nasopharyngeal carcinoma (NPC), breast and brain cancer, and oral hairy leukoplakia, among others. In addition, EBV has been implicated in a wide variety of diseases, including systemic lupus erythematosus (SLE), Sjögren's syndrome, multiple sclerosis (MS), myasthenia gravis (MG), rheumatoid arthritis (RA), autoimmune thyroid disorders, inflammatory bowel disease, celiac diseases, diabetes, Parkinson's disease, myopericarditis, dilated cardiomyopathy, and even death.

At the same time, anti-EBV antibody level was found to be higher in BL patients than in control subjects.<sup>3,15–17</sup> High level of anti-EBV immunoglobulin G antibodies were also found in subjects with NPC, <sup>18–21</sup> with IgG reactivity increasing

significantly with tumor stage<sup>21</sup>; Hodgkin and non-Hodgkin lymphomas<sup>22-24</sup>; precancerous gastric lesions<sup>25</sup>; MS<sup>26-29</sup>; RA<sup>30-32</sup>; MG<sup>33,34</sup>; and SLE,<sup>29,31,34</sup> among others. In general, high antibody titers to EBV appeared to be related to a worse prognosis, a phenomenon that has been described by Coutinho's laboratory<sup>35</sup> as "the advantage of being low-respondents." Currently, measurement of increased anti-EBV antibody titers is utilized to predict, to detect, and to monitor the progression of EBV-related cancers and progression of the various EBV-induced diseases.<sup>36-38</sup>

Today, in front of such well known clinical context, the molecular mechanism(s) by which anti-EBV immune responses relate to the EBV diseasome, from lymphomas to Parkinson's disease, are still obscure. From a logical point of view, a central question remains unanswered and perhaps, as far as we know, has never been clearly posed: why the powerful anti-EBV immune responses herald cancers, autoimmune diseases, and death instead of eradicating the viral infection and re-establishing a healthy status?

published online
August 31, 2020

DOI https://doi.org/ 10.1055/s-0040-1715641. ISSN 2699-9404. License terms



<sup>&</sup>lt;sup>1</sup> Department of Biosciences, Biotechnologies, and Biopharmaceutics, University of Bari, Bari, Italy

<sup>&</sup>lt;sup>2</sup> Zabludowicz Center for Autoimmune Diseases, Sheba Medical Center, Tel-Aviv University School of Medicine, Tel-Hashomer, Israel

<sup>&</sup>lt;sup>3</sup> I.M. Sechenov First Moscow State Medical University of the Ministry of Health of the Russian Federation, Sechenov University, Moscow, Russia

In the clinical frame exposed above and on the basis of previous scientific reports<sup>39-42</sup> that have detailed a high level of peptide sharing between EBV and human proteins involved in crucial functions, this study investigates whether the immune responses that accompany active EBV infection have the potential to cross-react with and damage human proteins that, when altered, can lead to various cancer and autoimmune diseases. That is, the thesis is explored according to which the anti-EBV immune responses that should be a "protective defense" from EBV infection actually cross-react with human proteins, in this way setting up an anti-human protein assault with catastrophic pathologic sequelae in the body. Specifically, the present study used the pentapeptide as an antigenic and immunogenic unit, 43-48 and analyzed 3,197 experimentally validated immunoreactive EBV-derived epitopes for pentapeptide matches with the human proteome. Data are reported on a vast peptide sharing between EBV epitopes and proteins involved in tumorigenesis, autoimmune disorders, diabetes, and death, among others. The data suggest that cross-reactivity is the mechanism underlying the causal connection between EBV infection, immune response, and the EBV-associated diseases.

### **Methods**

An EBV immunome formed by 3,197 immunopositive linear epitopes was assembled from Immune Epitope DataBase (IEDB, www.iedb.org).<sup>49</sup> The immunopositive EBV epitopes are listed in **Supplementary Table S1** (available in the online version). EBV epitope sequences were dissected into pentapeptides overlapped each other by four amino acid (aa) residues. The resulting 11,564 pentapeptides were analyzed for occurrence(s) within the human proteome using Pir Peptide Match Program.<sup>50</sup> Proteins related to EBV-induced diseases were annotated. UniProtKB database (http://www.uniprot.org/)<sup>51</sup> PubMed, and OMIM resources were used.

### Results

### Quantitation of the Peptide Sharing between EBV Epitopes and the Human Proteome

Following matching analyses of the 11,564 pentapeptides composing the 3,197 experimentally validated immunoreactive EBV epitopes, it was found that almost all of the epitope-derived pentapeptides (i.e., 93%) are widespread among thousands of human proteins ( $\succ$  **Table 1**). From a mathematical point of view, if one considers that the probability of a pentapeptide to occur in two proteins is  $20^{-5}$  (or 1 out of 3,200,000 or 0.0000003125), then the peptide overlap existing between the EBV immunome and the human proteome is staggering.

### **Distribution of the Peptide Sharing among EBV Epitopes**

A synthetic snapshot (i.e., 201 EBV epitopes) of the immunoreactive peptide sharing is shown in **- Table 2**, where peptide sequences shared with the human proteins are given in capital format and peptide fragments uniquely present in EBV are given with aa in small bold format. **- Table 2** clearly shows that the immunoreactive EBV epitopes are predominantly composed by peptide sequences common to human proteins.

**Table 1** Numerical description of the pentapeptide sharing between the set of 3,197 immunopositive EBV epitopes and the human proteome

Pentapeptides composing the 3,197 EBV epitope immunome	11,564
EBV epitope pentapeptides not shared with the human proteome	798
EBV epitope pentapeptides shared with the human proteome	10,766
Human proteins sharing pentapeptides with EBV epitopes	18,744
Occurrences of EBV epitope pentapeptides in the human proteome (including multiple occurrences)	137,805

Abbreviation: EBV, Epstein-Barr virus.

Immunologically, **Tables 1** and **2** document that the experimentally validated immunoreactive EBV epitopes mostly consist of pentapeptides that also occur in human proteins, in this way indicating a highest cross-reactivity potential, given the fact that a pentapeptide is a minimal immune determinant that contains the immunological information in terms of both immunogenicity and antigenicity.<sup>43–48</sup>

## The Pathological Implications of the Peptide Sharing between EBV Epitopes and Human Proteins: Lymphomas and Leukemias

Numerous cancer-related proteins share peptides with the here analyzed self-reactive EBV epitopes. Reasons of space do not permit a detailed peptide-by-peptide description of the sharing and only a few examples are described in ►Tables 3 and 4. Specifically, ►Table 3 shows the peptide sharing between the immunoreactive EBV epitopes and human proteins that-when mutated, modified, improperly functioning or deficient-are implicated in lymphomagenesis/ leukemogenesis. 52-91 It can be seen that the extent of the peptide sharing is very high and comes to the fore with glaring evidence when focusing on histone-lysine N-methyltransferase 2D (KMT2D), the disruption of which perturbs germinal center B cell development and promotes lymphomagenesis.<sup>77,78</sup> KMT2D alterations are involved in follicular lymphoma and diffuse large B-cell lymphoma, 92 cutaneous T-cell lymphoma and Sézary syndrome, 93 ocular adnexal MALT-type marginal zone lymphomas, 79 and chronic myeloid leukemia. 94 Moreover, KMT2D alterations are involved in intraocular medulloepithelioma, 95 small cell lung cancer, 96 bladder cancer, 97,98 and non-small-cell lung cancer. 99 Of not less importance, alterations of KMT2D have a causal role in Kabuki syndrome<sup>100</sup> that is characterized by skeletal and visceral abnormalities and cardiac anomalies, 101 hyperinsulinism, <sup>102</sup> epilepsy, <sup>103</sup> desmoid fibromatosis, <sup>104</sup> immunopathological manifestations, 105 lupus, 106 and oriental alterations, <sup>107</sup> among others.

Also, the intense peptide sharing between immunoreactive EBV epitopes and KMT2C is of relevance. KMT2C not only may act as a tumor suppressor in leukemias and T-cell lymphomas, <sup>75,76</sup> but it is also implicated in bladder, breast,

 Table 2
 Pentapeptide sharing between 201 immunoreactive EBV epitopes and human proteins<sup>aa</sup>

IEDB ID <sup>b</sup>	EPITOPE <sup>c,d</sup>	IEDB ID <sup>b</sup>	EPITOPE <sup>c,d</sup>	IEDB ID <sup>b</sup>	EPITOPE <sup>c,d</sup>
950	AEGLRALLARSHVER	45499	NPTQAPVIQLVHAVY	127195	TEMYIMYAM
1518	AGGAGAGGAGGA	46498	NVTQVGSEPISPEIG	127369	WEMRAGREI
1716	AGVFVYGGSKTSLYN	47613	PGAPGGSGSGP	127392	WPTPKTHPV
2390	ALAipqcrL	47760	PGTGPGNGLGEKGDT	127408	yamaiRQAI
2742	ALLVLYSFAL	48320	PLFDRKSDAK	137773	YNLRRGIAL
2743	ALLVLYSFALMLIIIILIIF	48486	PLSRLPFGM	138854	GAGAGAGA
3005	ALWNLHGQALFLGIVL	48852	PPPGRRP <b>ffhpv</b> GE	138856	GRGRGRGR
3600	<b>apify</b> PPVL	48876	PPP <b>qapyq</b> GY	138882	MTAASYARY
3782	APRLPDDPI	49864	PVFDRKSDAK	138873	LMARRARSL
3951	AQEILSDNSEISVFPK	50298	QAKWRLQTL	141342	LLDFVRMGV
5316	AVFDRKSDAK	51685	QNGALAINTF	144799	TLNLT
5317	AVFDRKSVAK	51946	QPRAPIRPI	167590	GPQRR
5326	AVFNRKSDAK	52142	QQ <b>rpvmfv</b> SRVPAKK	186702	PQPRAPIRPIPT
5439	AVLLheesm	53195	RARGRGRGRGEKRP	191290	FIVFLQTHI
8120	DEPASTEPVHDQLL	54367	RKIYDLIEL	227777	HPVAEADYFEY
8905	DKI <b>vqapify</b> PPVLQ	54728	RLRAEAQVK	230640	ASDYSQGAF
9644	DP <b>hgpvq</b> LSYYD	55251	RppifiRLL	230798	FYPPVLQPI
10448	DTPLIPLTIF	55298	RPQKRPSCI	231136	LAYArgqam
10858	DYDASTESEL	55327	RPRPPARSL	231402	RRVRRRVLV
10963	DYSQGAFTPL	55529	RRARSLSAERY	231547	TVFYnippm
11804	EENLLDFVRF	55619	RRIYDIEL	231696	YRTATLRTL
12183	EGGVGWRHW	56506	RYAREAEVRF	231699	YSQGAFTPL
13628	EPDVPPGAIEQGPAD	56523	RYEDPDAPL	231800	AQPAPQAPY
16876	FLRGRAYGI	56650	RYSIFFDY	231839	DSIMLTATF
17110	FMVFLQTHI	56651	RYSIFFDYM	231840	DTRaidqfF
17600	FRKAQIQGL	57755	SFFDRKSDAK	231880	FLQRTDLSY
18328	FVYGGSKTSL	59084	SLFDRKSDAK	231966	HVIQNAFRK
18438	FYnippmPL	59432	SLREWLLRI	232020	KPWLRAHPV
18946	GDDGDDGDEGGDGDE	62305	SVRDRLARL	232021	KQRKPGGPW
19674	GGAGGAGGAGGGAG	67456	TYSAGIVQI	232030	KTIGnfkpy
19737	GGGAGAGGAGAGGGGR	68561	VFSDGRVAC	232074	LPTPMQLAL
20023	GGSKTSLYNLRRGTA	69558	VLKDAIKDL	232076	LQALSNLIL
21719	GPPAA	70251	VPAPAGPIV	232078	LQSSSYPGY
21723	GPPAAGPPAAGPPAA	70624	VQPPQLTQV	232080	LSaerytLF
21870	GQGGSPTAM	70932	VSFlefvgw	232081	LSVIPSNPY
22159	GRPAVFDRKSDAKST	71968	VYAASFSPNL	232086	LTQAAGQAF
22976	GVFVYGGSKTSLYNL	72028	VYGGSKTSL	232095	LVSSGNTLY
23324	GydvghGPL	72251	WAPSV	232096	LVSSSAPSW
23449	GYRTATLRTL	73221	WVPSV	232103	MEQRVMATL
23994	Hhiwqnll	74120	yhlivDTDSL	232177	QEPGPVGPL
24170	HLAAQGMAY	75188	YNLRRGTAL	232178	QEQLSDTPL
24533	HPVgeady	75189	YNLRRGTALAIPQ	232199	RESIVCYFM
24666	HRCQAIRK	75356	YPL <b>heqhg</b> M	232214	RLHRLLLMR
	1		<u> </u>	1	

(Continued)

Table 2 (Continued)

IEDB ID <sup>b</sup>	EPITOPE <sup>c,d</sup>	IEDB ID <sup>b</sup>	EPITOPE <sup>c,d</sup>	IEDB ID <sup>b</sup>	EPITOPE <sup>c,d</sup>
24667	HRCQAIRKK	75360	YPLHKQHGM	232232	RPAP <b>pkiam</b>
26480	IIFIFRRDLLCPLGAL	75731	YSFALMLIIIILIIFIFRRD	232276	SEPCEALDL
26538	IIIILIIFI	79634	QPRAPIRPIT	232308	SQISNTEMY
27103	ILIIFIFRRDLLCPLGALCI	93251	LLARSHVER	232332	TEdnvppwl
27301	ILRQLLTGGVKKGRP	94034	THIFAEVLKD	232416	VTFSAGTFK
27375	ILTDFSVIK	97317	fwemrAGREITQ	232419	VTTQRQSVY
29618	IYLLEMLWRL	98084	GVFVYGGSK	232427	waqigHIPY
30430	KEHVIQNAF	101654	FVYGGSKTSLY	232437	WQRRYRRIY
30431	KEHVIQNAFRK	101878	LQTHIFAEV	232473	YQEPPAHGL
33207	KR <b>ppifi</b> RR	102253	YPL <b>heqyg</b> M	237896	QTAAAVVF
33866	KTSLYNLRRGTALA	106084	RPRSPSSQSSSSGSPPRRP	237920	RYKNRVASR
35162	LDFVRFMGV	107724	AARQRLQDI	540571	QPRLTPPQPL
35533	LEKARGSTY	107869	GPKVKRPPI	540583	RPTELQPTP
37153	LLDFVRFMGV	108006	LLDFV <b>rfmgy</b>	540628	TSSPSMPEL
38514	LPGPQVTAVLLHEES	108191	VMATLLPPV	548981	LLDFVRFMG
39102	<b>Irgkw</b> QRRYR	118970	PPPGRRP	548987	NGALAINTF
39634	LSRLPFGMA	124861	WNLHGQALFL	548994	QNGALAINT
41113	MARRARSLSaerytL	126528	LAsamrmLW	595247	FGLVLFPAQI
41147	MATLLPPVPQQPRAG	126967	RPRPrtpew	653929	AAQGMAY
41841	mkkawLSRAQQADAG	126980	RRAALSGHL	672845	PIFIRRL
42525	MSDEGPGTGPGNGLG	126985	RRLHRLLLM	674203	RamsfiATY
42941	MVFLQTHIFAEVLKD	126986	RRRRRAAL	675184	RppifiR
44181	NIAEGLRAL	126991	RRYRRIYDL	676208	RRIYDLI
45378	N <b>pkfen</b> IAEGLRALL	127118	SQAAFGLPI	695961	QAPYPGYEE

Abbreviations: EBV, Epstein-Barr virus; IEDB, Immune Epitope DataBase.

colorectal, endometrial, gastric, head and neck, lung, and liver cancer, and in medulloblastoma. 108

Then, in spite of the lack of space, it is mandatory noting the harmful cross-reactivity platform represented by the peptide commonality between the immunoreactive EBV epitopes and Wiskott-Aldrich syndrome protein (WASP) (►Table 4). The 29 pentapeptides shared with EBV epitopes mainly occur throughout the central and COOH regulatory domains of the WASP primary sequence (>Fig. 1, shared peptides in underlined bold character) and produce a "bull" for the EBV-activated immune system that is practically impossible not to hit. Hitting WASP can lead to lymphomagenesis. Indeed, WASP is a tumor suppressor frequently low or absent in anaplastic large cell lymphoma. 92 WASP deficiency relates to Wiskott-Aldrich syndrome (WAS). 109-112 WAS is characterized by eczema, thrombocytopenia, recurrent infections, immunodeficiency, neutropenia, and bloody diarrhea. 113 A large proportion of WAS patients develop autoimmunity and allergy since WASP appears to play an important role in the activation of CD4(+)

CD25(+)FOXP3(+) natural regulatory T cells.<sup>114</sup> Even in the absence of typical clinical manifestations of WAS, a low expression of WASP associates with the pathogenesis of a subtype of inflammatory bowel disease.<sup>115</sup> Furthermore, deficiency of WASP associates with exacerbated experimental arthritis.<sup>116</sup>

Overall, the peptide sharing between the immunoreactive EBV epitopes and KMT2D, KMT2C, and WASP proteins suffices to define the constellation of human diseases associated with EBV infection.

### The Pathological Implications of the Peptide Sharing between EBV Epitopes and Human Proteins: Various Cancers and Diseases

► **Table 4** illustrates that the EBV epitope-derived pentapeptides are widespread among the most disparate human proteins able to cause, when altered, a vast spectrum of diseases, from diabetes and sterility to myocarditis and death, <sup>117–167</sup> the latter two being possibly associated with the Titin imposing peptide sharing (250 shared pentapeptides).

<sup>&</sup>lt;sup>a</sup>Epitopes assembled from IEDB (www.iedb.org). <sup>49</sup> Epitope experimental details and references are available at www.iedb.org.

<sup>&</sup>lt;sup>b</sup>Epitopes listed according to IEDB ID number. <sup>49</sup>

<sup>&</sup>lt;sup>c</sup>Epitope sequences given in 1-letter code.

<sup>&</sup>lt;sup>d</sup>Pentapeptides shared between EBV epitopes and human proteins are given in capital letters, while pentapeptides present only in EBV are given in bold small format.

Table 3 Pentapeptide sharing between immunoreactive EBV epitopes and human proteins implicated in lymphomagenesis/ leukemogenesis

Shared peptides	Lymphoma/Leukemia-related proteins containing EBV epitope peptide(s) <sup>a</sup>	Ref <sup>b</sup>
LSPLL, RRQKR, LALRA, KEVLE, LGLGD, GNLVT, SLESV, LPTLL, PETVP, ALYLQ, ARVKE, PSLKL, KILLA, NPETL, EGLKD, LYLQQ, QKRPS, VAKVA, DRHSD, LQAIG, LSQVC, RPSCI	ATM: Serine-protein kinase ATM	52
PLPPP, PPLPP, LPTLL, REAIL, AERHG, CKKDH	BANK1: B-cell scaffold protein with ankyrin repeats	53,54
EEEEE, PPLPP, GAGGG, AGAGG, GAGGA, AGGGA, GRGGG, PPPVS, LSAAS, PPLGP, PPVSP, EPGPA, PVSPG, SSLTP, TPPPQ, GDDDD, AVAQS, DPSLG, GNSST, PGLFP, SEPVE, DDAGG, DDDAG	BC11B: B-cell lymphoma/leukemia 11B	55–57
SSSEE, GPPSP, APAST, GPEAR, PCPQA, PQARL, RFIQA	BCL6B: B-cell/lymphoma 6 member B protein	58-62
SPSPP, PLPPV, GSGAG, PAGSL, PVPPP, EPGPA, EQASL, EGTRL, LDLDF, LNQNL, VLQKL	BIN1: Myc box-dependent-interacting protein 1	63
LLLLL, LRLLL, RLRLL, KEDDG, EGGQN	CADM1: Cell adhesion molecule 1	64
PPPPP, LPPPP, GSGSG, GGGSG, GAGGG, GSGGS, SGGSG, GGSGS, SGSGG, LPPVP, PPVPA, PVPPT, QQGSG, CTPGD, PYILD	CBL: E3 ubiquitin-protein ligase CBL	65
PPPLP, PPGPS, LSPLS, SSPQP, ATSGA, ENLLD, EENLL, SPVLG, AFEEV, GPQDP, YDAPG	CRTC2: CREB-regulated transcription coactivator 2	66
LLARL, GASGS, VAGLL, PLHAL, LARLR, SGASP, CGLLR, VPKPR, FIRRL, TDGKT, TPLLT, ALIKT, SSCNS	DAPK1: Death-associated protein kinase 1	67,68
EEEAE, FGLSR, SDLSR, SLESV, KAIEE, VIQLV, IIAVV, VMDLL, IAVVA, IKAIE, ESFTQ, QDVGA, RLFAA, TTGGK, VIKAI, SFTQG	EPHA7: Ephrin type-A receptor 7 (998 aa)	69
LLLLL, PPPPP, LPPPP, ALLLL, LALLL, PPPPS, PPLPP, FPPPP, SLSST, GSPPR, PPQVP, SPSDS, TLSPS, TSEPV, SEDDP, ESVDV, GTPPQ, TDGGG, TSVVQ, VYAAS, EDDPQ, PSELD, DLRPL, FVGDY, KGTPP, PRLFA, VCSVA, HSPVV, ILQIS, LYEAS, PYEAF	FAT1: Protocadherin Fat 1	70,71
PPPPP, GGGGG, EEEEA, AAAAV, SSSEE, GGSGS, GGGGD, RGGSG, GAPGG, ASGPG, LPGVP, VSPAV, PGGLG, VEAHV, GGDGD, LRAAT, ERPLA, FPEGV, GGDKV	HIC1: Hypermethylated in cancer 1 protein	72
PPRPP, RRRKG, ATAAA, SVSQP, AEVLK, LLQTE, SHTAT	KDM6A: Lysine-specific demethylase 6A	73,74
PPPPP, QPPPP, SPPPP, SSSSA, SSSAP, PTPPP, GAPAA, KKRKR, RGGRG, GGRGR, PPPPY, SSSAG, GRGGR, LPPTP, APPTP, PPLGP, PTPLP, SGSPP, PQPPL, SQASA, DDEDL, STSVP, LPGVS, SSGTA, LTPRP, RPRGA, RQRSR, SGLGT, TPRPP, TPRPS, TSVPS, VTLPL, DLILQ, GTPRP, TPRPV, IAVSS, LDTED, TPRPR, LGATI, SAPRK, EGVEV, LSPAN, LSSCP, MQPPP, SLIQL, AKIEA, EDLFG, EEVEN, QGVQV, TPRSQ, VEDLF, LGLYA, PQSGP, DSREG, VSTAD, GPADD, PADDP, QSLIQ, VFPKD, DTDSL, GTFKP, IPQTL, PLQHW, TGQGK, EQHGM, IDDNS, LRPQW, QRHSD, TFKPP, GPRHT	KMT2C: Histone-lysine <i>N</i> -methyltransferase 2C	75,76
EEEEE, QPPPP, PLPPP, SAAAA, LRLLL, PAPAA, PAAAP, PTPPP, APPAP, GRGRG, PPSPG, PSPGS, PSPPP, RGRGR, SPLLP, AAPPA, GPAGP, LLAAL, PAQPP, SLGLA, LAPSP, LSPLL, PGPAG, SPSQS, SQSSS, GGRGR, GLPPP, PQGPP, RLRLL, LPPTP, LRSLG, PTLLL, SPSSQ, TPPPS, ALAPS, EGLRA, GPQPP, PEPPT, PLTEP, SSGSP, AASED, APVAP, AVGPP, DDEEL, ESPAR, GAHGG, GPPRL, KKRKR, LTPRP, PALDD, PPPGR, PPQGP, PPQVP, PPTQH, PTLGK, SDEAE, SPLLG, TPHTK, APYPG, ARPPE, ASDRL, CPSLD, DAAAR, EERPP, EGEGD, EGPST, EPRLA, FPDTK, FPEGL, GPLAI, GPWDP, GTQDP, IKVIE, LGLYA, LRLTP, LSPVI, PLLTV, PMSPP, PPTHP, PPVPQ, PQPLM, PQQPM, PSRPQ, QALAP, QEPPP, QTNQA, RGAFG, RPEFV, SDALG, SPVTP, SQTEL, SRVPA, SYTDP, TGSGG, TTPAG	KMT2D: Histone-lysine <i>N</i> -methyltransferase 2D	77-79
GGGGG, GGGGS, GGGGA, AGGGG, GGSGG, GGGSG, GAGGG, GGAGG, AGAGA, PPPEP, LRALL, LALRA, LTPPS, RALLA, RLLLK, PQAPE, TPLDL, GPETR, RVGAD	NFKB2: Nuclear factor NF-kappa-B p100 subunit	80-83
AAAPA, GAAAS, PAPGL, LLGGG, TPSPS, SLPHP, PHLPP, GSPTA, PLTSE, RDSYA, TTLAA, YPGYA, HRDSY, SYPGY	PRDM1: PR domain zinc finger protein 1	84

(Continued)

Table 3 (Continued)

Shared peptides	Lymphoma/Leukemia-related proteins containing EBV epitope peptide(s) <sup>a</sup>	Ref <sup>b</sup>
EEEEE, PSPPP, APAAA, SPSPP, PSPSP, SPSPS, PLDLS, DEGEE, LDLSV, LLTPV, PTVSP, KQLLQ, VLDLS, LTPVT, TVSPS, VTEDL, AIEEE, TSEET, PAPTV, TPVTV, EAVSF, FKPPP, SFKPP, NIPQT, YSLRL, PALRD, RSQVK, PFVGD	PRDM2: domain zinc finger PR protein 2	85–88
SSSSA, SPLLP, SSSAP, LSPLL, GTPSG, LQSET, PVSRF, AEGKL, PLRPT	SOCS6: Suppressor of cytokine signaling 6	68
KKRKR, AGAAR, LQSLA, TSPTS, RSLLT, LSLVF, AGPSV, DPVHG, GPSVA, QATLG, TQLTQ, DLQDP, LEKQS, PVQGE, QERDV, PKTAS, PLTQP, NIEEF, TPHQP, SHETP	TET1: Methylcytosine dioxygenase TET1	89
PPPLP, PPPPS, SPPPP, SSSEE, ELLEK, SASGS, QSSHL, APGGS, LQAPG, KLSSL, PPSQL, APPSQ, HLLQH, QQASV, VTKQE, VTVLT, PPTQH, PVTVL, GIKRT	TET2: Methylcytosine dioxygenase TET2	90
PPPPP, LPPPP, PLPPP, PPPLP, PPPPS, GGGRG, APGGG, GLPAP, QPPPQ, PAPGP, PRGPP, PPSSG, SLGLA, LPAPG, LPPVP, PLPPV, PPPSR, GGRPG, PPPGR, DLRSL, VGPLS, PMPPP, SEGLV, SGNGP, ADIGA, DIGAP, GGDQG, PVGPL	WASP: Wiskott-Aldrich syndrome protein	91

<sup>&</sup>lt;sup>a</sup>Human proteins reported by UniProt entry names.

**Table 4** Quantitative pentapeptide matching between immunoreactive EBV epitopes and human proteins related to various cancers and diseases

	Pentapep- tides:  Human proteins sharing pentapeptides with EBV epitopes, and disease involvement <sup>c,d</sup>		Refs.	
A <sup>a</sup>	Bb			
3	_	ACHA: Acetylcholine receptor subunit α. MG.	117	
7	-	ACHD: Acetylcholine receptor subunit delta. MG.	117	
8	-	ACHE: Acetylcholine receptor subunit epsilon. MG.	117	
9	11	ACHG: Acetylcholine receptor subunit gamma. MG.	117	
31	42	AGRB1: Adhesion G protein-coupled receptor B1. Inhibits glioma growth.	118,119	
15	-	AKA12: A-kinase anchor protein 12. MG autoantigen. Involved in breast cancer.	120	
27	_	APC: Adenomatous polyposis coli protein. Relates to colorectal adenomas and breast cancer.	121–123	
64	68	APCL: Adenomatous polyposis coli protein 2. Its repression promotes ovarian cancer.	123,124	
57	68	ARI1A: AT-rich interactive domain-containing protein 1A. Bladder, colorectal, endometrial, esophageal, gastric, kidney, liver, lung, ovarian cancers.	108	
68	92	ARI1B: AT-rich interactive domain-containing protein 1B. Liver cancer.	108	
33	-	ARID2: AT-rich interactive domain-containing protein 2. Liver, lung, melanoma cancers.	108	
23	-	BCOR: BCL-6 corepressor. Tumor suppressor in endometrial cancer and medulloblastoma.	108,125	
9	-	C1S: Complement C1s subcomponent precursor. SLE.	126	
20	-	CHD4: Chromodomain-helicase-DNA-binding protein 4. Endometrial cancer.	108	
32	34	CHD6. Chromodomain-helicase-DNA-binding protein 6. Bladder cancer.	108	
38	_	CHD8: Chromodomain-helicase-DNA-binding protein 8. glioblastoma.	108	
10	_	CLAT: Choline O-acetyltransferase. Myasthenic syndrome.	127	
17	_	CO4A: Complement C4-A precursor. SLE.	128	
29	56	CO4A1: Collagen α-1(IV) chain. Tumor suppressor; anti-angiogenic.	129	
17	-	CO4B: Complement C4-B precursor. SLE.	128	
12	13	CUL7: Cullin-7. 3M syndrome with growth restriction, skeletal abnormalities and dysmorphisms.	130	
25	26	DCC: Netrin receptor DCC. Required for axon guidance. Colorectal cancer suppressor.	131	
16	66	DMBT1: Deleted in malignant brain tumors 1 protein. Suppressed in human lung cancer.	132,133	
59	-	DYST: Dystonin. Bullous pemphigoid.	134,135	
42	_	FAT4: Protocadherin Fat 4. Involved in hepatocellular carcinoma. and in gastric cancer risk.	136,137	

<sup>&</sup>lt;sup>b</sup>Further references on the function/disease association at www.uniprot.org, OMIM, and PubMed resources.

Table 4 (Continued)

	Pentapep- ides:  Human proteins sharing pentapeptides with EBV epitopes, and disease involvement <sup>c,d</sup>		Refs.
A <sup>a</sup>	Bb		
34	38	FUBP2: Far upstream element-binding protein 2.	138
11	_	IGF1R: Insulin-like growth factor 1 receptor. Intrauterine and postnatal growth retardation.	139
14	_	INSR: Insulin receptor. Insulin resistance syndrome with pineal hyperplasia.	140
13	_	INSR2: Insulin, isoform 2. Diabetes.	141
27	31	IRS1: Insulin receptor substrate 1. Diabetes. cognitive impairment and Alzheimer's disease.	142
42	45	IRS2: Insulin receptor substrate 2. Diabetes. cognitive impairment and Alzheimer's disease.	142-144
38	50	IRS4: Insulin receptor substrate 4. Diabetes. cognitive impairment and Alzheimer's disease.	142
20	_	KDM5A: Lysine-specific demethylase 5A. Intellectual disability. Inhibits glioma cells migration.	145,146
2	_	LA: Lupus La protein. SLE.	147
16	_	LRP1B: Low-density lipoprotein receptor-related protein 1B precursor 4599.	148
6	_	MAG: Myelin-associated glycoprotein precursor. MS.	149
4	_	MOG: Myelin-oligodendrocyte glycoprotein precursor. MS.	150
13	19	MYRF: Myelin regulatory factor. MS.	151
17	12	MYT1L: Myelin transcription factor 1-like protein. MS.	152
45	47	NBEL2: Neurobeachin-like protein 2 Role in neutrophil and NK cell function and pathogen defense.	153
27	_	NF1: Neurofibromin. neurofibromatosis.	154
44	47	NMDE4, Glutamate receptor ionotropic, NMDA 2D. Epileptic encephalopathy.	155
97	113	Obscurin: Heart disease.	156
26	39	SMCA4: Transcription activator BRG1. Esophageal, medulloblastoma, lung cancers.	157
62	113	SRRM2: Serine/arginine repetitive matrix protein 2. Thyroid carcinoma; Parkinson's disease.	158,159
15	_	STA13: StAR-related lipid transfer protein 13. Deleted in liver cancer 2 protein.	160
8	9	TGFB1: Transforming growth factor β-1 proprotein. Lupus nephritis in SLE Patients.	161
250	341	TITIN: Titin. Myocarditis, acute myocardial ischemia, cardiac arrest.	162
32	34	TRNK1: TPR and ankyrin repeat-containing protein 1. SLE. Neural development and differentiation.	163
12	-	TSP1: Thrombospondin-1. Inhibits tumor angiogenesis and suppresses tumor growth.	164
24	-	ZAN: Zonadhesin . Crucial role in sperm-zona adhesion. Sterility.	165
40	_	ZEP1: Zinc finger protein 40. Tum or-suppressive effects in prostate and nonsmall cell lung cancer.	166,167

Abbreviations: EBV, Epstein-Barr virus; DNA, deoxyribonucleic acid; MG, myasthenia gravis; MS, multiple sclerosis; SLE, systemic lupus erythematosus.

MSGGPM**GGRPG**GRGAPAVQQNIPSTLLQDHENQRLFEMLGRKCL TLATAVVQLYLALPPGAEHWTKEHCGAVCFVKDNPQKSYFIRLY GLQAGRLLWEQELYSQLVYSTPTPFFHTFAGDDCQAGLNFADED EAQAFRALVQEKIQKRNQRQSGDRRQLPPPPTPANEERRGGLPP LPLHPGGDQGGPPVGPLSLGLATVDIQNPDITSSRYRGLPAPGP SPADKKRSGKKKISK**ADIGAP**SGFKHVSHVGWDPQNGFDVNNLD P**DLRSL**FSRAGISEAQLTDAETSKLIYDFIEDQGGLEAVRQEMR RQE**PLPPPPPSR**GGNQLPRPPIVGGNKGRSG**PLPPVP**LGIAPP PPT**PRGPPPPGR**GG**PPPPPPP**ATGRSG**PLPPPPP**GAGGP**PMPPP** PPPPPPPSSGNGPAPPPLPPALVPAGGLAPGGGRGALLDQIRQ GIQLNKTPGAPESSALQPPPQSSEGLVGALMHVMQKRSRAIHSS DEGEDQAGDEDEDDEWDD

Fig. 1 Distribution of EBV epitope-derived peptides throughout WASP primary aa sequence. WASP sequence from Uniprot (http://www.uniprot. org/).<sup>51</sup> EBV epitope-derived peptides are underlined and bold marked. EBV, Epstein-Barr virus; WASP, Wiskott-Aldrich syndrome protein.

### **Discussion**

We summarize here the vast peptide platform that, with impressive mathematical unexpectedness, connects immunoreactive EBV epitopes and human proteins.

Quantitatively, **Table 1** shows that the peptide sharing does not obey to any theoretical probability expectations or constraints such as, for example, protein dimension. The case is best illustrated by the far upstream element-binding protein 2 (FUBP2; 711 aa) and the low-density lipoprotein receptor-related protein 1B (LRP1B; 4,599 aa). FUBP2 has 34 pentapeptides in common with the herpesviral proteome, whereas the much longer LRP1B shares 16 pentapeptides (**Table 4**). That is, a high number of shared pentapeptides can be found in a protein irrespective of the protein length.

aColumn A: number of shared peptides.

<sup>&</sup>lt;sup>b</sup>Column B: number of shared peptides including multiple occurrences.

<sup>&</sup>lt;sup>c</sup>Human proteins reported by UniProt entry names. Protein details, sequence, and aa length available at www.uniprot.org.

<sup>&</sup>lt;sup>d</sup>Further references on the function/associated disease are available at UniProt, OMIM, and PubMed resources.

Pathologically, the peptide sharing between the immunoreactive EBV epitopes and the human proteome implies the possibility of cross-reactions and of a consequent wide spectrum of diseases, from lymphomas and leukemias to diabetes and spermatogenesis (>Tables 3 and 4). From this point of view, **Tables 3** and **4** offer a scientific explanation of the clinical fact that EBV infection can trigger so many and so various diseases in so different and distant parts of the body. Moreover, given the number of human proteins involved in the sharing, the possibility of cross-reacting with a specific protein or group of proteins and inducing a specific disease or group of diseases will depend on the "when and where" the disease-associated protein(s) will be expressed. Consequently, the EBV diseasome will manifest with different diseases depending on the age of the subjects and on the immunological imprinting by previous pathogen infections, 168 thus explaining also why, once the immune system has been activated by EBV, some subjects will develop a lymphoma while other subjects contract diabetes or lupus or will die.

Immunologically, the vast peptide sharing between immunoreactive EBV epitopes and human proteins fails to support the theory of microbial or of human immunological specificity and nullifies the current concept of self-tolerance. Indeed, it was advanced in the "50s and still persists today the Burnet's hypothesis according to which self-tolerance is achieved by the so-called negative selection" of self-reactive lymphocytes. 169–171 That is, lymphocytes with specificity for peptide sequences that are expressed in the human host are hypothesized to be deleted from the immunological repertoire during fetal or early life to avoid self-reactivity and the consequent autoimmunity. Clearly, such a hypothesis breaks down in front of the pervasive peptide overlap between immunoreactive EBV epitopes and human proteins. If the "negative selection" assumptions were true, the self-reactive lymphocytes targeting the experimentally validated EBV epitopes described here and almost exclusively composed by peptides common to human proteins would have had to be eliminated from the immunological repertoire in the fetal life. It seems that the postulated deletion of potentially self-reactive lymphocytes did not occur. Similar results have been obtained analyzing hepatitis C virus and human papillomavirus immunoreactive epitopes. 172,173 Altogether, our data indicate that potentially self-reactive lymphocytes are regularly produced by the immune system. It seems that the immune system, under physiological conditions, does not engage reactions with self-proteins or pathogens just in virtue of their peptide commonality. As already discussed, 174–176 it seems that it is just the vast peptide commonality to confer or, better, to reify protein immunotolerance.

As a collateral note, we observe that, while **Tables 1** and **2** militate against the assumption of a "negative selection" of self-reactive lymphocytes, **Tables 3** and **4** also question the defensive role of the immune response. By definition, immune system attacks pathogenic enemies and protects self-entities. That is, it is assumed that the immune system is endowed with the capacity of discerning a pathogen antigen from a self-protein and of behaving consequentially by attacking the

"foes" and defending the "friends." Instead of being analyzed and defined as an aggregate of molecules organized into functional biological pathways, the immune system is considered as a "thinking entity" that sees, discriminates, decides, and then attacks. Against such an anthropomorphous view, the present mathematical and biochemical data document that pathogenic immune responses can routinely occur following infections, as already experimentally demonstrated. 177,178 Pathogenic autoantibodies—that are usually considered as rare phenomena due to the so-called "immunological holes" deriving from an incomplete negative selection of the self-reactive lymphocytes 169–171 or that, even, have been denied as pure fantasies 179—seem to be the rule.

► **Tables 3** and **4** show that anti-EBV immunoreactivity can hit a myriad of human proteins that, when (epi)genetically altered, can lead to cancers, autoimmune diseases, and even death. Such cross-reactive potential explains why higher the anti-EBV IgG antibody titer, worse may be the disease prognosis and faster the disease progression as described by a continuum of reports since the 1970s. <sup>16–34</sup> That is, autoimmunity is not a matter of "rare immunological holes," but it is intrinsic to the immune response that involves most of the human proteome by being most of the human proteome shared with microbial entities as a result of a long evolutionary path that from viruses and bacteria led to the eukaryotic cell. <sup>180</sup>

In conclusion, this study highlights the necessity of reviewing the hypothesis of the "negative selection" of self-reactive lymphocytes and, at the same time, emphasizes the importance of the "peptide uniqueness" concept to develop immunotherapies against EBV infection, and infections in general. Only immunotherapies based on peptides uniquely owned by the infectious agents would offer high specificity as well as the advantage of a lack of adverse events in the human host.<sup>39,181–183</sup>

Funding None.

Conflict of Interest

D.K. declares no conflicts. Y.S. is a medical consultant in vaccine compensation court, United States.

### References

- 1 Epstein MA, Achong BG, Barr YM. Virus particles in cultured lymphoblasts from Burkitt's lymphoma. Lancet 1964;1 (7335):702–703
- 2 Old LJ, Boyse EA, Oettgen HF, et al. Precipitating antibody in human serum to an antigen present in cultured Burkitt's lymphoma cells. Proc Natl Acad Sci U S A 1966;56(06):1699–1704
- 3 Henle G, Henle W, Clifford P, et al. Antibodies to Epstein-Barr virus in Burkitt's lymphoma and control groups. J Natl Cancer Inst 1969;43(05):1147–1157
- 4 Gunvén P, Klein G, Henle G, Henle W, Clifford P. Epstein-Barr virus in Burkitt's lymphoma and nasopharyngeal carcinoma. Antibodies to EBV associated membrane and viral capsid antigens in Burkitt lymphoma patients. Nature 1970;228(5276):1053–1056
- 5 Harley B, Shivapathasundram G, Astradsson A, Muthurajah V, Wickremesekera A. An unusual presentation of cerebellar lymphoma. J Clin Neurosci 2018;57:177–180

- 6 Yahia ZA, Adam AA, Elgizouli M, et al. Epstein Barr virus: a prime candidate of breast cancer aetiology in Sudanese patients. Infect Agent Cancer 2014;9(01):9
- 7 Fołtyn S, Strycharz-Dudziak M, Drop B, Boguszewska A, Polz-Dacewicz M. Serum EBV antibodies and LMP-1 in Polish patients with oropharyngeal and laryngeal cancer. Infect Agent Cancer 2017:12:31
- 8 Khammissa RA, Fourie J, Chandran R, Lemmer J, Feller L. Epstein-Barr virus and its association with Oral Hairy Leukoplakia: a short review. Int J Dent 2016;2016:4941783
- 9 Kivity S, Agmon-Levin N, Blank M, Shoenfeld Y. Infections and autoimmunity—friends or foes? Trends Immunol 2009;30(08): 409–414
- 10 Maślińska M. The role of Epstein-Barr virus infection in primary Sjögren's syndrome. Curr Opin Rheumatol 2019;31(05): 475–483
- 11 Dittfeld A, Gwizdek K, Michalski M, Wojnicz R. A possible link between the Epstein-Barr virus infection and autoimmune thyroid disorders. Cent Eur J Immunol 2016;41(03):297–301
- 12 Woulfe JM, Gray MT, Gray DA, Munoz DG, Middeldorp JM. Hypothesis: a role for EBV-induced molecular mimicry in Parkinson's disease. Parkinsonism Relat Disord 2014;20(07): 685-694
- 13 Di Loreto S, Fabiano C, Nigro G. High prevalence of streptococcal or Epstein-Barr virus infections in children with acute non-septic monoarthritis. New Microbiol 2014;37(01):81–86
- 14 Hasegawa D, Kaji M, Takeda H, et al. Fatal degeneration of specialized cardiac muscle associated with chronic active Epstein-Barr virus infection. Pediatr Int 2009;51(06):846–848
- 15 Henle W, Hummeler K, Henle G. Antibody coating and agglutination of virus particles separated from the EB3 line of Burkitt lymphoma cells. J Bacteriol 1966;92(01):269–271
- 16 Henle W, Henle G, Gunvén P, Klein G, Clifford P, Singh S. Patterns of antibodies to Epstein-Barr virus-induced early antigens in Burkitt's lymphoma. Comparison of dying patients with longterm survivors. J Natl Cancer Inst 1973;50(05):1163–1173
- 17 Asito AS, Piriou E, Odada PS, et al. Elevated anti-Zta IgG levels and EBV viral load are associated with site of tumor presentation in endemic Burkitt's lymphoma patients: a case control study. Infect Agent Cancer 2010;5:13
- 18 Liu MY, Shih YY, Chou SP, et al. Antibody against the Epstein-Barr virus BHRF1 protein, a homologue of Bcl-2, in patients with nasopharyngeal carcinoma. J Med Virol 1998;56(03):179–185
- 19 Cheng WM, Chan KH, Chen HL, et al. Assessing the risk of nasopharyngeal carcinoma on the basis of EBV antibody spectrum. Int J Cancer 2002;97(04):489–492
- 20 Fachiroh J, Schouten T, Hariwiyanto B, et al. Molecular diversity of Epstein-Barr virus IgG and IgA antibody responses in nasopharyngeal carcinoma: a comparison of Indonesian, Chinese, and European subjects. J Infect Dis 2004;190(01):53–62
- 21 Guo X, Li T, Li F, et al. Intermittent abortive reactivation of Epstein-Barr virus during the progression of nasopharyngeal cancer as indicated by elevated antibody levels. Oral Oncol 2019; 93:85–90
- 22 Johansson B, Klein G, Henle W, Henle G. Epstein-Barr virus (EBV)-associated antibody patterns in malignant lymphoma and leukemia. I. Hodgkin's disease. Int J Cancer 1970;6(03): 450–462
- 23 Mueller N, Evans A, Harris NL, et al. Hodgkin's disease and Epstein-Barr virus. Altered antibody pattern before diagnosis. N Engl J Med 1989;320(11):689–695
- 24 Mueller NE, Lennette ET, Dupnik K, Birmann BM. Antibody titers against EBNA1 and EBNA2 in relation to Hodgkin lymphoma and history of infectious mononucleosis. Int J Cancer 2012;130(12): 2886–2891
- 25 Schetter AJ, You WC, Lennette ET, Gail MT, Rabkin CS. Association of Epstein-Barr virus antibody levels with precancerous gastric lesions in a high-risk cohort. Cancer Sci 2008;99(02):350–354

- 26 Sumaya CV, Myers LW, Ellison GW, Ench Y. Increased prevalence and titer of Epstein-Barr virus antibodies in patients with multiple sclerosis. Ann Neurol 1985;17(04):371–377
- 27 Farrell RA, Antony D, Wall GR, et al. Humoral immune response to EBV in multiple sclerosis is associated with disease activity on MRI. Neurology 2009;73(01):32–38
- 28 Jakimovski D, Ramanathan M, Weinstock-Guttman B, et al. Higher EBV response is associated with more severe gray matter and lesion pathology in relapsing multiple sclerosis patients: a case-controlled magnetization transfer ratio study. Mult Scler 2020;26(03):322–332
- 29 Lossius A, Johansen JN, Torkildsen Ø, Vartdal F, Holmøy T. Epstein-Barr virus in systemic lupus erythematosus, rheumatoid arthritis and multiple sclerosis—association and causation. Viruses 2012;4(12):3701–3730
- 30 Alspaugh MA, Henle G, Lennette ET, Henle W. Elevated levels of antibodies to Epstein-Barr virus antigens in sera and synovial fluids of patients with rheumatoid arthritis. J Clin Invest 1981;67 (04):1134–1140
- 31 Toussirot E, Roudier J. Epstein-Barr virus in autoimmune diseases. Best Pract Res Clin Rheumatol 2008;22(05):883–896
- 32 Cavalcante P, Serafini B, Rosicarelli B, et al. Epstein-Barr virus persistence and reactivation in myasthenia gravis thymus. Ann Neurol 2010;67(06):726–738
- 33 Csuka D, Banati M, Rozsa C, Füst G, Illes Z. High anti-EBNA-1 IgG levels are associated with early-onset myasthenia gravis. Eur J Neurol 2012;19(06):842–846
- 34 Draborg AH, Lydolph MC, Westergaard M, et al. Elevated concentrations of serum immunoglobulin free light chains in systemic lupus erythematosus patients in relation to disease activity, inflammatory status, B cell activity and Epstein-Barr virus antibodies. PLoS One 2015;10(09):e0138753
- 35 Martínez-A C, Marcos MA, de la Hera A, et al. Immunological consequences of HIV infection: advantage of being low responder casts doubts on vaccine development. Lancet 1988;1 (8583):454–457
- 36 Fachiroh J, Paramita DK, Hariwiyanto B, et al. Single-assay combination of Epstein-Barr Virus (EBV) EBNA1- and viral capsid antigen-p18-derived synthetic peptides for measuring anti-EBV immunoglobulin G (IgG) and IgA antibody levels in sera from nasopharyngeal carcinoma patients: options for field screening. J Clin Microbiol 2006;44(04):1459–1467
- 37 Paramita DK, Fachiroh J, Haryana SM, Middeldorp JM. Evaluation of commercial EBV RecombLine assay for diagnosis of nasopharyngeal carcinoma. J Clin Virol 2008;42(04):343–352
- 38 Levin LI, Chang ET, Ambinder RF, et al. Atypical prediagnosis Epstein-Barr virus serology restricted to EBV-positive Hodgkin lymphoma. Blood 2012;120(18):3750–3755
- 39 Capone G, Calabrò M, Lucchese G, et al. Peptide matching between Epstein-Barr virus and human proteins. Pathog Dis 2013;69(03):205–212
- 40 Capone G, Fasano C, Lucchese G, Calabrò M, Kanduc D. EBV-associated cancer and autoimmunity: searching for therapies. Vaccines (Basel) 2015;3(01):74–89
- 41 Calabrò M. Epstein Barr Virus Immunoevasion. Sequence Similarity between Human Proteins and Epstein Barr Virus [PhD thesis]. Bari: University of Bari; 2015
- 42 Kanduc D. Proteome-wide Epstein-Barr virus analysis of peptide sharing with human systemic lupus erythematosus autoantigens. Isr Med Assoc J 2019;21(07):444–448
- 43 Reddehase MJ, Rothbard JB, Koszinowski UH. A pentapeptide as minimal antigenic determinant for MHC class I-restricted T lymphocytes. Nature 1989;337(6208):651–653
- 44 Zeng W, Pagnon J, Jackson DC. The C-terminal pentapeptide of LHRH is a dominant B cell epitope with antigenic and biological function. Mol Immunol 2007;44(15):3724–3731
- 45 Kanduc D. Homology, similarity, and identity in peptide epitope immunodefinition. J Pept Sci 2012;18(08):487–494

- 46 Raychaudhuri S, Sandor C, Stahl EA, et al. Five amino acids in three HLA proteins explain most of the association between MHC and seropositive rheumatoid arthritis. Nat Genet 2012;44 (03):291–296
- 47 Kanduc D. Pentapeptides as minimal functional units in cell biology and immunology. Curr Protein Pept Sci 2013;14(02): 111–120
- 48 Hao SS, Zong MM, Zhang Z, et al. The inducing roles of the new isolated bursal hexapeptide and pentapeptide on the immune response of AIV vaccine in mice. Protein Pept Lett 2019;26(07): 542–549
- 49 Vita R, Mahajan S, Overton JA, et al. The Immune Epitope Database (IEDB): 2018 update. Nucleic Acids Res 2019;47(D1): D339–D343
- 50 Chen C, Li Z, Huang H, Suzek BE, Wu CH; UniProt Consortium. A fast Peptide Match service for UniProt Knowledgebase. Bioinformatics 2013;29(21):2808–2809
- 51 UniProt Consortium. UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res 2019;47(D1):D506–D515
- 52 Grønbaek K, Worm J, Ralfkiaer E, Ahrenkiel V, Hokland P, Guldberg P. ATM mutations are associated with inactivation of the ARF-TP53 tumor suppressor pathway in diffuse large B-cell lymphoma. Blood 2002;100(04):1430–1437
- 53 Yan J, Nie K, Mathew S, et al. Inactivation of BANK1 in a novel IGH-associated translocation t(4;14)(q24;q32) suggests a tumor suppressor role in B-cell lymphoma. Blood Cancer J 2014; 4:e215
- 54 Dam EM, Habib T, Chen J, et al. The BANK1 SLE-risk variants are associated with alterations in peripheral B cell signaling and development in humans. Clin Immunol 2016;173:171–180
- 55 Kurosawa N, Fujimoto R, Ozawa T, Itoyama T, Sadamori N, Isobe M. Reduced level of the BCL11B protein is associated with adult T-cell leukemia/lymphoma. PLoS One 2013;8(01):e55147
- 56 Punwani D, Zhang Y, Yu J, et al. Multisystem anomalies in severe combined immunodeficiency with mutant BCL11B. N Engl J Med 2016;375(22):2165–2176
- 57 Gutierrez A, Kentsis A, Sanda T, et al. The BCL11B tumor suppressor is mutated across the major molecular subtypes of T-cell acute lymphoblastic leukemia. Blood 2011;118(15): 4169–4173
- 58 Greipp PT, Smoley SA, Viswanatha DS, et al. Patients with chronic lymphocytic leukaemia and clonal deletion of both 17p13.1 and 11q22.3 have a very poor prognosis. Br J Haematol 2013;163 (03):326–333
- 59 Xu L, Li X, Chu ES, et al. Epigenetic inactivation of BCL6B, a novel functional tumour suppressor for gastric cancer, is associated with poor survival. Gut 2012;61(07):977–985
- 60 Yang Q, Gao J, Xu L, Zeng Z, Sung JJ, Yu J. Promoter hypermethylation of BCL6B gene is a potential plasma DNA biomarker for gastric cancer. Biomarkers 2013;18(08):721–725
- 61 Deng J, Liang H, Dong Q, et al. The survival decrease in gastric cancer is associated with the methylation of B-cell CLL/lymphoma 6 member B promoter. Open Biol 2014;4(07): 140067
- 62 Wang J, Dong L, Xu L, et al. B cell CLL/lymphoma 6 member B inhibits hepatocellular carcinoma metastases in vitro and in mice. Cancer Lett 2014;355(02):192–200
- 63 Esmailzadeh S, Huang Y, Su MW, Zhou Y, Jiang X. BIN1 tumor suppressor regulates Fas/Fas ligand-mediated apoptosis through c-FLIP in cutaneous T-cell lymphoma. Leukemia 2015;29(06): 1402–1413
- 64 Fu L, Gao Z, Zhang X, et al. Frequent concomitant epigenetic silencing of the stress-responsive tumor suppressor gene CADM1, and its interacting partner DAL-1 in nasal NK/T-cell lymphoma. Int J Cancer 2009;124(07):1572–1578
- 65 Martinelli S, Checquolo S, Consoli F, et al. Loss of CBL E3-ligase activity in B-lineage childhood acute lymphoblastic leukaemia. Br J Haematol 2012;159(01):115–119

- 66 Fang M, Pak ML, Chamberlain L, Xing W, Yu H, Green MR. The CREB coactivator CRTC2 is a lymphoma tumor suppressor that preserves genome integrity through transcription of DNA mismatch repair genes. Cell Reports 2015;11(09):1350–1357
- 67 Shawky SA, El-Borai MH, Khaled HM, et al. The prognostic impact of hypermethylation for a panel of tumor suppressor genes and cell of origin subtype on diffuse large B-cell lymphoma. Mol Biol Rep 2019;46(04):4063–4076
- 68 Küçük C, Hu X, Jiang B, et al. Global promoter methylation analysis reveals novel candidate tumor suppressor genes in natural killer cell lymphoma. Clin Cancer Res 2015;21(07): 1699–1711
- 69 Oricchio E, Nanjangud G, Wolfe AL, et al. The Eph-receptor A7 is a soluble tumor suppressor for follicular lymphoma. Cell 2011;147 (03):554–564
- 70 Laginestra MA, Cascione L, Motta G, et al. Whole exome sequencing reveals mutations in FAT1 tumor suppressor gene clinically impacting on peripheral T-cell lymphoma not otherwise specified. Mod Pathol 2020;33(02):179–187
- 71 Hu X, Zhai Y, Kong P, et al. FAT1 prevents epithelial mesenchymal transition (EMT) via MAPK/ERK signaling pathway in esophageal squamous cell cancer. Cancer Lett 2017;397:83–93
- 72 Stöcklein H, Smardova J, Macak J, et al. Detailed mapping of chromosome 17p deletions reveals HIC1 as a novel tumor suppressor gene candidate telomeric to TP53 in diffuse large B-cell lymphoma. Oncogene 2008;27(18):2613–2625
- 73 Chang S, Yim S, Park H. The cancer driver genes IDH1/2, JARID1C/KDM5C, and UTX/KDM6A: crosstalk between histone demethylation and hypoxic reprogramming in cancer metabolism. Exp Mol Med 2019;51(06):1–17
- 74 Schulz WA, Lang A, Koch J, Greife A. The histone demethylase UTX/KDM6A in cancer: progress and puzzles. Int J Cancer 2019; 145(03):614–620
- 75 Yang W, Ernst P. Distinct functions of histone H3, lysine 4 methyltransferases in normal and malignant hematopoiesis. Curr Opin Hematol 2017;24(04):322–328
- 76 Fernandez-Pol S, Ma L, Joshi RP, Arber DA. A survey of somatic mutations in 41 genes in a cohort of T-cell lymphomas identifies frequent mutations in genes involved in epigenetic modification. Appl Immunohistochem Mol Morphol 2019;27(06): 416–422
- 77 Froimchuk E, Jang Y, Ge K. Histone H3 lysine 4 methyltransferase KMT2D. Gene 2017;627:337–342
- 78 Zhang J, Dominguez-Sola D, Hussein S, et al. Disruption of KMT2D perturbs germinal center B cell development and promotes lymphomagenesis. Nat Med 2015;21(10):1190–1198
- 79 Johansson P, Klein-Hitpass L, Grabellus F, et al. Recurrent mutations in NF-κB pathway components, KMT2D, and NOTCH1/2 in ocular adnexal MALT-type marginal zone lymphomas. Oncotarget 2016;7(38):62627–62639
- 80 Wang Y, Cui H, Schroering A, et al. NF-kappa B2 p100 is a proapoptotic protein with anti-oncogenic function. Nat Cell Biol 2002;4(11):888–893
- 81 Keller U, Huber J, Nilsson JA, et al. Myc suppression of Nfkb2 accelerates lymphomagenesis. BMC Cancer 2010;10:348
- 82 Thakur S, Lin HC, Tseng WT, et al. Rearrangement and altered expression of the NFKB-2 gene in human cutaneous T-lymphoma cells. Oncogene 1994;9(08):2335–2344
- 83 Klemann C, Camacho-Ordonez N, Yang L, et al. Clinical and immunological phenotype of patients with primary immunodeficiency due to damaging mutations in NFKB2. Front Immunol 2019;10:297
- 84 Zhang T, Ma J, Nie K, et al. Hypermethylation of the tumor suppressor gene PRDM1/Blimp-1 supports a pathogenetic role in EBV-positive Burkitt lymphoma. Blood Cancer J 2014;4:e261
- 85 Johansson P, Klein-Hitpass L, Choidas A, et al. SAMHD1 is recurrently mutated in T-cell prolymphocytic leukemia. Blood Cancer J 2018;8(01):11

- 86 Geli J, Kiss N, Kogner P, Larsson C. Suppression of RIZ in biologically unfavourable neuroblastomas. Int J Oncol 2010;37(05): 1323-1330
- 87 Xie W, Li X, Chen X, Huang S, Huang S. Decreased expression of PRDM2 (RIZ1) and its correlation with risk stratification in patients with myelodysplastic syndrome. Br J Haematol 2010; 150(02):242-244
- 88 Sasaki O, Meguro K, Tohmiya Y, Funato T, Shibahara S, Sasaki T. Altered expression of retinoblastoma protein-interacting zinc finger gene, RIZ, in human leukaemia. Br J Haematol 2002;119 (04):940-948
- 89 Cimmino L, Dawlaty MM, Ndiaye-Lobry D, et al. TET1 is a tumor suppressor of hematopoietic malignancy. Nat Immunol 2015;16 (06):653-662
- 90 Kosmider O, Gelsi-Boyer V, Ciudad M, et al; Groupe Francophone des Myélodysplasies. TET2 gene mutation is a frequent and adverse event in chronic myelomonocytic leukemia. Haematologica 2009;94(12):1676-1681
- 91 Menotti M, Ambrogio C, Cheong TC, et al. Wiskott-Aldrich syndrome protein (WASP) is a tumor suppressor in T cell lymphoma. Nat Med 2019;25(01):130-140
- 92 Morin RD, Mendez-Lago M, Mungall AJ, et al. Frequent mutation of histone-modifying genes in non-Hodgkin lymphoma. Nature 2011;476(7360):298-303
- 93 da Silva Almeida AC, Abate F, Khiabanian H, et al. The mutational landscape of cutaneous T cell lymphoma and Sézary syndrome. Nat Genet 2015;47(12):1465–1470
- 94 Rabello DDA, Ferreira VDDS, Berzoti-Coelho MG, et al. MLL2/ KMT2D and MLL3/KMT2C expression correlates with disease progression and response to imatinib mesylate in chronic myeloid leukemia. Cancer Cell Int 2018;18:26
- 95 Sahm F, Jakobiec FA, Meyer J, et al. Somatic mutations of DICER1 and KMT2D are frequent in intraocular medulloepitheliomas. Genes Chromosomes Cancer 2016;55(05):418-427
- 96 Augert A, Zhang Q, Bates B, et al. Small cell lung cancer exhibits frequent inactivating mutations in the histone methyltransferase KMT2D/MLL2: CALGB 151111 (Alliance). J Thorac Oncol 2017;12(04):704-713
- 97 Sun P, Wu T, Sun X, et al. KMT2D inhibits the growth and metastasis of bladder cancer cells by maintaining the tumor suppressor genes. Biomed Pharmacother 2019;115:108924
- 98 Ding B, Yan L, Zhang Y, et al. Analysis of the role of mutations in the KMT2D histone lysine methyltransferase in bladder cancer. FEBS Open Bio 2019;9(04):693-706
- 99 Ardeshir-Larijani F, Bhateja P, Lipka MB, Sharma N, Fu P, Dowlati A. KMT2D mutation is associated with poor prognosis in nonsmall-cell lung cancer. Clin Lung Cancer 2018;19(04):e489-e501
- 100 Ng SB, Bigham AW, Buckingham KJ, et al. Exome sequencing identifies MLL2 mutations as a cause of Kabuki syndrome. Nat Genet 2010;42(09):790-793
- 101 Cheon CK, Ko JM. Kabuki syndrome: clinical and molecular characteristics. Korean J Pediatr 2015;58(09):317-324
- 102 Yap KL, Johnson AEK, Fischer D, et al. Congenital hyperinsulinism as the presenting feature of Kabuki syndrome: clinical and molecular characterization of 9 affected individuals. Genet Med 2019;21(01):233-242
- 103 Kurahashi N, Miyake N, Mizuno S, et al. Characteristics of epilepsy in patients with Kabuki syndrome with KMT2D mutations. Brain Dev 2017;39(08):672-677
- 104 Scala M, Morana G, Sementa AR, et al. Aggressive desmoid fibromatosis in Kabuki syndrome: expanding the tumor spectrum. Pediatr Blood Cancer 2019;66(09):e27831
- 105 Margot H, Boursier G, Duflos C, et al. Immunopathological manifestations in Kabuki syndrome: a registry study of 177 individuals. Genet Med 2020;22(01):181-188
- 106 Arsov T, Sestan M, Cekada N, et al. Systemic lupus erythematosus: a new autoimmune disorder in Kabuki syndrome. Eur J Med Genet 2019;62(06):103538

- 107 Porntaveetus T, Abid MF, Theerapanon T, et al. Expanding the oro-dental and mutational spectra of Kabuki Syndrome and expression of KMT2D and KDM6A in human tooth germs. Int J Biol Sci 2018;14(04):381-389
- 108 Herz HM. Enhancer deregulation in cancer and other diseases. BioEssays 2016;38(10):1003-1015
- 109 Sullivan KE, Mullen CA, Blaese RM, Winkelstein JA. A multiinstitutional survey of the Wiskott-Aldrich syndrome. J Pediatr 1994; 125(6 Pt 1):876-885
- 110 Thrasher AJ, Burns SO. WASP: a key immunological multitasker. Nat Rev Immunol 2010;10(03):182-192
- 111 Rivers E, Thrasher AJ. Wiskott-Aldrich syndrome protein: emerging mechanisms in immunity. Eur J Immunol 2017;47(11): 1857-1866
- 112 MacCarthy-Morrogh L, Gaspar HB, Wang YC, et al. Absence of expression of the Wiskott-Aldrich syndrome protein in peripheral blood cells of Wiskott-Aldrich syndrome patients. Clin Immunol Immunopathol 1998;88(01):22-27
- 113 Candotti F. Clinical manifestations and pathophysiological mechanisms of the Wiskott-Aldrich Syndrome. J Clin Immunol 2018;38(01):13-27
- 114 Marangoni F, Trifari S, Scaramuzza S, et al. WASP regulates suppressor activity of human and murine CD4(+)CD25(+) FOXP3(+) natural regulatory T cells. J Exp Med 2007;204(02): 369-380
- 115 Ohya T, Yanagimachi M, Iwasawa K, et al. Childhood-onset inflammatory bowel diseases associated with mutation of Wiskott-Aldrich syndrome protein gene. World J Gastroenterol 2017;23(48):8544-8552
- 116 Bouma G, Carter NA, Recher M, et al. Exacerbated experimental arthritis in Wiskott-Aldrich syndrome protein deficiency: modulatory role of regulatory B cells. Eur J Immunol 2014;44(09): 2692-2702
- 117 Huang GZ, Lo YL. Correlation between acetylcholine receptor antibody levels and thymic pathology in myasthenia gravis: a review. J Clin Neuromuscul Dis 2013;14(04):209-217
- 118 Cork SM, Kaur B, Devi NS, et al. A proprotein convertase/MMP-14 proteolytic cascade releases a novel 40 kDa vasculostatin from tumor suppressor BAI1. Oncogene 2012;31(50):5144-5152
- 119 Kaur B, Cork SM, Sandberg EM, et al. Vasculostatin inhibits intracranial glioma growth and negatively regulates in vivo angiogenesis through a CD36-dependent mechanism. Cancer Res 2009;69(03):1212-1220
- 120 Soh RYZ, Lim JP, Samy RP, Chua PJ, Bay BH. A-kinase anchor protein 12 (AKAP12) inhibits cell migration in breast cancer. Exp Mol Pathol 2018;105(03):364-370
- 121 Bortlik M, Vitkova I, Papezova M, et al. Deficiency of adenomatous polyposis coli protein in sporadic colorectal adenomas and its associations with clinical phenotype and histology. World I Gastroenterol 2006;12(24):3901-3905
- 122 Carson DJ, Santoro IM, Groden J. Isoforms of the APC tumor suppressor and their ability to inhibit cell growth and tumorigenicity. Oncogene 2004;23(42):7144-7148
- 123 Daly CS, Shaw P, Ordonez LD, et al. Functional redundancy between Apc and Apc2 regulates tissue homeostasis and prevents tumorigenesis in murine mammary epithelium. Oncogene 2017;36(13):1793-1803
- 124 Ying X, Li-ya Q, Feng Z, Yin W, Ji-hong L. MiR-939 promotes the proliferation of human ovarian cancer cells by repressing APC2 expression. Biomed Pharmacother 2015;71:64-69
- 125 Tiberi L, Bonnefont J, van den Ameele J, et al. A BCL6/BCOR/SIRT1 complex triggers neurogenesis and suppresses medulloblastoma by repressing Sonic Hedgehog signaling. Cancer Cell 2014;26 (06):797-812
- 126 Dragon-Durey MA, Quartier P, Frémeaux-Bacchi V, et al. Molecular basis of a selective C1s deficiency associated with early onset multiple autoimmune diseases. J Immunol 2001;166(12): 7612-7616

- 127 Ohno K, Tsujino A, Brengman JM, et al. Choline acetyltransferase mutations cause myasthenic syndrome associated with episodic apnea in humans. Proc Natl Acad Sci U S A 2001;98(04): 2017-2022
- 128 Lokki ML, Circolo A, Ahokas P, Rupert KL, Yu CY, Colten HR. Deficiency of human complement protein C4 due to identical frameshift mutations in the C4A and C4B genes. J Immunol 1999; 162(06):3687-3693
- 129 Nyberg P, Xie L, Sugimoto H, et al. Characterization of the antiangiogenic properties of arresten, an alpha1beta1 integrin-dependent collagen-derived tumor suppressor. Exp Cell Res 2008; 314(18):3292-3305
- 130 Shaikh S, Shettigar SKG, Kumar S, Kantharia S, Kurva J, Cherian S. Novel mutation in Cul7 gene in a family diagnosed with 3M syndrome. J Genet 2019;98:21
- 131 Hedrick L, Cho KR, Fearon ER, Wu TC, Kinzler KW, Vogelstein B. The DCC gene product in cellular differentiation and colorectal tumorigenesis. Genes Dev 1994;8(10):1174-1183
- 132 Mollenhauer J, Wiemann S, Scheurlen W, et al. DMBT1, a new member of the SRCR superfamily, on chromosome 10q25.3-26.1 is deleted in malignant brain tumours. Nat Genet 1997;17(01):
- 133 Takeshita H, Sato M, Shiwaku HO, et al. Expression of the DMBT1 gene is frequently suppressed in human lung cancer. Jpn J Cancer Res 1999;90(09):903-908
- 134 Baldari U, Raccagni AA, Celli B, Righini MG. Chronic bullous disease of childhood following Epstein-Barr virus seroconversion: a case report. Clin Exp Dermatol 1996;21(02):123-126
- 135 Sugi T, Hashimoto T, Hibi T, Nishikawa T. Production of human monoclonal anti-basement membrane zone (BMZ) antibodies from a patient with bullous pemphigoid (BP) by Epstein-Barr virus transformation. Analyses of the heterogeneity of anti-BMZ antibodies in BP sera using them. J Clin Invest 1989;84(04): 1050-1055
- 136 Huang FY, Wong DK, Tsui VW, et al. Targeted genomic profiling identifies frequent deleterious mutations in FAT4 and TP53 genes in HBV-associated hepatocellular carcinoma. BMC Cancer 2019:19(01):789
- 137 Sun H, Zhou H, Zhang Y, et al. Aberrant methylation of FAT4 and SOX11 in peripheral blood leukocytes and their association with gastric cancer risk. J Cancer 2018;9(13):2275-2283
- 138 Lu YJ, Wu CS, Li HP, et al. Aberrant methylation impairs low density lipoprotein receptor-related protein 1B tumor suppressor function in gastric cancer. Genes Chromosomes Cancer 2010; 49(05):412-424
- 139 Abuzzahab MJ, Schneider A, Goddard A, et al; Intrauterine Growth Retardation (IUGR) Study Group. IGF-I receptor mutations resulting in intrauterine and postnatal growth retardation. N Engl | Med 2003;349(23):2211-2222
- 140 Longo N, Wang Y, Pasquali M. Progressive decline in insulin levels in Rabson-Mendenhall syndrome. J Clin Endocrinol Metab 1999; 84(08):2623-2629
- 141 Kanatsuna N, Delli A, Andersson C, et al. Doubly reactive INS-IGF2 autoantibodies in children with newly diagnosed autoimmune (type 1) diabetes. Scand J Immunol 2015;82 (04):361-369
- 142 Tanokashira D, Fukuokaya W, Taguchi A. Involvement of insulin receptor substrates in cognitive impairment and Alzheimer's disease. Neural Regen Res 2019;14(08):1330-1334
- 143 Norquay LD, D'Aquino KE, Opare-Addo LM, et al. Insulin receptor substrate-2 in beta-cells decreases diabetes in nonobese diabetic mice. Endocrinology 2009;150(10):4531-4540
- 144 Arai T, Hashimoto H, Kawai K, et al. Fulminant type 1 diabetes mellitus observed in insulin receptor substrate 2 deficient mice. Clin Exp Med 2008;8(02):93-99
- 145 Vallianatos CN, Iwase S. Disrupted intricacy of histone H3K4 methylation in neurodevelopmental disorders. Epigenomics 2015;7(03):503-519

- 146 Dai B, Huang H, Guan F, et al. Histone demethylase KDM5A inhibits glioma cells migration and invasion by down regulating ZEB1. Biomed Pharmacother 2018;99:72-80
- 147 Gaidamakov S, Maximova OA, Chon H, et al. Targeted deletion of the gene encoding the La autoantigen (Sjögren's syndrome antigen B) in B cells or the frontal brain causes extensive tissue loss. Mol Cell Biol 2014;34(01):123-131
- 148 Cam JA, Zerbinatti CV, Knisely JM, Hecimovic S, Li Y, Bu G. The low density lipoprotein receptor-related protein 1B retains beta-amyloid precursor protein at the cell surface and reduces amyloid-beta peptide production. J Biol Chem 2004;279(28):29639-29646
- 149 Herrendorff R, Hänggi P, Pfister H, et al. Selective in vivo removal of pathogenic anti-MAG autoantibodies, an antigen-specific treatment option for anti-MAG neuropathy. Proc Natl Acad Sci U S A 2017;114(18):E3689-E3698
- 150 Ramanathan S, Dale RC, Brilot F. Anti-MOG antibody: the history, clinical phenotype, and pathogenicity of a serum biomarker for demyelination. Autoimmun Rev 2016;15(04):307-324
- 151 Duncan GJ, Plemel JR, Assinck P, et al. Myelin regulatory factor drives remyelination in multiple sclerosis. Acta Neuropathol 2017;134(03):403-422
- 152 Shi Y, Shao Q, Li Z, et al. Myt1L promotes differentiation of oligodendrocyte precursor cells and is necessary for remyelination after lysolecithin-induced demyelination. Neurosci Bull 2018;34(02):247-260
- 153 Sowerby JM, Thomas DC, Clare S, et al. NBEAL2 is required for neutrophil and NK cell function and pathogen defense. J Clin Invest 2017;127(09):3521-3526
- 154 Abramowicz A, Gos M. Neurofibromin in neurofibromatosis type 1-mutations in NF1gene as a cause of disease. Dev Period Med 2014;18(03):297-306
- 155 Li D, Yuan H, Ortiz-Gonzalez XR, et al. GRIN2D recurrent de novo dominant mutation causes a severe epileptic encephalopathy treatable with NMDA receptor channel blockers. Am J Hum Genet 2016;99(04):802-816
- 156 Grogan A, Kontrogianni-Konstantopoulos A. Unraveling obscurins in heart disease. Pflugers Arch 2019;471(05):735-743
- Schneppenheim R, Frühwald MC, Gesk S, et al. Germline nonsense mutation and somatic inactivation of SMARCA4/BRG1 in a family with rhabdoid tumor predisposition syndrome. Am J Hum Genet 2010;86(02):279-284
- 158 Tomsic J, He H, Akagi K, et al. A germline mutation in SRRM2, a splicing factor gene, is implicated in papillary thyroid carcinoma predisposition. Sci Rep 2015;5:10566
- 159 Shehadeh LA, Yu K, Wang L, et al. SRRM2, a potential blood biomarker revealing high alternative splicing in Parkinson's disease. PLoS One 2010;5(02):e9104
- 160 Ching YP, Wong CM, Chan SF, et al. Deleted in liver cancer (DLC) 2 encodes a RhoGAP protein with growth suppressor function and is underexpressed in hepatocellular carcinoma. J Biol Chem 2003;278(12):10824-10830
- 161 Rashad NM, El-Shabrawy RM, Said D, El-Shabrawy SM, Emad G. Serum levels of transforming growth factor beta -1 (TGF-β1) as an early no invasive marker for diagnosis of lupus nephritis in systemic lupus erythematosus patients. Egypt J Immunol 2019; 26(01):31-42
- 162 Pérez-Serra A, Toro R, Sarquella-Brugada G, et al. Genetic basis of dilated cardiomyopathy. Int J Cardiol 2016;224:461–472
- 163 Jiang X, Detera-Wadleigh SD, Akula N, et al. Sodium valproate rescues expression of TRANK1 in iPSC-derived neural cells that carry a genetic variant associated with serious mental illness. Mol Psychiatry 2019;24(04):613-624
- 164 Miyanaga K, Kato Y, Nakamura T, et al. Expression and role of thrombospondin-1 in colorectal cancer. Anticancer Res 2002;22
- 165 Tardif S, Cormier N. Role of zonadhesin during sperm-egg interaction: a species-specific acrosomal molecule with multiple functions. Mol Hum Reprod 2011;17(11):661-668

- 166 Ghosh AK, Steele R, Ray RB. Carboxyl-terminal repressor domain of MBP-1 is sufficient for regression of prostate tumor growth in nude mice. Cancer Res 2005;65(03):718–721
- 167 Ghosh AK, Steele R, Ryerse J, Ray RB. Tumor-suppressive effects of MBP-1 in non-small cell lung cancer cells. Cancer Res 2006;66 (24):11907–11912
- 168 Kanduc D, Shoenfeld Y. Inter-pathogen peptide sharing and the original antigenic sin: solving a paradox. Open Immunol J 2018; 8:16–27
- 169 Cohn M. Two unresolved problems facing models of the selfnonself discrimination. J Theor Biol 2015;387:31–38
- 170 Rose NR. Negative selection, epitope mimicry and autoimmunity. Curr Opin Immunol 2017;49:51–55
- 171 Rose NR. Learning from myocarditis: mimicry, chaos and black holes. F1000Prime Rep 2014;6:25
- 172 Kanduc D. From hepatitis C virus immunoproteomics to rheumatology via cross-reactivity in one table. Curr Opin Rheumatol 2019;31(05):488–492
- 173 Kanduc D, Shoenfeld Y. Human papillomavirus epitope mimicry and autoimmunity: the molecular truth of peptide sharing. Pathobiology 2019;86(5–6):285–295
- 174 Kanduc D. HCV: written in our DNA. Self Nonself 2011;2(02):108–113
- 175 Kanduc D. The self/nonself issue: a confrontation between proteomes. Self Nonself 2010;1(03):255–258

- 176 Kanduc D. Immunogenicity, immunopathogenicity, and immunotolerance in one graph. Anticancer Agents Med Chem 2015;15 (10):1264–1268
- 177 Ray SK, Putterman C, Diamond B. Pathogenic autoantibodies are routinely generated during the response to foreign antigen: a paradigm for autoimmune disease. Proc Natl Acad Sci U S A 1996; 93(05):2019–2024
- 178 Ruiz JT, Luján L, Blank M, Shoenfeld Y. Adjuvants- and vaccinesinduced autoimmunity: animal models. Immunol Res 2017;65 (01):55–65
- 179 Whitton JL, Fujinami RS. Viruses as triggers of autoimmunity: facts and fantasies. Curr Opin Microbiol 1999;2(04):392–397
- 180 Kanduc D. The comparative biochemistry of viruses and humans: an evolutionary path towards autoimmunity. Biol Chem 2019; 400(05):629–638
- 181 Kanduc D, Lucchese A, Mittelman A. Non-redundant peptidomes from DAPs: towards "the vaccine"? Autoimmun Rev 2007;6(05): 290–294
- 182 Kanduc D. Peptide cross-reactivity: the original sin of vaccines. Front Biosci (Schol Ed) 2012;4:1393–1401
- 183 Kanduc D, Shoenfeld Y. From HBV to HPV: designing vaccines for extensive and intensive vaccination campaigns worldwide. Autoimmun Rev 2016;15(11):1054–1061